Solvent effect and fluorescence response of the 7-tert-butylpyrene-dipicolyl amine linkage for the selective and sensitive response toward Zn(II) and Cd(II) ions

Zannatul Kowser,¹ Hirotsugu Tomiyasu,¹ Xuekai Jiang,¹ Ummey Rayhan, Carl Redshaw² and Takehiko Yamato*¹

The different binding behaviour of 7-tert-butylpyrene based chemosensors bearing dipicolylamine (Dpa) linkages at the 1,3-positions was investigated in various solvents for the sensing of Zn(II) and Cd(II). The potential mono-chelating ligand \(\text{L}_1\) follows the same binding pattern in both THF and methanol-water solvent systems, exhibiting high selectivity and sensitivity for Cd(II) than Zn(II) mainly in THF solvent system. The potential bis-chelate ligand \(\text{L}_2\) can selectively bind both Zn(II) and Cd(II) in a 1:1 ratio in THF, whereas in methanol-water (7:3) at pH = 7.0; a 1:2 binding ratio was observed. In THF, two sites of ligand \(\text{L}_2\) can only selectively and sensitively bind one Zn(II) or Cd(II). The different complexation behaviours of \(\text{L}_1\) and \(\text{L}_2\) in different solvents were studied by means of fluorescence spectra and \(^1\)H-NMR titration experiments in the presence of Zn(II) and Cd(II).

Introduction

The design and synthesis of molecular receptors for the detection of environmentally and biologically important species has attracted growing interest in recent years.¹ Amongst them, chemosensors whose fluorescence emission is sensitive to the environment and solvent media are especially important.²⁻⁵ In this regard, many fluorescence mechanisms have also been reported by probing sensing properties based on Photinduced Electron Transfer (PET), Intermolecular Charge Transfer (ICT), Chelation Enhanced Fluorescence (CHEF). Indeed, their application in the field of supramolecular chemistry has been elegantly illustrated.⁶ In case of PET,⁷ there is little or no change of the spectral shifts with changes of emission intensities, whereas both spectral shifts and intensity changes are observed for ICT, whilst CHEF also exhibited fluorescence enrichment with or without accompanying spectral changes.

The detection of Zn⁴⁺ is important both in vitro and in vivo due to its biological relevance.⁸⁻¹¹ It is an indispensable element for the human body and in many physiological and pathological processes, it performs an essential role.¹² It has been reported that its deficiency give rise to acrodermatitis enteropathica,¹³ but it is detrimental when present in excess, causing severe health problems such as superficial skin diseases, prostate cancer, diabetes and brain diseases. Unfortunately spectroscopically silent Zn⁴⁺ is difficult to detect directly.¹⁴ By contrast, a trace amount of Cd⁴⁺ is highly toxic towards the human body. Its intake causes serious diseases such as renal dysfunction, calcium metabolism disorders and prostate cancer.¹⁵

It is known that fluorescence quenching sometimes creates an unfavourable condition for a high signal output upon recognition of ions and also interferes with temporal separation of spectrally similar complexes with time-resolved fluorometry.¹⁶ Thus, our main focus is to design a chemosensor that does not quench the fluorescence upon binding with a metal ion. In this regard, the PET which is responsible for fluorescence quenching is minimized in the signaling moiety upon binding and results in the enhancement of the fluorescence.

Recently, pyrene has been utilized widely as a fluorophore to detect ion pairs, cations, anions¹⁷ and neutral species,¹⁸ because of the photoluminescence properties and chemical stabilities associated with pyrene. Given this, we have developed chemosensors that contain a 7-tert-butylpyrene as a fluorophore moiety and dipicolylamine as a receptor moiety connected through a C–N bond. Such an efficient and simple ligand system was also proposed by Ojida et al.¹⁹ They synthesized the binuclear anthracene complex Zn(II)-Dpa, and used it as an anion sensor for phosphorylated peptides. In our present work, we have established the ligands as efficient cation sensors which reveal different behaviour in different solvent systems.

