

1 **Validity and sensitivity of a human cranial finite element model:**  
2 **implications for comparative studies of biting performance.**

3 Running title: Cranial finite element model validation

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## 29 Abstract

30 Finite element analysis (FEA) is a modelling technique increasingly used in anatomical  
31 studies investigating skeletal form and function. In the case of the cranium this approach  
32 has been applied to both living and fossil taxa to (for example) investigate how form relates  
33 to function or infer diet or behaviour. However, FE models of complex musculoskeletal  
34 structures always rely on simplified representations because it is impossible to completely  
35 image and represent every detail of skeletal morphology, variations in material properties  
36 and the complexities of loading at all spatial and temporal scales. The effects of necessary  
37 simplifications merit investigation. To this end, this study focusses on one aspect, model  
38 geometry, which is particularly pertinent to fossil material where taphonomic processes  
39 often destroy the finer details of anatomy or in models built from clinical CTs where the  
40 resolution is limited and anatomical details are lost. We manipulated the details of a finite  
41 element (FE) model of an adult human male cranium and examined the impact on model  
42 performance. First, using digital speckle interferometry, we directly measured strains from  
43 the infraorbital region and frontal process of the maxilla of the physical cranium under  
44 simplified loading conditions, simulating incisor biting. These measured strains were then  
45 compared with predicted values from FE models with simplified geometries that included  
46 modifications to model resolution, and how cancellous bone and the thin bones of the  
47 circum-nasal and maxillary regions were represented. Distributions of regions of relatively  
48 high and low principal strains and principal strain vector magnitudes and directions,  
49 predicted by the most detailed FE model, are generally similar to those achieved *in vitro*.  
50 Representing cancellous bone as solid cortical bone lowers strain magnitudes substantially  
51 but the mode of deformation of the FE model is relatively constant. In contrast, omitting  
52 thin plates of bone in the circum-nasal region affects both mode and magnitude of  
53 deformation. Our findings provide a useful frame of reference with regard to the effects of  
54 simplifications on the performance of FE models of the cranium and call for caution in the  
55 interpretation and comparison of FEA results.

## 56 Keywords

57 Human cranium, finite element analysis, digital speckle interferometry, finite element  
58 model validation.

59

60

## 61 Introduction

62 Finite element analysis (FEA) is increasingly applied in studies of skeletal form and  
63 function. A focus of interest is the craniofacial skeleton where mechanical loading during  
64 ontogeny is important in ensuring balanced, normal growth and so, normal adult form and  
65 function (Lieberman 1996; Moss 2007; Menegaz et al. 2010). Further, comparative analyses  
66 of craniofacial strains predicted by FEA are potentially informative in relation to ecology  
67 and diet in both living and fossil taxa (Rayfield 2007; Kupczik et al. 2009; Strait et al. 2009;  
68 Wroe et al. 2010; Gröning et al. 2011b; Ross et al. 2011; O'Higgins et al. 2012; Smith et al.  
69 2015b). However, the results of an FEA depend on model geometry, material properties,  
70 applied loads and kinematic constraints. Full reproduction of these characteristics in a  
71 model of a structure like the human cranium is currently extremely difficult. Among model  
72 characteristics, detailed anatomy can be difficult to achieve because of limitations in  
73 imaging and thus reconstruction. Representation of anatomy is particularly error prone in  
74 the case of fossil material, because of taphonomic alteration of bone internal anatomy (e.g.  
75 due to sediment deposition) and tissue characteristics (e.g. similar image characteristics of  
76 fossilised bone and sediments) (Turner-Walker and Parry 1995; Olesiak et al. 2010; Fitton  
77 et al. 2015), or in the case of models built from clinical computed tomograms where image  
78 resolution is limited (Toro-Ibacache et al. 2015). Thus, simplification is inevitably necessary  
79 and it is important to assess the validity of FE models and, in particular, to understand how  
80 different modelling simplifications impact on performance.

