# **ARTICLE**

Cite this: DOI: 10.1039/x0xx00000x

Received 00th January 2021, Accepted 00th January 2021

DOI: 10.1039/x0xx00000x

www.rsc.org/

Allosteric binding properties of a 1,3-alternate thiacalix[4]arene-based receptor having phenyl-thiolurea and 2-pyridylmethyl moieties on opposite faces

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The synthesis of three heteroditopic receptors (5a-c) based on a thiacalix[4]arene in the 1,3-alternate conformation, which have two thiourea moieties linking various phenyl groups substituted with electron- withdrawing groups at their p-positions, with two 2-pyridylmethyl groups at the opposite side of the thiacalix[4]arene cavity, have been synthesized. One example (5a) has been characterized by X-ray crystallography. The binding properties of these receptors were investigated by means of <sup>1</sup>H NMR spectroscopy UV-vis absorption titration experiments CHCl<sub>3</sub>-DMSO-CD<sub>3</sub>CN (10:1:1, v/v) using various anions. The structures and complexation energies were also studied by density functional theory (DFT) methods. The results suggested that receptor 5c, which possesses two p-nitrophenyl thioureido moieties, can complex most efficiently in the thiourea cavity and exhibits high selectivity towards F<sup>-</sup> and AcO<sup>-</sup> ions. Interestingly, the formation of a heteroditopic dinuclear complex of receptor 5b, with F<sup>-</sup> and Ag<sup>+</sup> ions by a positive allosteric effect was observed.

# Introduction

Calix[n]arenes, which are macrocyclic compounds comprised of alternating numbers [n = 3-8] of phenolic groups typically linked via  $-CH_2-$  groups, have proven to be versatile and useful building blocks for a variety of applications, especially in host-guest chemistry. This is due to the fact that they are easily synthesized and can be modified with a wide range of functional groups which can be fine-tuned for a variety of applications. One of the most widely explored area of calix[n]arene chemistry has been in the development of sensitive and selective ionophoric receptors for cations, and also anions. Included in the class of calix[n]arenes are the thiacalix[4]arene analogues, in which four alternating phenolic

groups are linked via divalent sulfur atoms.<sup>2</sup> While not as extensively studied as the "classical" calix[4]arenes they nevertheless have shown similarities in host-guest chemistry as chemosensors for metal cations since they can also be relatively easily functionalized, not only at their narrow-rim phenolic oxygen atoms, but also the sulfur linkage can be synthetically modified.<sup>3</sup>

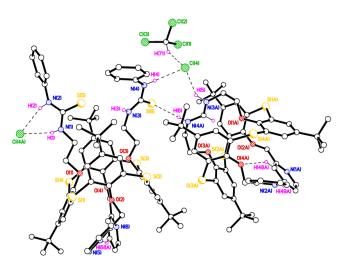
Several different types of artificial systems including calix[4]arenes have been shown to be suitable for allosteric effects in host-guest interactions with metal cations. Similar allosteric effects and regulation<sup>4</sup> in biological systems are of great importance in organic processes. Anions are also important for biological processes, involving DNA and as enzyme substrates. There is therefore much interest in developing anion selective sensors which,

however, can be more challenging. Besides halide anions which are spherical, anions can also be trigonal or Y-shaped such as *e.g.* acetate (AcO<sup>-</sup>) or benzoate ions; or tetrahedral, such as e.g. dihydrogenphosphate (H<sub>2</sub>PO<sub>4</sub><sup>-</sup>) and perchlorate ions to name just a few. Many colorimetric anion chemosensors<sup>7,8</sup> have been reported with chromogenic signaling moieties such as anthraquinone, benzenediimide and *p*-nitro-phenylazo groups which have been incorporated in a variety of structural scaffolds which contain urea groups. These have proven to be efficient naked-eye colorimetric detectors for various anions since hydrogen-bonding interactions can occur between anions and the urea NH protons. However, there are relatively few reports of colorimetric anion chemosensors based on calix[4]arene scaffolds. 9.10

Previously, we have reported the observation of allosteric effects resulting from the interaction of alkali metal cations and anions with thiacalix[4]arene derivatives. Herein, we report further studies on such allosteric effects being observed with tetra-*p-tert*-butyl thiacalix[4]arene, the latter is in a 1,3-alternate conformation and is di-substituted on one rim with thiourea moieties linked to various *p*-substituted phenyl groups with electron-withdrawing groups (5a-c). At the opposite face of the thiacalix[4]arene cavity there are two 2-pyridylmethyl groups. We herein demonstrate that this heteroditopic system undergoes complexation with both anions and Ag<sup>+</sup> at the opposite rims of the receptor molecule and with effective, positive allosteric effects.

protons, two singlets (4H each) for aromatic protons and two singlets (2H each) for the four urea NH protons (Figures S1–S12).

The molecular structure of **5a**, which was recrystallized from a mixture of CHCl<sub>3</sub>–CH<sub>3</sub>CN (1:1, v/v) in the presence of one equiv. of tetrabutylammonium chloride by slow evaporation, was also verified by a single-crystal X-ray analysis (Fig. 1 and Figures S13–S17). The