The purpose of this work is to shed light on the mechanism of the different fluorescence response of receptor \(\text{L}_2\) with Zn⁴⁺.
and Cd²⁺ in various solvent systems. Interestingly, a 1:1 ligand to metal binding ratio was observed in case of THF for both Zn²⁺ and Cd²⁺ ions, whereas when using a methanol-water solvent system, it can selectively interact with Cd²⁺ and Zn²⁺ ions in a 1:2 (ligand/metal) stoichiometry. In case of methanol-water, L₂ exhibits a significant fluorescence enhancement for Zn²⁺, which is twice that observed for the THF solvent system. However, the potentially mononuclear receptor L₁ is highly selective in coordinating with Zn(II) and Cd(II).

Results and discussions

We have designed and successfully synthesized L₁ and L₂ using the reaction pathway shown in scheme 1. The fluorogenic molecule L₂ is synthesized from 7-tert-butylpyrene-1,3-dicarbaldehyde by treatment with 2,2ʹ-dipicolylamine, following which, the Schiff base is reduced by the gradual addition of NaBH(OAc)₃ to obtain L₂ in 82 % yield. Following the same reaction pathway, the potentially mono-chelate L₁ has also been prepared from 7-tert-butylpyrene-1-carbaldehyde in order to compare the binding affinities for Zn²⁺ and Cd²⁺ in different solvent systems. The characterization of these compounds was confirmed by ¹H and ¹³C NMR spectroscopy and by High-Mass spectrometry. In the absence of Zn²⁺ and Cd²⁺ ion, both L₁ and L₂ only afford weak fluorescence because of PET; the lone pair electrons from the amino group are transferred to the excited pyrenyl moiety and are presumed to quench the emission intensity of the pyrenyl fluorophore. After addition of Zn²⁺ and Cd²⁺ at small concentrations, preferential binding with dipicolylamine occurs to terminate the PET. In this way, the 7-tert-butylpyrene binuclear-Dpa complex exhibits a significant fluorescent enhancement for Zn²⁺ and can detect both Zn²⁺ and Cd²⁺ ions upon changing the solvent system. Addition of Zn²⁺ and Cd²⁺ ions using THF as solvent reveals a fluorescence at 402 nm. On the other hand, ligand L₂ can only detect Zn²⁺ ion with almost twice the fluorescence enhancement with on changing the solvent media, i.e., methanol-water instead of THF.

Firstly, the fluorescence properties of the receptor L₂ were investigated in different solvents (Fig. 1a) following addition of Zn²⁺. L₂ itself exhibits very weak fluorescence. It was then found that a large fluorescence enhancement (8-fold) was observed upon addition of...
To investigate the sensitivity of L2 toward Zn2+ ion, 9,10-bis[(2,2’-dipicolylamino)methyl]anthracene, L3 was also synthesized. As indicated in Fig. 1b, like L2, neither L3 nor L1 exhibit a distinct fluorescence emission after addition of Zn2+ (10 equiv) in methanol-water (10 mM HEPES/MeOH = 7:3, pH = 7.0). These observations suggest that in methanol-water, the ligand L2 was highly sensitive toward the Zn2+ ion. Fig. 2 shows the selective fluorescence enhancement after addition of Zn2+ and Cd2+ (100 µM) and I0 is fluorescence intensity for free receptor.

Fig. 1 (a) Fluorescence response of ligand L2 (7 µM) upon addition of Zn2+ in different solvent systems with excitation at 353 nm. (b) Fluorescence spectra of L1, L2 and L3 in CH3OH/H2O (10mM HEPES/CH3OH = 3:7, pH = 7.0) with excitation at 347 and 353 nm, respectively. By contrast, in methanol, THF or acetonitrile, the fluorescence intensity increase was monitored for similar ratios and was found to be almost half that observed in methanol-water. This suggests that the addition of water has a great impact on the fluorescence enhancement in the methanol-water system.

The changes of the fluorescence emission spectrum of L2 with Zn2+ using different ratios of methanol-water was also monitored, which suggested that a 7:3 ratio methanol-water solvent system was the ideal solvent media for the present work. In other words, on either increasing or decreasing the amount of water present, the emission intensity decreased (Figure SI 23).