81 Several studies have assessed FE model validity and sensitivity (Kupczik et al. 2007; Bright  
82 and Gröning 2011; Ross et al. 2011; Fitton et al. 2012; Cox et al. 2015; Fitton et al. 2015;  
83 Smith et al. 2015a). Collecting *in vivo* strain measurements for validation is impossible in  
84 many cases (e.g. because of ethical constraints and in fossils) and, where it is practicable,  
85 strain data are usually limited to a few point locations where the siting of strain gauges is  
86 feasible. More detailed and comprehensive measurement of surface strains is possible using  
87 post mortem material (Gröning et al. 2009) but replicating physiological loading *in vitro*  
88 then becomes an issue. In any case, the gathering of experimental data against which FE  
89 model performance can be assessed is time consuming, often destructive, subject specific,  
90 error prone and only possible in extant, not fossil specimens. A practical solution is to  
91 validate one or a limited number of FE models in detail and to base further models on  
92 what has been learnt from the validation and accompanying sensitivity analyses. The aim in  
93 this scenario is to validate the modelling approach and to understand the sensitivity of

94 models to variants of this approach, with the aims of increasing the accuracy of FE model  
95 behaviour and knowing more about the limits of interpretation imposed by simplifications.

96 Several prior studies of FE models of the skull have compared predicted strains with those  
97 measured *in vivo* (Strait et al. 2005; Ross et al. 2011), or with strains resulting from loading  
98 of wet cadaveric or dried skeletal material (Marinescu et al. 2005; Kupczik et al. 2007;  
99 Gröning et al. 2009; Smith et al. 2015a). To our knowledge, only one study to date has  
100 validated a model of a human cranium. This used 13 gauges to measure the strains over a  
101 cadaveric cranium that was loaded to perform a block-bite using half the dental arch  
102 (Szwedowski et al. 2011). The model was built using area-specific linearly elastic and  
103 isotropic material properties based on a map of bone density, as well as a hybrid solid-shell  
104 mesh, representing cancellous and cortical bone respectively. Sensitivity analyses were  
105 performed by varying the elasticity modulus, Poisson's ratio and homogeneous cortical  
106 shell thicknesses. The authors showed that the model with the most detailed cortical bone  
107 reconstruction and material properties correlated best with the experimental data, however  
108 the impact of different simplifications on strain contours and directions was not examined.

109 Among simplification approaches, it is common to omit structures that are very small and  
110 not feasible to reproduce accurately at the given model resolution. Such structures include  
111 fine plates of bone, cancellous bone, sutures and the periodontal ligaments (Kupczik et al.  
112 2007; Wood et al. 2011; Bright 2012). Thus, cancellous bone is often modelled as a bulk  
113 material because even relatively large trabeculae are not always distinguishable in computed  
114 tomograms (Gröning et al. 2012). Further, in FEA studies of the skull and postcranial  
115 skeleton, bone is often allocated simplified homogeneous and isotropic material properties  
116 obtained either from the literature or from average values of the specimen itself, rather  
117 than by mapping directly measured, heterogeneous orthotropic material properties (Strait et  
118 al. 2005; Kupczik et al. 2007) which are often unavailable and, particularly in the case of  
119 fossils and living humans, impossible to obtain.

120 Given the need for simplifications in modelling (including the extent to which cortical and  
121 cancellous bone are differentiated), the aim of the present study is to provide a frame of  
122 reference for the construction of models of the human cranium and those of our  
123 anatomically close primate and fossil relatives. Five voxel-based FE models of the same  
124 human cranium were built varying their model geometry (anatomical detail and  
125 composition). Two manipulations are applied, the first involves changes in anatomical  
126 detail that are inevitable when finite element (voxel) sizes vary according to the typical

127 limited range of resolution of primary CT data used in most studies to date, and the second  
128 by representing or omitting cancellous bone in the model. To assess the validity of the  
129 predictions of the FE models, strains were compared with those measured *in vitro*, in the  
130 actual specimen.