# Results and discussions

#### Synthesis

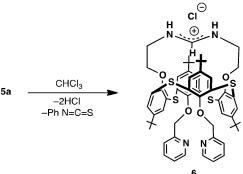
O-Alkylation of 1<sup>14</sup> with 2 mol equiv. of 2-bromoacetamide in the presence of Na<sub>2</sub>CO<sub>3</sub> using a reported procedure<sup>15</sup> afforded the 1,3-di-O-substitution product distal-2 in 64 % yield as the major product. Other possible isomers were not observed. The reaction of 2 with 2-(chloromethyl)pyridine in acetone in the presence of Cs<sub>2</sub>CO<sub>3</sub> formed 1,3-alternate-3 in 52 % yield. Reduction of 3 with BH<sub>3</sub> under THF reflux conditions afforded 1,3-alternate-4 in 82% yield. Condensation reactions of 4 with 2.2 equivalents of the appropriate thioisocyanate in CH<sub>2</sub>Cl<sub>2</sub> furnished the corresponding thiourea receptors 5a-c in good to excellent yields (Scheme 1). Each of the <sup>1</sup>H-NMR spectra of 5a-c in CDCl<sub>3</sub>-DMSO (10:1, v/v) exhibited the characteristics of 1,3-alternate canformations. The spectra showed two singlets (18H each) for the tert-butyl protons, one singlet (4H) for OCH<sub>2</sub>CO protons, two triplets (4H each) for the -OCH<sub>2</sub>CH<sub>2</sub>-

asymmetric unit comprises two different calixarenes with moieties  ${}^{\prime}C_{70}H_{78}N_6O_4S_6{}^{\prime}$  (5a) and  ${}^{\prime}C_{57}H_{67}N_4O_4S_4{}^{+\prime}$ , a Cl<sup>-</sup> counter ion and one molecule of CHCl<sub>3</sub> as the solvent of crystallization. The charge on the Cl<sup>-</sup> anion is stabilised by charge-assisted hydrogen bonding from a fairly rare [-NH-CH-NH-]+ functional group. This group has a delocalised positive charge on the second, modified, thiacalixarene molecule in the asymmetric unit (Fig. 2 & Figures S13-S14). From the N-C bond lengths, it is clear that individually they are neither single nor double bonds, but share the delocalised positive charge across both bonds. The asymmetry of the bond lengths is a result of H(5) interacting with the Cl<sup>-</sup> anion, thus lengthening the N(3A)–H(5) bond and, as a result, shortening the adjacent N-C bond (Fig. 2). This cationic moiety is demonstrating an ability to capture chloride ions. There are currently 17 structures in the CSD which exhibit the same functional group (e.g. XANFIC, RURWUY, MAJSEV), 16 in which both C-N bonds are roughly 1.30 Å, with any asymmetry in the two bond lengths being attributed to different substituents either side of this functional group.

**Fig. 1** X–ray crystal structure of receptor **5a** (left hand side). The asymmetric unit shows an intermolecular  $S\cdots H-N$  hydrogen bond between the two different thiacalixarenes and the  $N-H\cdots Cl^-$  interactions between each thiacalixarene and the  $Cl(4)^-$  anion; Cl(4A) is a symmetry equivalent. Minor disorder components and H atoms not involved in H-bonding are omitted for clarity.

Interestingly, it was found that one of the thiacalixarene molecules in the asymmetric unit of 5a, which is in a 1,3-alternate conformation, shows weaker intramolecular C-H···N hydrogen bonding between the opposing pyridine rings (Fig. S15). The second thiacalixarene in the asymmetric unit also shows intramolecular hydrogen between the opposing pyridine groups in addition to the charge-assisted N-H···Cl hydrogen bonding (Figures S16–S17). There is an intermolecular S···H–N hydrogen bond between the two thiacalixarenes (Fig. 1 and Figures S16–S17). The receptor **5a** has a three-dimentional cavity which is large enough to accommodate a metal cation between the opposing 2-pyridyl side arms. The exact mechanism for the formation of compound 6 is not clear from the available data. However, it is clear that under the crystallization conditions employed, the two thiourea moieties react with a molecule of CHCl<sub>3</sub>. The intramolecular condensation product is formed either via a step-wise or concerted elimination of two molecules of HCl and two thiourea moeites as two phenyl isothiocyanates.

Fig. 2 A zoomed-in view showing the bond lengths (Å) of the [-NH-CH-NH-]<sup>+</sup> moiety with it's delocalised positive charge.



Scheme 2 Formation of 6 from 5a.

# Binding studies

The binding properties of receptors 5a-c in the presence of F as its tetrabutylammonium (TBA) salt, in CDCl<sub>3</sub>–DMSO–CD<sub>3</sub>CN (10:1:1, v/v) solution, were investigated by means of 'H-NMR spectroscopic titration experiments. As shown in Fig. 3, for the complexation of F with 5b for example, the signals for the NH<sub>a</sub> protons (red) progressively shifted downfield by 5.99 ppm ( $\delta = 8.06$  to 14.05 ppm) until five equivalents of F were added. On the other hand, the signals for the NH<sub>b</sub> protons (blue) progressively shifted downfield by 3.62 ppm ( $\delta = 7.34$  to 11.02 ppm) until five equiv. of F were added (Figure S19). These results are strongly suggestive of F<sup>-</sup> recognition by the receptor **5b** via hydrogen-bonding interactions between F and the N-H protons. On the other hand, the methylene protons adjacent to the NH<sub>b</sub> moiety (green) are shifted slightly upfield. A similar <sup>1</sup>H-NMR spectroscopic titration experiment with receptor 5a in CDCl<sub>3</sub>-DMSO-CD<sub>3</sub>CN solution (Figure S18) showed that addition of F<sup>-</sup> also resulted in clear downfield shifts of the <sup>1</sup>H NMR signals of the NH<sub>a</sub> protons. Moreover, the addition of F<sup>-</sup> (1.0 equiv.) to solutions of receptor 5c in CHCl<sub>3</sub>-DMSO-CD<sub>3</sub>CN (10:1:1, v/v) during the titration experiments resulted in the disappearance of the signals for urea NH<sub>a</sub> and NH<sub>b</sub> protons. These results also indicate that strong interactions between these anions and the thiourea NH groups in the receptor **5c** occur and that the kinetics of these anion exchanges are on the NMR time scale (Fig. S20).