To verify the fluorescence intensity changes in different solvents, fluorescence titration experiments and job’s plot were carried out. Figure 4 illustrates a gradual enhancement of fluorescence upon the addition of Zn2+ in L2 (7µM) was observed at 406 nm when excited at 353 nm. The change was almost terminated after addition of 2 equiv. of Zn2+ which suggested a 1:2 stoichiometry for the metal-ligand complex. This was again confirmed by the Job’s plot analysis. The fluorescence intensity exhibited a maximum at the mole fraction 0.65, suggestive of 1:2 complexation. The association constant for the complexation of L2 with Zn2+ was determined to be 3.3 × 10⁴ M⁻¹ (Fig SI 31). Figure 5a shows the fluorescence titrations of Zn2+ with L1 in THF. Stepwise addition of Zn2+ led to an increase of the fluorescence intensity until the complete addition of 1 equiv. of Zn2+. To confirm the binding sites of the sensor, the stoichiometries of L1 with Zn2+ were calculated using the Job’s plot, for which there was a maximum at 0.5 mole fraction, indicative of a 1:1 stoichiometry. Figure 5b presents the change of the fluorescence spectra of L2 upon addition of Zn2+ in THF. After addition of 1 equiv. of Zn2+, no obvious change occurred, which signified the 1:1 stoichiometry between L2 and Zn2+.
These results indicated that ligands L2 and L1 exhibit similar behaviour and binding toward Zn²⁺ and Cd²⁺ ions in THF. The US Environmental Protection Agency (EPA) set the maximum contaminant levels of Zn²⁺ and Cd²⁺ in drinking water at 7.6 and 4.5 \times 10^{-8} \text{M}, respectively.21 Given this, the receptors L1 and L2 can be considered to be highly selective for the detection of Zn²⁺ and Cd²⁺ (Table 1).

Figure 6a shows the selectivity among various metal ions. Probe L2 exhibited high selectivity toward Zn²⁺ over (Cu²⁺, Pb²⁺, Ag⁺, Hg⁺, K⁺, Li⁺ (as their perchlorate salts) and Co³⁺, Cr³⁺, Ni²⁺ including Cd²⁺ (as nitrate salts). Therefore, the affinity of L1 was observed with each of the respective metal cations and the results implied that L1 can selectively detect both Cd²⁺ and Zn²⁺ ions, but with a slightly higher affinity for Cd²⁺ versus Zn²⁺. Figure 6b reveals that L1 and L2 were more sensitive toward Cd²⁺ than Zn²⁺ when using THF as solvent. By contrast, the addition of other cations (Cu²⁺, Pb²⁺, Ag⁺, Hg⁺, K⁺, Li⁺, Co³⁺, Cr³⁺, Ni²⁺, Na⁺, Li⁺) showed almost no fluorescence enhancement. These results indicated that L1 and L2 exhibit selective emission enhancement toward Zn²⁺ and Cd²⁺ both in THF and methanol-water solvents. On the other hand, observations for the fluorescence emissions for the L2 (7 µM) and Zn²⁺ (100 µM) system, indicated that most of the competitive cations such as Pb²⁺, Ag⁺, Hg⁺, K⁺, Li⁺, Co³⁺, Cr³⁺, Ni²⁺, Cd²⁺ caused no obvious change at higher concentration (100µM) (figure SI 22). However, Cu²⁺, Ag⁺, Hg⁺ all strongly quenched the fluorescence in the L2+Zn²⁺ system. These results suggested that the co-ordination of Zn²⁺ with L2 is more selective than other metal ions, with the exception of Cu²⁺, Ag⁺ and Hg⁺.20, 21

<table>
<thead>
<tr>
<th>Compd.</th>
<th>Solvent</th>
<th>Binding model, L:M²⁺</th>
<th>( K_a (\text{M}^{-1}) )</th>
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<tbody>
<tr>
<td>L2:Zn²⁺</td>
<td>MeOH/H₂O</td>
<td>1:2</td>
<td>3.3 \times 10⁴</td>
</tr>
<tr>
<td>L2:Zn²⁺</td>
<td>THF</td>
<td>1:1</td>
<td>6.6 \times 10⁴</td>
</tr>
<tr>
<td>L1:Zn²⁺</td>
<td>THF</td>
<td>1:1</td>
<td>5.0 \times 10⁴</td>
</tr>
<tr>
<td>L2:Cd²⁺</td>
<td>THF</td>
<td>1:1</td>
<td>5.0 \times 10⁴</td>
</tr>
</tbody>
</table>