131 *In vitro* strains were measured using an optical technique; digital speckle pattern  
132 interferometry (DSPI; Yang and Etemeyer 2003; Yang et al. 2007) which provides a full-  
133 field surface measurement of microscopic deformation, from which the surface  
134 displacements and strains of an object under load can be calculated. This approach has  
135 previously been used to validate predicted stresses and strains from FE models of a human  
136 mandible (Gröning et al. 2009) and a pig cranium (Bright and Gröning 2011). It offers  
137 several advantages over strain gauges, most notably, DSPI measures strains over the entire  
138 field of view, while strain gauges measure them at distinct points.

139 Model sensitivity was assessed by comparing the FEA results among models. Additionally  
140 larger, global changes in size and shape of the skull under loading can be compared among  
141 model variants using Procrustes size and shape analysis, from geometric morphometrics  
142 (Milne and O'Higgins 2012; O'Higgins and Milne 2013). This approach has previously been  
143 used in conjunction with strain maps from FEA of skeletal structures (Milne and O'Higgins  
144 2012; Fitton et al. 2015). It provides additional insights into modes of global deformation  
145 that are useful when assessing the impact of subtle differences among FE models in  
146 sensitivity analyses (Gröning et al. 2011a; Fitton et al. 2012; Fitton et al. 2015).

147 The following null hypotheses (H0) were tested:

148 H01: There are no differences in distribution, magnitude and direction between the  
149 principal strains predicted by the different FE models, and between these and the principal  
150 strains measured *in vitro*.

151 H02: There are no differences in magnitudes and modes of global deformation among the  
152 different finite element models.

153 The testing of these hypotheses allows us to assess the magnitude and nature of any  
154 differences in performance among the models and between the models and the cadaveric  
155 cranium. This consideration leads to some important insights into sources of error and  
156 their impact on FEA studies of crania.

157

## 158 **Materials and methods**

### 159 **Anatomical data**

160 The cadaveric head of a 74 year old man from the repository of the Centre for Anatomical  
161 and Human Sciences (Hull York Medical School, HYMS, UK) was used in this study. The  
162 subject signed consent for experimental anatomical studies in life, when he donated his  
163 remains and ethical approval was obtained from the HYMS Ethics Committee. All  
164 experimental work was carried out in accordance with the Human Tissue Act (available at  
165 [www.hta.gov.uk](http://www.hta.gov.uk)) and HYMS protocols for the handling and storage of cadaveric material.

166 The cadaver had been embalmed two years prior to this study using a modified version of  
167 the University of Bristol embalming fluid formulation (1.4% formaldehyde and 70%  
168 ethanol, Vickers Laboratories Ltd., Pudsey, UK). The head was scanned using computed  
169 tomography (CT) at the York Teaching Hospital (York, UK) with a Siemens 16-channel  
170 multidetector CT scanner equipped with a STRATON tube (Siemens Somatom Sensation  
171 16, Siemens Healthcare, Erlangen, Germany) at 120 kV and 320 mA with an H60s edge  
172 enhancing kernel. Voxel size was 0.48 x 0.48 x 0.7 mm. Initial reconstruction of images was  
173 performed using a specialist system (Syngo Multimodality workplace, Siemens Healthcare,  
174 Erlangen, Germany) to ensure adequate field of view and image quality. The image stacks  
175 were then exported as DICOM files for detailed segmentation and reconstruction as  
176 described further below.

### 177 ***In vitro* strain measurement.**

178 The head was skeletonised by dissection, removing the soft tissues and the periosteum,  
179 taking precautions not to damage the bone surface. The cranium was placed on the  
180 platform of a universal material testing machine with a 1 kN load cell (Lloyd's EZ50,  
181 Ametek-Lloyd Instruments Inc., Sussex, UK). The position and loading of the cranium was  
182 chosen as an easily replicable loading scenario; while the loading was not physiological the  
183 loading at the teeth was comparable to the way a tooth is loaded during biting. Steel blocks  
184 were used to support the cranium at both mastoid processes and the left central incisor.  
185 Compressive vertical forces were applied to the midplane of the frontal squama, 13 mm  
186 anterior to bregma (see experimental setup in Fig. 1a). The load was applied in 11 steps of  
187 50 N to achieve a final load of 550 N. The final arrangement of steel supports and load was  
188 arrived at by trial and error, with earlier runs of the loading experiment failing due to  
189 instability that was corrected by increasing friction between the steel blocks and platform