All of the results obtained from the <sup>1</sup>H-NMR spectroscopic titration experiments clearly suggest that anion recognition by the receptors is via hydrogen-bonding interactions between the anion and the NH protons. The binding or association constant  $(K_a)$  values for 5a and 5b and F<sup>-</sup> from the <sup>1</sup>H NMR experiments were calculated using a 1:1 global fit analysis of the chemical shifts of the  $NH_a$ protons  $(K_a = (2.65 \pm 0.19) \times 10^6 \,\mathrm{M}^{-1} \,\mathrm{for}\,\mathbf{5a}$  and  $K_a = (9.31 \pm 0.65) \times 10^6 \,\mathrm{M}^{-1} \,\mathrm{for}\,\mathbf{5a}$  $10^6 \,\mathrm{M}^{-1}$  for **5b**) (Figure S21). The  $K_a$  value for **5b**, which has the electron-withdrawing F atoms on the phenyl ureido moieties, is greater than that for 5a. These values indicate that the  $K_a$  values are influenced by the electron-withdrawing groups located at the p-position of the phenyl ureido moieties. The introduction of electron-withdrawing groups at the p-position of the phenyl ureido groups increases the acidity of the thiourea protons, and hence enhances the anion-binding ability through hydrogen-bonding interactions.

Fig. 3 Binding mode of receptor 5b upon addition of  $F^-$  and partial  $^1H$  NMR (300 MHz) titration spectra of 5b (4.0  $\times$  10  $^3$  M) in CDCl3–DMSO–CD3CN (10:1:1, v/v) upon addition of TBAF at 298 K.

Further complexation studies of **5a–c** with F<sup>-</sup>, Cl<sup>-</sup>, AcO<sup>-</sup> and  $H_2PO_4^-$  ions were carried out using UV–vis spectroscopic titration experiments in  $CH_2Cl_2$ . Receptor **5c** (2.5  $\mu$ M) exhibits a broad absorption band at 305 nm in its UV–vis absorption spectrum (Fig. 4).

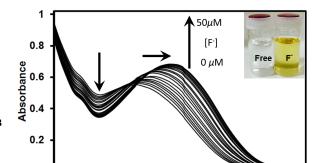


Fig. 4 UV–vis absorption spectra of receptor 5c (2.5  $\mu M) upon the addition of <math display="inline">Bu_4NF$  (0–50  $\mu M)$  in  $CH_2Cl_2$ –DMSO (10:1, v/v).

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**Table 1** Association constants  $(K_a [M^{-1}])^a$  of receptors **5a–c** with anions.

Table 1 Association constants (Ra [M ]) of receptors on a with amons.								
	$K_{\rm a} \left[ { m M}^{-1}  ight] \left( \pm { m K}_{ m error}{}^c  ight)$							
Host	F <sup>-</sup> (spherical)	Cl <sup>-</sup> (spherical)	AcO (spherical)	H <sub>2</sub> PO <sub>4</sub> <sup>-</sup> (spherical)				
5a	2.3×10 <sup>6</sup> (±1.6×10 <sup>5</sup> )	$\begin{array}{c} 2.0 \times 10^4 \\ (\pm 1.4 \times 10^3) \end{array}$	1.9×10 <sup>6</sup> (±1.3×10 <sup>5</sup> )	$4.6 \times 10^4  (\pm 3.3 \times 10^3)$				
5b	9.3×10 <sup>6</sup> (±1.6×10 <sup>5</sup> )	$4.4 \times 10^4$ (±3.0×10 <sup>3</sup> )	4.6×10 <sup>6</sup> (±3.3×10 <sup>5</sup> )	9.4×10 <sup>4</sup> (±6.6×10 <sup>3</sup> )				
5c	3.1×10 <sup>7</sup> (±2.2×10 <sup>6</sup> )	$2.5 \times 10^5$ (±1.7×10 <sup>3</sup> )	2.3×10 <sup>7</sup> (±1.6×10 <sup>6</sup> )	4.5×10 <sup>5</sup> (±1.7×10 <sup>3</sup> )				

<sup>a</sup> Measured in CH<sub>2</sub>Cl<sub>2</sub> at 298 K by UV-vis titration method; host concentration was 2.5 μM. <sup>b</sup> Guests used: TBA salts. <sup>c</sup> The ± error here is not a measure of precision of replicates but represents the %error resulting from the global-fit calculations since only single titration experiments in each case were conducted.

Upon addition of aliqots of  $F^-$  (0–50  $\mu$ M) to the CH<sub>2</sub>Cl<sub>2</sub>–DMSO solution of **5c** (Fig. 4 and Fig. S24), a gradual decrease in the absorption of the band at 305 nm with a simultaneous increase in the absorption at 360 nm and a clear isosbestic point at 335 nm can be seen. From a Job's plot, <sup>17c</sup> a 1:1 stoichiometry for the binding between the receptor **5c** and  $F^-$  (Figure S25) was established, and the  $K_a$  was determined to be  $3.1\times10^7$  M<sup>-1</sup> from the UV–vis titrations. <sup>17b</sup> The colour of the solution changed and was easily visible, from colourless to yellow upon addition of 5 equiv.  $F^-$ , indicating that a quinoidal structure was formed by the deprotonation of urea NH groups in the p-nitrophenyl ureido moiety.

The  $K_a$  values obtained from similar UV-vis titration experiments of 5a-c with F are summarized in Table 1. The order of the F complexation abilities is 5c > 5b > 5a, with the NO<sub>2</sub>-bearing receptor 5c showing the best recognition ability toward F-. Also, 5b has a larger  $K_a$  than 5a. The presence of electron-withdrawing groups at the p-position of the phenyl thioureido groups clearly increases the acidity of the thiourea protons, and hence enhances the ability through hydrogen-fluoride anion-binding interactions. Similarly, further UV-vis spectroscopic titration complexation studies of 5a-c with Cl<sup>-</sup> and AcO<sup>-</sup> and H<sub>2</sub>PO<sub>4</sub><sup>-</sup> were conducted and the  $K_a$  values are summarized in Table 1. The order of the anion complexation abilities of receptors **5a-c** is found to be: F  $> AcO^- > H_2PO_4^- > Cl^-$ . Receptor 5c therefore was capable of complexing with all of the anions tested and demonstrated the best recognition ability toward all of the anions tested, irrespective of their shapes, but was especially sensitive to F<sup>-</sup>.