*Measured at 27 °C by fluorescence titration experiments (Figure SI. 31–34); host concentration was 7 µM.
Fig. 5
Fluorescence response of (a) ligand L1 (7 µM) (b) ligand L2 (7 µM) in addition with Zn²⁺ at 298 K. The excitation was performed at 347 nm for L1 and 353 nm for L2.

Fig. 6 Fluorescence intensity changes of ligand L1 and L2 (7 µM) in (a) CH₃OH/H₂O (10 mM HEPES/MeOH = 3:7, pH = 7.0) and (b) THF solvent at 298 K after addition of various metal ions (100 µM). I is the fluorescence intensity after addition of metal ions and I₀ is fluorescence intensity for free receptor.
**Fig. 7** Partial $^1$H-NMR titration of L2/guest (H/G = 1:2); (a) Free ligand L2 (1.5 × 10$^{-2}$ M); (b) $\text{L2} \supset \text{Cd}^{2+}$ (1 equiv.); (c) $\text{L2} \supset \text{Cd}^{2+}$ (2 equiv.) Solvent: (d) CD$_3$OD–D$_2$O (9:1, v/v, pD = 7.0). 300 MHz at 298 K.

**Fig. 8** Partial $^1$H-NMR titration of L2/guest (H/G = 1:1); (a) Free ligand L2 (0.5 × 10$^{-3}$ M); (b) $\text{L2} \supset \text{Cd}^{2+}$ (0.5 equiv.); (c) $\text{L2} \supset \text{Cd}^{2+}$ (1 equiv.); $\text{L2} \supset \text{Cd}^{2+}$ (2 equiv.). Solvent: THF-d$_8$. 400 MHz at 298 K.

**Table 2.** Chemical shift of dipicolylamine and methylene protons of free L2 and L2 with Zn$^{2+}$ or Cd$^{2+}$.

<table>
<thead>
<tr>
<th>Proton</th>
<th>Chemical Shift, $\delta$ ppm in MeOH- H$_2$O (H/G = 1:2)</th>
<th>Chemical Shift, $\delta$ ppm in THF (H/G = 1:1)</th>
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</thead>
<tbody>
<tr>
<td>H$_a$</td>
<td>3.85, 3.80, -0.05, 3.88, 0.03</td>
<td>3.87, 3.81, -0.06</td>
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<tr>
<td>H$_b$</td>
<td>4.28, 4.68, 0.40, 4.64, 0.36</td>
<td>4.39, 4.25, -0.14</td>
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<tr>
<td>H$_c$</td>
<td>7.42, 7.18, -0.06, 7.49, 0.07</td>
<td>7.48, 7.23, -0.25</td>
</tr>
<tr>
<td>H$_d$</td>
<td>8.35, 8.75, 0.40, 8.83, 0.48</td>
<td>8.46, 8.68, 0.22</td>
</tr>
</tbody>
</table>

$\Delta \delta$ values are the difference of the chemical shift between L2 and Zn$^{2+}$ or Cd$^{2+}$ at 27 °C. Here, minus sign (-) denotes a shift to higher magnetic field.