190 using emery paper. Stability of the cranium after each step was assessed by repeatedly  
191 checking that increases in the reaction force at the constrained border of the left central  
192 incisor scaled linearly with increasing loads. Five successive and successful experimental  
193 rounds (i.e. with stability of the set up and replicable recording of strains and reaction  
194 force) for *in vitro* strain measurement in the infraorbital region and four for the frontal  
195 process of the maxilla were achieved. The position of the loading point on the cranium was  
196 marked to control the position of the load between loading experiments. Incisor reaction  
197 forces were measured using a strain meter equipped with a 5 kN load cell (Omega DP25B-  
198 S, Omega Engineering Inc., Stamford, USA) previously calibrated by applying known  
199 compressive loads with the Lloyd's testing machine described above.

200 Full-field surface strains were measured using a Q-100 DSPI system (DANTEC Dynamics  
201 GmbH, Ulm, Germany). The regions selected for strain measurement in this study were  
202 the left infraorbital area and the frontal process of the maxilla, since both show high strains  
203 in FEAs of simulated incisor bites in primates (Gross et al. 2001; Kupczik et al. 2009;  
204 Fitton et al. 2012). This system provides a maximum field of view (FOV) of 25 x 33 mm<sup>2</sup>.  
205 The measured surfaces were covered with a thin layer of white spray (DIFFU-THERM  
206 developer BAB-BCB, Technische Chemie KG, Herten, Germany) to prevent surface  
207 reflection of ambient light. The Q-100 sensor was glued using its three legs to the  
208 boundaries of the treated surface using an acrylic-based adhesive (X60, HBM Inc.,  
209 Darmstadt, Germany). Sensor attachment to the surface is standard procedure in using the  
210 Q-100 system for safety critical engineering work. While there is a theoretical impact on  
211 measured strains, in practice any effect is restricted to close to the points of attachment  
212 which were not included in the analyses. This procedure was undertaken once for each  
213 surface, thus avoiding variations in the location of the measured surface between loading  
214 runs. Surface characterisation, phase calculation and deformation estimation (see steps in  
215 Fig. 1b) were carried out using the Istra Q-100 (v.2.7, DANTEC Dynamics GmbH, Ulm,  
216 Germany). The primary strain data produced by the Q-100 system, maximum ( $\epsilon_1$ ) and  
217 minimum ( $\epsilon_3$ ) principal strain magnitudes, plus 2D and 3D colour-coded strain contour  
218 plots (representing strain distributions, i.e. relative locations of high and low strain) were  
219 exported and used for comparison of FEA results.

## 220 FE model construction

221 The cranium was reconstructed from the CT images through a combined approach of  
222 thresholding and manual segmentation of bone and teeth using the visualisation program

223 Avizo (v.7.0.1, Visualization Sciences Group, Burlington, USA). Five different models were  
224 built (Table 1). To assess the impact of simplifying cancellous bone representation, in one  
225 model (model 1) cancellous bone was omitted, and hence all bone was modelled as a solid  
226 material with the Young's modulus of cortical bone. This approach has been used in  
227 previous studies of cranial FE models (Wroe et al. 2010; Bright and Gröning 2011; Fitton  
228 et al. 2012; Jansen van Rensburg et al. 2012; Toro-Ibacache et al. 2015) and is particularly  
229 relevant in cases where, because of model resolution, fossilisation and taphonomic  
230 processes, or in order to generate hypothetical model geometries via surface warping,  
231 modelling cancellous bone is impractical (Bright and Gröning 2011; O'Higgins et al. 2011;  
232 Fitton et al. 2015). The remaining models (models 2-5) have a cortical shell with cancellous  
233 bone defined as a bulk material of much lower modulus than cortical bone, approach also  
234 used in previous studies (Kupczik et al. 2009; Smith et al. 2015a). In these four remaining  
235 models, cancellous bone was represented as a bulk material in the regions normally strained  
236 during FE biting simulations, below the level of the fronto-zygomatic suture, including the  
237 anterior and middle portions of the cranial base.