A computational study was undertaken to further investigate the binding properties of receptors **5a–c** with the anions tested. The individual structures were fully geometry-optimized in the gas-phase using Gaussian 09<sup>18</sup> at the B3LYP level of DFT and the LANL2DZ basis set. Significant changes were observed for the distances between the two thiourea NH moieties in each of the receptors in their anion complexes. The conformational changes for example as shown for the 1:1 complex of **5b** with F<sup>-</sup>can be seen in Fig. 5 (more precise details for the computation studies for receptors **5a–5c** with the different anions are shown in Table S1, and Figures S28 and S30).

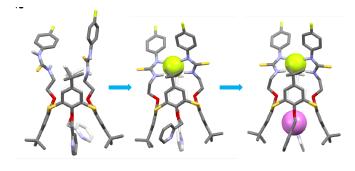


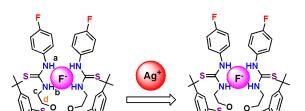
Fig. 5 Geometry-optimized (ball-and-stick) structures (in the gas phase) of: (a): Free ligand 5b; (b):  $5b\supset F^-$  and (c):  $Ag^+\subset [5b\supset F^-]$ . Colour code:  $F^-=$  green, nitrogen = blue, sulfur = yellow, oxygen = red, carbon = grey, hydrogen = white and  $Ag^+=$  violet.

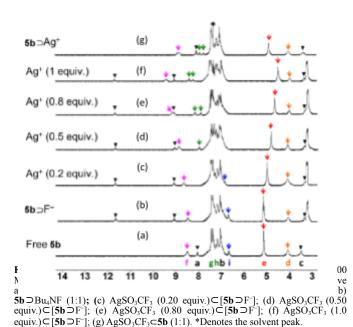
Fig. 5 shows the computed structures of (a) **5b** and (b) the 1:1 complex of **5b** with F<sup>-</sup>. The hydrogen-bonding distances between the F<sup>-</sup> ion and each of the thiourea NH protons (NH<sub>a</sub>···NH<sub>a'</sub> and NH<sub>b</sub>···NH<sub>b'</sub>) on the two *p*-fluorophenyl ureido moieties (Fig. 6) decrease from 8.783 to 2.530 (Å) and from 8.379 to 3.251 (Å), respectively. This also strongly supports the experimental evidence obtained for the formation of the 1:1 (**5b** $\supset$ F<sup>-</sup>) complex. The calculated interaction energies ( $\Delta$ *IE* kJ mol<sup>-1</sup>) for receptor **5b** with all anion complexes are shown in Table 2. The trend for the interaction energies for **5b** are in the order: F<sup>-</sup>> AcO<sup>-</sup>> H<sub>2</sub>PO<sub>4</sub><sup>-</sup>> Cl<sup>-</sup>, which is in agreement with the trend observed for the observed complexation data obtained from the UV-vis absorption titration experiments.

Table 2. Calculated interaction energies for receptors 5a-c with anions.

	_	Interaction energies ΔIE (kJ mol <sup>-1)</sup>			
Host	Ar	F-	Cl <sup>-</sup>	$AcO^-$	$\mathrm{H_2PO_4}^-$
5a	C <sub>6</sub> H <sub>5</sub> -	-481.33	-296.45	-294.52	-286.66
5b	$p$ -F–C $_6$ H $_4$ –	-501.95	-317.00	-314.05	-289.58
5c	$p\text{-}\mathrm{NO}_{2}\text{-}\mathrm{C}_{6}\mathrm{H}_{4}\text{-}$	-551.35	-362.33	-361.34	-339.47

<sup>1</sup>H-NMR titration experiments of **5b** with AgSO<sub>3</sub>CF<sub>3</sub> were conducted in CDCl<sub>3</sub>-DMSO (Fig. 6). Addition of a molar equiv. of AgSO<sub>3</sub>CF<sub>3</sub> causes an upfield shift ( $\Delta \delta = 0.25$  ppm) from  $\delta = 5.13$  to 4.88 ppm for the methylene  $-OCH_2$ Py protons of **5b**. The pyridine protons (H<sub>fi</sub>) all show downfield shifts, indicating that the Ag<sup>+</sup> is bound to the phenolic oxygens and the nitrogen atoms of the pyridine appendage (1:1  $Ag^+ \subset 5b$  complex in Fig. 6g). The chemical shift change ( $\Delta\delta$ ) of the H<sub>r</sub>-pyridyl protons adjacent to the nitrogen is shifted downfield by a larger amount ( $\Delta \delta = -0.39$  ppm, from  $\delta = 8.50$ to 8.89 ppm) than those of the other pyridyl protons. Spectral changes of the 1:1  $Ag^+ \subset 5b$  complex in the presence of an excess of AgSO<sub>3</sub>CF<sub>3</sub> were not detectable, which supports the exclusive formation of the 1:1  $Ag^+ \subset 5b$  complex. From these observations, it is clear that for the complexation of **5b** with Ag<sup>+</sup>, the pyridine nitrogens turn inward to bind with the Ag<sup>+</sup> within the cavity of the receptor, as is also shown in Fig. 5c. On the other hand, the methylene protons in  $\neg OCH_2$ Py shifted in the opposite direction due to the ring current effect of the benzene moieties. The Job's plot for the titration of **5b** with Ag<sup>+</sup> exhibited a 1:1 stoichiometry. From the <sup>1</sup>H-NMR spectroscopic titration experiments in CH<sub>2</sub>Cl<sub>2</sub>–DMSO (10:1, v/v), the  $K_a$  value for the complexation with  $Ag^+$  ion was determined to be  $K_a = 7.58 \pm 0.53 \times 10^2 \,\mathrm{M}^{-1}$ .