(pD = 7.0) was applied for these analysis. The $^1$H NMR signals reveal the aromatic and methylene regions of L2 (Fig. 7 and Figure S1 28). After addition of 2 equiv. of Cd$^{2+}$ and Zn$^{2+}$ the proton signals of L2, when in the presence of Zn$^{2+}$ ion, undergo a larger downfield shift than when the Cd$^{2+}$ ion was present. Moreover, two sets of four methylene H$_a$ protons were split into two peaks and broadened following binding with Cd$^{2+}$ and Zn$^{2+}$. The other proton signals overlapped with each other among the four pyridine rings of the two sets of Dpas as does of pyrene ring protons and leads to a downfield shift which is due to the decrease of electron density by the metal-nitrogen co-ordination. The H$_d$ protons of adjacent pyrene rings underwent a significant downfield shift ($\delta$ 8.35 to 8.75 and 8.83 ppm) for Cd$^{2+}$ and Zn$^{2+}$ ions, respectively. Furthermore, two sets of two methylene H$_b$ protons also broadened and underwent a large downfield shift. These results suggested that two sets of dpas were equally assigned for making a co-ordination bond with two metal ions and confirmed a 1:2 metal-ligand stoichiometry. The $^1$H NMR analysis also revealed larger chemical shift differences for L2 for the complexation with Zn$^{2+}$ versus the Cd$^{2+}$ ion.

In contrast, when using THF as solvent, there is no such change after addition of 1 equiv. of Cd$^{2+}$ ion which confirmed the 1:1 binding mode for the complexation of L2 with Cd$^{2+}$ (Fig. 8). Here, the same H$_a$ protons of the adjacent four pyridine rings undergo a smaller downfield shift (from $\delta$ 8.46 to 8.68 ppm) than in methanol-water solvent after addition of 1 equiv. of Cd$^{2+}$. Another three protons (H$_c$, H$_e$ and H$_f$) also experience a downfield shift. Moreover, two sets of four methylene H$_a$ protons are split into two broad peaks from $\delta$ 3.87 to 3.81 and 4.16 ppm following binding with Cd$^{2+}$ akin to the methanol-water system. On the other hand, the H$_b$ proton of the pyrene ring exhibits a large upfield shift from $\delta$ 8.35 to 7.64 ppm, and unlike the methanol-water system, the H$_b$ protons split into two peaks from $\delta$ 4.39 to 4.25 and 4.56 ppm, which suggested that the methylene H$_b$ and pyrene H$_b$ protons directly contribute to the binding with the metal ion. This phenomenon is only possible when the Cd$^{2+}$ is positioned at the centre between the two binding sites. However, in THF, addition of Zn$^{2+}$ induces vigorous precipitation which does not allow for analysis using $^1$H NMR spectra for elucidation of the binding mode. Moreover, the fluorescence spectra and Job’s plot confirmed the 1:1 binding mode of a L2$\supset$Zn$^{2+}$ complex.

The above NMR and fluorescence spectra together with the Job’s plot suggested that in methanol-water solvent system, two binding sites equally co-ordinate with two metal ions. On the other hand, Zn$^{2+}$ or Cd$^{2+}$ is positioned between two binding sites in THF. Given the shape of THF (a five membered ring), it can lead to a pronounced pseudorotational effect which is responsible for the stable twisted conformation. It is assumed that this structural property plays an important role in the 1:1 ligand to metal binding system.

**Conclusion**

In conclusion, the novel fluorogenic molecules L1 and L2 based on 7-tert-butylpyrene have been synthesized. The binding of Zn$^{2+}$ and Cd$^{2+}$ ions at the pyrene linked dipicolylamine moieties was investigated by using fluorescence and $^1$H NMR titration experiments. It was found that receptor L1 exhibits a similar binding toward Cd$^{2+}$ and Zn$^{2+}$ in both solvent systems. Herein, L1 displayed higher fluorescence sensitivity for Cd$^{2+}$ versus Zn$^{2+}$. On the other hand, receptor L2 exhibited different binding behaviour in different solvent systems. When the molecule was dissolved in methanol-water solvent system, it selectively detected Cd$^{2+}$ and Zn$^{2+}$ with a 1:2 (ligand:metal) binding ratio. It was noticeable that L2 had the strongest affinity for binding with Zn$^{2+}$ ion versus Cd$^{2+}$ and all the other competitive metal ions. In contrast, using THF as solvent, Zn$^{2+}$ or Cd$^{2+}$ is positioned between two binding sites and followed a 1:1 property plays an important role in the 1:1 ligand to metal binding system.
binding mode. It was concluded that ligands \( L_1 \) and \( L_2 \) exhibited similar binding behaviour in THF.