238 The inner walls of the frontal, ethmoidal, sphenoidal and maxillary sinuses are often  
239 thinner than a single voxel and so are prone to being incompletely and poorly represented  
240 in the CT. In consequence, the question arose as to how best to represent them in an FE  
241 model. To assess the impact of omitting or including them in the model their anatomies  
242 were either fully reconstructed manually, albeit using one or two voxels to represent their  
243 thickness, or left as assigned by grey level thresholding, resulting in thin plates of bone with  
244 irregular holes. Model resolution was varied via resampling by using two different voxel  
245 sizes (0.48 mm and 0.35 mm) to simulate the effect of typical differences in resolution in  
246 CT scans used in previous FE studies of crania. Reducing voxel size achieves a more  
247 accurate representation of the thin inner nasal walls compared to using the larger voxel  
248 size. It is of interest to assess the effect of such differences between corresponding models  
249 (models 2 vs. 4 and 3 vs. 5). We were unable to carry out a more detailed convergence  
250 analysis comparing a range of mesh resolutions because of limitations of resolution of the  
251 clinical CT scanner in relation to the finest details of bony anatomy.

252 Anatomical details were refined manually in each model where needed, thus varying the  
253 total number of voxels and so, elements among models. In all cases, teeth were modelled as  
254 one material with a higher elastic modulus ( $E$ ) than bone. The characteristics of each model  
255 are detailed in Table 1 and their features are depicted in Fig. 2a. Subsequently, data were

256 exported as BMP stacks and converted into FE meshes of eight-noded linear cubic  
257 elements by direct voxel conversion. Model pre- and post-processing were performed using  
258 the custom FEA program VOX-FE (Fagan et al. 2007; Liu et al. 2012).

259 In all models cortical bone, cancellous bone and teeth were allocated homogeneous linearly  
260 elastic and isotropic material properties (with Poisson's ratio=0.3), following the approach  
261 used in previously validated models of human and macaque crania (Kupczik et al. 2007;  
262 Szwedowski et al. 2011) and the human mandible (Gröning et al. 2009). In models 2-5,  
263 cancellous bone was represented as a different material and was allocated an  $E$  of 56 MPa  
264 (Misch et al. 1999) and an  $E$  of 50 GPa was assigned to teeth, this being approximately the  
265 mean of the large range of values found in the literature for enamel and dentine (Meredith  
266 et al. 1996; Barak et al. 2009; Benazzi et al. 2012). The material properties of cortical bone  
267 are particularly important in relation to overall model stiffness (Marinescu et al. 2005; Strait  
268 et al. 2005) and these vary throughout the cranium. For this reason material properties of  
269 the cadaveric cranium were measured directly from two different regions before settling on  
270 a suitable uniform value. A bone sample was collected from the maxillary tuberosity and  
271 from the zygomatic arch.  $E$  was measured using a nano-hardness tester with a Berkovitch  
272 diamond indenter (CSM Instruments SA, Peseux, Switzerland) following the protocol in  
273 Kupczik et al. (2007). The average value was found to be  $16.3 \pm 3.7$  GPa for the tuberosity  
274 and  $21.9 \pm 2.7$  GPa for the zygomatic arch. Since these values lie within the range used in  
275 the literature for models of the human cranium (Horgan and Gilchrist 2003; Wroe et al.  
276 2010; Jansen van Rensburg et al. 2012), a single  $E$  of 17 GPa, which has been used in  
277 previous models (Kupczik et al. 2009; Gröning et al. 2011b; Fitton et al. 2012), was  
278 assigned to all cortical bone.