The different sites of complexation for the anions and Ag suggested the potential for an effective positive or negative allosteric effect between receptor 5b⊃F and Ag<sup>+</sup>. Thus, a <sup>1</sup>H-NMR spectroscopic titration experiment was conducted to determine this possibility. Figs. 6c-f show that when Ag+ ion was added to the solution of 5b⊃F-, the addition induces a downfield shift of the pyridine protons and upfield shifts of the -OCH<sub>2</sub>Py methylene protons, while the chemical shifts for the amido urea protons did not change. The addition of 1.0 equiv. of AgSO<sub>3</sub>CF<sub>3</sub> to **5b**⊃F<sup>-</sup> causes a larger upfield shift for the  $-OCH_2$ Py methylene protons ( $\Delta \delta = 0.79$ ppm,  $\delta = 5.13$  to 4.56 ppm) than that previously seen when a molar equivalent of Ag<sup>+</sup> was added to **5b** ( $\Delta \delta = 0.25$  ppm) itself. Moreover, a larger downfield shift was observed for the pyridyl  $H_f$  proton ( $\Delta \delta$  = 0.89 ppm,  $\delta = 8.50$  to 9.39 ppm) than that previously seen with **5b**. These observations show that a stronger binding ability occurs for the preformed  $5b \supset F^-$  complex with  $Ag^+$  than for 5b alone. This is supported by the fact that the  $K_a$  value for the complexation of **5b**⊃F<sup>-</sup> with Ag<sup>+</sup> was determined to be  $(4.49 \pm 0.31) \times 10^3 \text{ M}^{-1}$ , which is 6 times higher than that of the complexation of 5b with Ag<sup>+</sup>. These results suggest the formation of a heteroditopic dinuclear complex such as  $Ag^+ \subset [5b \supset F^-]$  as shown in Figs. 5c and 6, and that a positive allosteric effect of receptor **5b** towards Ag<sup>+</sup> in the presence of F occurs. Presumably this occurs as a result of the anion-electrostatic interactions with the N-H atoms of the thioureido pair resulting in a conformational change of the flexible thiacalix[4]arene allowing for the Ag+ complexation within the

The binding properties of receptor **5b** with  $Ag^+$  and fluoride anion were subjected to an additional computational study. The individual structures were fully geometry-optimized, in the gas-phase, using Gaussian  $09^{18}$  at the B3LYP level of DFT and the LANL2DZ basis set. The calculated interaction energies ( $\Delta IE \text{ kJ mol}^{-1}$ ) are shown in Table 3.

**Table 3.** The DFT interaction energies  $\Delta/E$  (kJ mol<sup>-1</sup>) and selected distances (in Å) for receptor **5b** and its complexes with F<sup>-</sup>, in the absence of and in the presence of Ag<sup>+</sup>.

	Δ <i>IE</i> (kJ mol <sup>-1</sup> )	Pyridine (N-N')	Thiourea (NH-N'H')	Thiourea (NH-N'H')-F	Pyridine (N-N')-Ag++
5b	-	6.437	6.806	-	-
<b>5b⊃</b> F⁻⁻	-501.95	-	3.050	1.742	6.512
Ag <sup>+</sup> ⊂5b	-436.59	3.677	4.076	-	2.386
Ag <sup>+</sup> ⊂[ <b>5b</b> ⊃F	-1090.36	3.708	3.710	1.734	2.417

The interaction energies for  $Ag^+ \subset \mathbf{5b}$ ,  $\mathbf{5b} \supset F^-$ , and  $Ag^+ \subset [\mathbf{5b} \supset F^-]$  are -501.95, -436.59, and 1090.36 kJ  $mol^{-1}$ , respectively, and are in agreement with the trend for the observed experimental complexation data obtained. The geometry-optimized structures (in the gas phase)  $\mathbf{5b} \supset F^-$  and  $Ag^+ \subset [\mathbf{5b} \supset F^-]$  are shown in Fig. 5.

#### Conclusion

In summary, a new family of heteroditopic receptors 5a-c based on a thiacalix[4] arene which is in the 1,3-alternate conformation has been synthesized and their anion complexation ability studied. These receptors have two thiourea moieties bearing various phenyl groups substituted with electron-withdrawing groups at their p-positions, as well as 2-pyridyl moieties at the opposite rims of the thiacalix[4]arene cavity. Using <sup>1</sup>H-NMR spectroscopic and UV-vis titration experiments, receptor 5c which has electron-withdrawing NO<sub>2</sub> groups at the p-positions of the phenylthioureido moieties was shown to have the most effective recognition ability towards the selected anions. The binding of Ag<sup>+</sup> at the 2-pyridyl moieties and the binding of the anions at the two thiourea NH groups of the two p-nitrophenylureido moieties, respectively, was investigated. The results indicated the likely complexation mode, and it was found that receptor 5c was able to bind all of the anions tested, irrespective of their shapes. Receptor 5c exhibited the high selectivity towards F amongst all of the anions tested and indicated that this receptor might be a promising candidate as a colorimetric chemosensor. The appearance of a positive allosteric effects with receptor 5b was also found using <sup>1</sup>H-NMR spectroscopic titration experiments. Interestingly, the formation of a heteroditopic dinuclear complex of receptor 5b with F and Ag ions by a positive allosteric effect could be observed.

# **Experimental Section**

# General

All melting points were determined with Yanagimoto MP-S1. 

<sup>1</sup>H-NMR spectra were determined at 300 MHz with a Nippon Denshi JEOL FT-300 NMR spectrometer with SiMe<sub>4</sub> as an internal reference; *J*-values are given in Hz. UV spectra were measured by a Shimadzu 240 spectrophotometer. Mass spectra were obtained on a Nippon Denshi JMS-01SG-2 mass spectrometer at an ionization energy of 70 eV using a direct inlet system through GLC. Elemental analyses were performed by Yanaco MT-5.

#### Materials

Unless otherwise stated, all reagents used were purchased from commercial sources and used without further purification. 5,11,17,23-Tetra-*tert*-butyl-2,8,14,20-tetrathiacalix[4]arene-25,26,27, 28-tetraol 1<sup>14</sup> and *distal-*5,11,17,23-tetra-*tert*-butyl-25,27-bis(carbamoylmethoxy)-26,2 8-dihydroxy-2,8,14,20-tetrathiacalix[4]arene 2<sup>15</sup> were prepared following the reported procedures.