**Experimental Section**

**General**

**General**: Unless otherwise stated, all other reagents used were purchased from commercial sources and were used without further purification. Compounds \( 1,23^3,22^4,2^5 \) and receptor \( L_3^{10a} \) were prepared following the reported procedures. All the solvents were dried and distilled by the usual procedures before use. All melting points were determined using a Yanagimoto MP-S1. JEOL FT-300 NMR spectrometer and Varian-400MR-vnmrs400 with SiMe\(_4\) as an internal reference: \( J \)-values are given in Hz. UV-vis spectra were recorded using a Shimadzu UV-3150UV-vis-NIR spectrophotometer. Fluorescence spectroscopic studies of compounds in solution were performed in a semimicro fluorescence cell (Hellma®, 104F-QS, 10 × 4 mm, 1400 \( \mu \)L) with a Varian Cary Eclipse spectrophotometer. Mass spectra were obtained with a Nippon Denshi JMS-HX110A Ultrahigh Performance mass spectrometer at 75 eV by using a direct-inlet system.

**Synthesis of Compound 2**

To 7-tert-butylpyrylene (500 mg, 1.93 mmol), 1,1-dichloromethyl methyl ether (333 mg, 2.90 mmol) was added in \( \text{CH}_2\text{Cl}_2 \) (20 ml) at 0 °C with stirring for 10 min. A solution of \( \text{TiCl}_4 \) (0.53 ml, 4.8 mmol) was added drop wise to the stirred solution over 10 min. After this addition, the reaction mixture was continuously stirred for 18 h at 45 °C. Then, the reaction mixture was quenched with ice water and extracted with \( \text{CH}_2\text{Cl}_2 \) (2 × 100 mL). The organic layer was washed with water (2 × 200 mL), brine (2 × 200 mL), dried over \( \text{MgSO}_4 \) and then evaporated. The crude product was purified by column chromatography eluting with ethyl acetate-hexane (3:1) to afford a yellow solid (230 mg, 62 %). Mp: 134–135 °C; \( \text{\textsuperscript{1}H NMR} \) (300 MHz, CDCl\(_3\)): \( \delta \) = 1.58 (9H, s, tBu), 3.92 (4H, s, \( \text{CH}_2 \)), 4.39 (2H, s, \( \text{CH}_2 \)), 7.14–7.09 (2H, m, pyridine-H), 7.47 (2H, d, \( J = 7.8 \) Hz, pyridine-H), 7.60 (2H, d, \( J = 5.6 \) Hz, pyridine-H), 7.96 (2H, s, pyrene-H\(_{6,8}\)), 8.04 (1H, d, \( J = 9.3 \) Hz, pyrene-H\(_3\)), 8.07 (2H, s, pyrene-H\(_{6,8}\)), 8.19 (2H, dd, \( J = 1.7, 6.3 \) Hz, pyrene-H\(_{7,8}\)), 8.33 (1H, d, \( J = 9.2 \) Hz, pyrene-H\(_3\)) and 8.53 (2H, d, \( J = 4.9 \) Hz, pyridine-H) ppm. \( \text{\textsuperscript{13}C NMR} \) (100 MHz, CDCl\(_3\)): \( \delta \) = 31.9, 35.2, 57.1, 60.4, 122.0, 122.1, 122.3, 122.9, 123.3, 123.9, 124.3, 124.9, 127.2, 127.3, 127.9, 129.6, 130.5, 130.6, 131.1, 132.3, 136.4, 148.8, 148.9 and 159.6 ppm. HRMS: \( m/z \) calcd. for \( \text{C}_{46}\text{H}_{44}\text{N}_6 \) 681.3706; found 681.3707 [M\(^+\)].