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# Synthesis of *distal-5*,11,17,23-tetra-*tert*-butyl-25,27-bis-(carbamoylmethoxy)-26,28-dihydroxy-2,8,14,20-tetrathia-calix[4]arene 2

A mixture of 1 (1.0 g, 1.4 mmol) and Na<sub>2</sub>CO<sub>3</sub> (0.22 g, 1.5 mmol) in dry acetone (20 mL) was heated at reflux for 1 h under argon. Then 2-bromoacetamide (418 mg, 3.04 mmol) was added and the mixture was heated at reflux for an additional 48 h under argon. After cooling the reaction mixture to room temperature, the solvent was evaporated under reduced pressure. The residue was made alkaline with 30% NaOH (20 mL) and extracted with  $CH_2Cl_2$  (30 mL × 3). The combined extracts were washed with water (30 mL  $\times$  3) and brine (30 mL × 3). After washing, the organic layer was dried over magnesium sulfate and condensed under reduced pressure to give a crude product. Crystallization from CHCl<sub>3</sub>-MeOH (3:1, v/v) gave compound 2 (738 mg, 64 %) as colourless prisms. M.p. 148–151 °C. IR (KBr)/cm<sup>-1</sup> 3467 (NH), 3334 (OH), 3187 (NH) and 1694 (CO). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.05$  (s, 18H, tBu), 1.24 (s, 18H, tBu), 4.75 (s, 4H, OCH<sub>2</sub>CO), 6.19 (br, 2H, NH), 7.05 (br, 2H, NH), 7.43 (s, 4H, ArH), 7.67 (s, 4H, ArH) and 8.40 (s, 2H, OH) ppm. <sup>13</sup>C NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 30.9$ , 34.2, 68.8, 120.0, 120.7, 122.8, 123.3, 144.2, 164.8, 166.1 and 170.0 ppm. FABMS: m/z: 835.29  $(M^+)$ . Anal. calcd for  $C_{44}H_{54}N_2O_6S_4$  (835.17): C 63.28, H 6.52, N 3.35; found: C 63.02, H 6.49, N 3.33.

#### Synthesis of compound 3

To a solution of 2 (1.0 g, 1.2 mmol) and  $Cs_2CO_3$  (3.9 g, 12 mmol) in dry acetone (20 mL) was added 2-(chloromethyl)pyridine (2.0 g, 12 mmol) and the reaction mixture was heated at refluxed for 48 h under argon. After cooling the reaction mixture to room temperature, the solvent was evaporated under reduced pressure. The residue was made alkaline with 30% NaOH (20 mL) and extracted with CHCl<sub>3</sub> (30 mL  $\times$  3). The combined extracts were washed with water (30 mL  $\times$  3) and brine (30 mL  $\times$  3). After washing, the organic layer was dried over magnesium sulfate and condensed under reduced pressure to give a crude product. The residue was purified by column chromatography using CHCl<sub>3</sub> as eluent to provide a pale-yellow powder. Crystallization from CHCl<sub>3</sub>-hexane (3:1, v/v) gave compound 3 (634 mg, 52 %) as pale-yellow prisms. M.p. 195-197 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.83$  (18H, s, tBu), 1.28 (18H, s, tBu), 4.48 (4H, s, OCH<sub>2</sub>), 5.10 (2H, br, NH), 5.20 (4H, s, OCH<sub>2</sub>), 5.51 (2H, br, NH), 6.57 (2H, d, J = 8.5 Hz, Pyridine– $H_3$ ), 7.09 (4H, s, ArH), 7.00–7.42. (2H, m, Pyridine– $H_{4.5}$ ), 7.40 (4H, s, ArH) and 8.51 (2H, d, J = 7.7 Hz, Pyridine– $H_6$ ) ppm. <sup>13</sup>C NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 31.5, 31.9, 34.2, 34.3, 66.1, 70.7, 121.8, 122.0, 126.4,$ 126.8, 127.5, 127.8, 128.1, 135.9, 146.9, 147.0, 147.4 153.7, 155.3, 156.1 and 169.8 ppm. FABMS: m/z: 1017.40 (M<sup>+</sup>). Anal. calcd for C<sub>56</sub>H<sub>64</sub>N<sub>4</sub>O<sub>6</sub>S<sub>4</sub> (1017.39): C 66.11, H 6.34, N 5.51; found: C 66.23, H 6.29, N 5.55.

#### Synthesis of compound 4

A solution of BH<sub>3</sub>/THF (50 mL, large excess) was added to **3** (600 mg, 0.590 mmol) and the reaction mixture was heated at reflux for 20 h under argon. After cooling the reaction mixture to room temperature, it was quenched by the slow addition of aqueous 1.0 M HCl (30 mL). The mixture was again heated at reflux for 1 h under argon. After cooling the reaction mixture to room temperature, the solvent was evaporated under reduced pressure. The residue was made alkaline with 30% NaOH (20 mL) and extracted with CHCl<sub>3</sub>

(30 mL  $\times$  3). The combined extracts were washed with water (30 mL  $\times$  3) and brine (30 mL  $\times$  3). After washing, the organic layer was dried over magnesium sulfate and condensed under reduced pressure to give a crude product. The residue was purified by column chromatography using CHCl<sub>3</sub>-MeOH (10:1) as eluent to provide a colourless powder. Crystallization from CHCl<sub>3</sub>-hexane (7:3, v/v) gave compound 4 (479 mg, 82 %) as colourless prisms. M.p. 180–182 °C. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.82$  (18H, s, tBu), 1.31(18H, s, tBu), 2.20 (4H, br,  $NH_2$ ), 2.52 (4H, t, J = 9.0 Hz,  $OCH_2CH_2NH_2$ ), 4.02 (t, J = 9.0Hz, 4H,  $OCH_2CH_2NH_2$ ), 5.12 (s, 4H, Pyridine– $CH_2$ ), 6.50 (2H, d, J = 8.5 Hz, Pyridine– $H_3$ ), 7.05 (s, 4H, Ar-H), 7.13 (2H, br, Pyridine- $H_4$ ), 7.42 (s, 4H, Ar-H), 7.49-7.60. (2H, m, Pyridine– $H_5$ ) and 8.49 (2H, d, J = 7.7 Hz, Pyridine– $H_6$ ) ppm. <sup>13</sup>C NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 30.0, 33.8, 33.9, 68.9, 70.6,$ 120.0, 128.1, 128.5, 128.8, 129.4, 129.9, 136.0, 149.8, 150.5, 150.8, 154.0, 156.4 and 158.0 ppm. FABMS: m/z: 989.40 (M<sup>+</sup>). Anal. calcd for C<sub>56</sub>H<sub>68</sub>N<sub>4</sub>O<sub>4</sub>S<sub>4</sub> (989.43): C 67.89, H 6.93, N 5.66; found: C 67.45, H 6.50, N 5.45.

#### Synthesis of compound 5a

To compound 4 (100 mg, 0.101 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (15 mL) was added phenyl isothiocyanate (82 mg, 0.61 mmol) and the mixture was stirred at room temperature for 12 h under argon. The solvent was then evaporated under reduced pressure. The residue was purified by column chromatography using CHCl<sub>3</sub>-MeOH-aqueous 28% NH<sub>3</sub> solution (95:4:1, v/v) as eluent to provide a colourless powder. Crystallization from CHCl<sub>3</sub>-Hexane (3:1, v/v) gave 5a (94 mg, 74 %) as colourless prisms. M.p. 223–224 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>–DMSO, 10:1, v/v):  $\delta = 0.87$  (18H, s, tBu), 1.29 (18H, s, tBu), 3.52 (4H, br, OCH<sub>2</sub>CH<sub>2</sub>NH), 4.10 (4H, br, OCH<sub>2</sub>CH<sub>2</sub>NH), 5.12 (4H, s, Pyridine– $CH_2$ ), 6.70 (2H, d, J = 7.8 Hz, Pyridine– $H_3$ ), 7.03 (4H, s, Ar-H), 7.10-7.70. (m, 16H, Phenyl-H, CH<sub>2</sub>NH and Pyridine $-H_{4.5}$ ), 7.46 (4H, s, Ar–H), 7.72 (2H, s, Phenyl–NH) and 8.60 (2H, s, Pyridine–*H*<sub>6</sub>) ppm. <sup>13</sup>C NMR (400 MHz, CDCl<sub>3</sub>–DMSO, 10:1):  $\delta = 30.7$ , 34.0, 45.9, 64.8, 68.8, 122.3, 124.0, 126.2, 129.2, 130.1, 131.2, 132.4, 134.5, 136.0, 146.0, 146.3, 155.3, 156.0, 169.4 and 182.0 ppm. FABMS: m/z: 1259.46 (M<sup>+</sup>). Anal. calcd for C<sub>70</sub>H<sub>78</sub>N<sub>6</sub>O<sub>4</sub>S<sub>6</sub> (1259.80): C 66.74, H 6.24 N 6.67; found: C 66.69, H 6.32, N 6.76.

# Synthesis of compound 5b

To compound 4 (100 mg, 0.101 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (15 mL) was added 4-fluorophenyl isothiocyanate (93 mg, 0.61 mmol) and the mixture was stirred at room temperature for 12 h under argon. The solvent was then evaporated under reduced pressure. The residue was purified by column chromatography using CHCl<sub>3</sub>-MeOH-28% aqueous NH<sub>3</sub> solution (95:4:1) as eluent to provide a colourless powder. Recrystallization from CHCl3-Hexane (3:1, v/v) gave receptor **5b** (102 mg, 78 %) as colourless prisms. M.p. 209–210 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>–DMSO, 10:1, v/v):  $\delta = 0.82$  (18H, s, tBu), 1.12 (18H, s, tBu), 3.49 (4H, br, OCH<sub>2</sub>CH<sub>2</sub>NH), 4.10 (4H, br,  $OCH_2CH_2NH$ ), 5.13 (4H, s, Pyridine– $CH_2$ ), 6.68 (2H, d, J = 7.8 Hz, Pyridine- $H_3$ ), 7.04 (4H, s, Ar-H), 6.88-7.80. (m, 14H, p-F- $C_6H_4$ ,  $CH_2NH$  and Pyridine- $H_{4.5}$ ), 7.40 (4H, s, Ar-H), 8.06 (2H, br,  $p-F-C_6H_4-NH$ ) and 8.50 (2H, s, Pyridine- $H_6$ ) ppm. <sup>13</sup>C NMR (400) MHz, CDCl<sub>3</sub>–DMSO, 10:1):  $\delta$  = 31.0, 34.0, 44.8, 66.0, 69.9, 121.8, 122.6, 124.0, 124.3, 126.1, 126.5, 128.3, 131.9, 132.4, 133.0, 133.8, 135.2, 135.9, 146.8, 148.2, 155.1, 155.6, 156.2, 156.4, 160.0, 160.7 and 181.8 ppm. FABMS: m/z: 1295.5188 (M<sup>+</sup>).  $C_{70}H_{76}F_2N_6O_4S_6$ 

(1295.5386): calcd C 64.88, H 6.49, N 5.91; found: C 64.76, H 6.33, N 5.76.