**Synthesis of Receptor L1**

To a solution of 7-tert-butylpyrylene-1-carbaldehyde (225 mg, 0.79 mmol) in 1,2-dichloroethane (18 mL), 2,2'-dipropylamine (156 mg, 0.79 mmol) was added drop wise. Then the mixture was stirred for 18 h at 45°C. After that, sodium triacetoxoborohydride (500 mg, 2.36 mmol) was added, and the mixture was further stirred for 24 h at 50 °C. The reaction mixture was quenched with ice and extracted with \( \text{CH}_2\text{Cl}_2 \) (2 × 100 mL). The organic layer was washed with water (2 × 200 mL), brine (2 × 200 mL), dried over \( \text{MgSO}_4 \) and then evaporated. The crude product was purified by column chromatography eluting with ethyl acetate-hexane (3:1) to afford a yellow solid (230 mg, 62 %). Mp: 134–135 °C; \( \text{\textsuperscript{1}H NMR} \) (300 MHz, CDCl\(_3\)): \( \delta \) = 1.58 (9H, s, tBu), 3.92 (4H, s, \( \text{CH}_2 \)), 4.39 (2H, s, \( \text{CH}_2 \)), 7.14–7.09 (2H, m, pyridine-H), 7.47 (2H, d, \( J = 7.8 \) Hz, pyridine-H), 7.60 (2H, d, \( J = 5.6 \) Hz, pyridine-H), 7.96 (2H, s, pyrene-H\(_{6,8}\)), 8.04 (1H, d, \( J = 9.3 \) Hz, pyrene-H\(_3\)), 8.07 (2H, s, pyrene-H\(_{6,8}\)), 8.19 (2H, dd, \( J = 1.7, 6.3 \) Hz, pyrene-H\(_{7,8}\)), 8.33 (1H, d, \( J = 9.2 \) Hz, pyrene-H\(_3\)) and 8.53 (2H, d, \( J = 4.9 \) Hz, pyridine-H) ppm. \( \text{\textsuperscript{13}C NMR} \) (100 MHz, CDCl\(_3\)): \( \delta \) = 31.9, 35.2, 57.1, 60.4, 122.0, 122.1, 122.3, 122.9, 123.3, 123.9, 124.3, 124.9, 127.2, 127.3, 127.9, 129.6, 130.5, 130.6, 131.1, 132.3, 136.4, 148.8, 148.9 and 159.6 ppm. HRMS: \( m/z \) calcd. for \( \text{C}_{46}\text{H}_{44}\text{N}_6 \) 681.3706; found 681.3707 [M\(^+\)].

**Determination of the Association Constants**

The association constants were determined by using the fluorescent titration experiment of \( L_1 \) and \( L_2 \) in a constant concentration of host receptor (7 × 10\(^{-6}\) M) and varying the guest concentration (0–20 × 10\(^{-6}\) M). The association constant (\( K_a \)) for the complexes of receptor \( L_1 \) and \( L_2 \) were calculated by observing the integral intensities of the complex and of free host molecules using nonlinear curve–fitting analysis according to the literature procedure.

**\( ^{1}H \) NMR Titration Experiments**

A solution of \( \text{Zn(ClO}_4\text{)}_2 \cdot 6\text{H}_2\text{O} \) or \( \text{Cd(NO}_3\text{)}_2 \cdot 4\text{H}_2\text{O} \) in \( \text{D}_2\text{O} \) (1.5 × 10\(^{-2}\) M) was added to a \( \text{D}_2\text{O} \) (11:1, v/v) solution of receptor \( L_2 \) in the absence or presence of \( \text{Zn}^{2+} \) and \( \text{Cd}^{2+} \) ion in an NMR tube (300 MHz NMR). Similarly, a solution of \( \text{Zn(ClO}_4\text{)}_2 \cdot 6\text{H}_2\text{O} \) or \( \text{Cd(NO}_3\text{)}_2 \cdot 4\text{H}_2\text{O} \) in THF-d\(_8\) (0.5 × 10\(^{-3}\) M) was added to a THF-d\(_8\) solution of \( L_2 \) (400 MHz NMR). \( ^{1}H \) NMR spectra were recorded after addition of the reactants and the temperature of the NMR probe was kept constant at 27 °C.
Supporting information: Detailed fluorescence and $^1$H NMR titration data.

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Notes and references

† Electronic Supplementary Information (ESI) available: Details of the $^1$H/$^13$C NMR spectra, $^1$H NMR spectroscopic and UV-vis titration experimental data, the Bensei–Hilderbrand plot and Job’s plot, See DOI: 10.1039/b000000x/