#### Synthesis of compound 5c

To compound 4 (100 mg, 0.101 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (15 mL) was added 4-nitrophenyl isothiocyanate (109 mg, 0.606 mmol) and the mixture was stirred for at room temperature for 12 h under argon. After the reaction, the solvent was evaporated under reduced pressure. The residue was purified by column chromatography using CHCl<sub>3</sub>-MeOH- aqueous 28% NH<sub>3</sub> solution (95:4:1, v/v) as eluent to provide a colourless powder. Crystallization from CHCl3-CH3CN (3:2, v/v) gave receptor 5c (98 mg, 72 %) as pale-yellow prisms. M.p. 206–207 °C. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>–DMSO, 10:1):  $\delta$  = 0.82 (18H, s, tBu), 1.31 (18H, s, tBu), 3.50 (4H, br, OCH<sub>2</sub>CH<sub>2</sub>NH), 4.10 (4H, br, OCH<sub>2</sub>CH<sub>2</sub>NH), 5.19 (4H, s, Pyridine–CH<sub>2</sub>), 6.69 (2H, d, J = 7.8 Hz, Pyridine- $H_3$ ), 7.10 (4H, s, Ar-H), 7.20–7.68. (4H, m, Pyridine- $H_{4.5}$ ), 7.53 (4H, s, Ar-H), 7.80 (4H, d, J = 9.0 Hz,  $p-NO_2-C_6H_4$ ), 8.15 (4H, d, J = 9.0 Hz,  $p-NO_2-C_6H_4$ ), 8.50 (2H, s, Pyridine- $H_6$ ), 8.70 (2H, br,  $CH_2NH$ ), and 9.72 (2H, br, p-NO<sub>2</sub>-C<sub>6</sub>H<sub>4</sub>-NH) ppm. <sup>13</sup>C NMR (400 MHz, CDCl<sub>3</sub>-DMSO, 10:1):  $\delta = 30.2, 34.0, 44.1, 66.0, 71.3, 120.1, 122.0, 125.8, 127.0, 128.2,$ 129.6, 131.2, 132.4, 134.0, 134.9, 136.0, 138.0, 148.1, 151.0, 156.1, 157.7, 158.4, 164.0 and 183.9 ppm. FABMS: m/z: 1349.4189 (M<sup>+</sup>). C<sub>70</sub>H<sub>76</sub>N<sub>8</sub>O<sub>8</sub>S<sub>6</sub> (1349.5154): calcd C 62.29, H 5.68, N 8.30.

#### **Determination of the Association Constants**

The association constants were determined using  $^{1}$ H-NMR spectroscopic titration experiments at a constant concentration of host receptor (4.0 × 10<sup>-3</sup> M) and varying the guest concentration (0–8.0 × 10<sup>-3</sup> M). The  $^{1}$ H-NMR chemical shifts of the urea protons (NH) signal was used as a probe. The association constants ( $K_a$ ) for the complexes of receptors **5a–c** were calculated by nonlinear curve-fitting analysis of the observed chemical shifts of the NH protons according to the literature procedure.  $^{17}$ 

# <sup>1</sup>H NMR Titration Experiments

A solution of  $Bu_4NF$  in  $CD_3CN$  (4.0  $\times$  10<sup>-3</sup> M) was added to  $CDCl_3$  solutions of receptors **5a–c** in the absence of, or presence of  $AgSO_3CF_3$ . <sup>1</sup>H-NMR spectra were recorded after addition of the reactants and the temperature of the NMR probe was kept constant at 27 °C.

#### Crystallographic Analysis of Receptor 5a

Crystal data for 5a:  $C_{70}H_{78}N_6O_4S_6 \cdot C_{57}H_{67}N_4O_4S_4^+ \cdot Cl^- \cdot CHCl_3$ ,  $M_r$ =2415.05. Orthorhombic, Pbca; a = 26.9078 (10), b = 27.1717 (11), c= 34.4369 (13) Å; V = 25177.9 (17) Å<sup>3</sup>; Z = 8;  $D_x = 1.274$  Mg m<sup>-3</sup>; F(000) = 10208; T = 150 K;  $\mu = 0.40$  mm<sup>-1</sup>;  $\lambda = 0.7749$  Å, crystal size  $0.15 \times 0.15 \times 0.03$  mm<sup>3</sup>. Crystals were yellow blocks. Diffraction data were measured at ALS Station 11.3.1 using synchrotron radiation on a Bruker D8 with PHOTON 100 detector diffractometer equipped with a silicon 111 monchromator using thin-slice ω-scans. 19 218318 measured reflections, 23240 independent reflections ( $R_{\text{int}} = 0.055$ ) to  $\theta_{\text{max}} = 27.9^{\circ}$ ; 17514 reflections with I > $2\sigma(I)$ . The structure was determined by iterative, dual-space methods using the SHELXT program and refined by the full-matrix least-squares method, on F2, in SHELXL-2013/14.20-21 The non-hydrogen atoms were refined with anisotropic thermal parameters. Hydrogen atoms on C were included in idealized positions and their  $U_{\rm iso}$  values were set to ride on the  $U_{\rm eq}$  values of the parent atoms. H atoms on N were freely refined. At the conclusion of the refinement,  $wR_2 = 0.156$  (all data) and  $R_1 = 0.051$  (observed data), 1520 parameters,  $\Delta$ )<sub>max</sub> = 0.61 eÅ<sup>-3</sup>;  $\Delta$ )<sub>min</sub> = -0.88 eÅ<sup>-3</sup>; 134 restraints. The choroform molecule was modelled as disordered over two, closely-spaced sites with major site occupancy of 68.1(6) %. The crystallographic data (excluding structure factors) for the structure in this paper have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication number CCDC 2075593 for **5a**.

# Acknowledgements

This work was performed under the Cooperative Research Program of "Network Joint Research Center for Materials and Devices (Institute for Materials Chemistry and Engineering, Kyushu University)". We would like to thank the OTEC at Saga University for financial support. CR thanks the EPSRC for a travel grant (EP/R023816/1). The Advanced Light Source is supported by the Director, Office of Science, Office of Basic Energy Sciences, of the of U.S. Department Energy under Contract DE-AC02-05CH11231. SR and AA thank the Deanship of Scientific Research, King Saud University for financial support through the Vice Deanship of Scientific Research Chairs. Compute Canada is thanked for providing the computing resources.

#### Notes and references

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- † Electronic Supplementary Information (ESI) available: Details of the <sup>1</sup>H/<sup>13</sup>C NMR spectra, Crystallographic data, <sup>1</sup>H NMR spectroscopic and UV-vis titration experimental data, and Job's plot, See DOI: 10.1039/b000000x/
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