279 The points of applied vertical load, the biting point and mastoid support were replicated in  
280 the model. The predicted bite force in model 5 was used to check the loading condition by  
281 confirming that this matched the reaction force measured *in vitro* at the left upper incisor.  
282 Based on the experimental setup and to simulate loading conditions (i.e. vertically loaded  
283 incisor and immobilised mastoids), a vertical kinematic constraint was applied to the tooth,  
284 and constraints in all three-axes at each mastoid process. Loads and constraints were  
285 applied to the model in the form of selected nodes in the border of the incisor, and  
286 punctiform regions of nodes at the point of load application and tips of the mastoid  
287 processes.

**288 Measured vs. predicted strains**

289 The procedure to compare strains measured *in vitro* and those predicted by the FE models  
290 comprised three steps: (1) matching the FOV of the DSPI with the area of interest of the  
291 FE model, (2) data extraction and (3) data comparison.

292 To compare visually strain contours (representing strain distribution) similar colours were  
293 mapped to equivalent strain ranges from DSPI and FEA. The surface geometry of the  
294 region of the face measured by DSPI was exported as a Virtual Reality Modeling Language  
295 (VRML) file and visualised in 3D using Avizo. The surface of the cranium extracted from  
296 the CT was loaded into the same scene as the DSPI surface. The DSPI surface was then  
297 manually positioned to obtain the best fit with the cranium surface guided by anatomical  
298 structures and high magnification photographs of the skull surface. Best-fit was assessed by  
299 two observers (VT-I and PO). Coordinates marking the location of the DSPI surface on  
300 the CT-derived cranial surface were saved using Avizo in order to match the positions of  
301 sampling points among models.

302 The strain magnitude outputs from DSPI and FEA are not the same in both dimensionality  
303 (2D for DSPI and 3D for FEA) and resolution, making one-to-one comparison  
304 impossible. We therefore used an approach that compares profiles of strain magnitudes  
305 along corresponding lines traced over the surfaces of the specimen and model. The DSPI  
306 computes strain magnitudes over a regular 2D grid in the plane of the lens. Two straight  
307 lines in this plane (lines 1 and 2) were traced across the infraorbital and two across the  
308 frontal process fields of view (FOV; lines 3 and 4) using the vertices of the FOVs to  
309 optimise replicability of measurement. Line correspondence between the models and the  
310 DSPI surfaces is shown in Figs. 3a and 3b. Strain magnitudes at each point along the lines  
311 from DSPI were extracted and smoothed by once-averaging of single adjacent points on  
312 either side to reduce noise. To extract corresponding data from the 3D surface of the FE  
313 model, lines of landmarks were traced on the model surface forming equivalent straight  
314 lines to those used to extract strain magnitudes from the DSPI FOVs. Lines comprising 37  
315 (line 1), 30 (line 2), 28 (line 3) and 33 (line 4) landmarks were traced over the model in  
316 Avizo. These lines replicate those traced on the DSPI FOVs but they inscribe curves over  
317 the surface of the FE model. These curves have two dimensions, distance and depth, while  
318 DSPI traced lines have just one dimension, distance. The depth dimension was removed  
319 from each FE model curve by projecting it onto the plane described by its first two  
320 principal components. The first principal component, which represented distance rather

321 than depth, was then rotated into the plane of the DSPI FOV to achieve best fit. The strain  
322 values were smoothed in VOX-FE by once-averaging of neighbouring voxels in order to  
323 reduce strain fluctuations due to voxelation (Liu et al. 2012). After smoothing, predicted  
324 strain magnitudes at each of the landmarks were extracted for comparison against strains  
325 measured *in vitro*. The impact of simplifications of the model on relative (rather than  
326 absolute) strain magnitudes was assessed by calculating the correlation coefficient among  
327 models.

328 Both systems output surface strain magnitudes and vectors, the Istra Q-100 (DSPI) in 2D  
329 and VOX-FE in 3D. These software tools show vectors differently; with directions and  
330 magnitudes being represented in the VOX-FE output and directions alone in the Istra Q-  
331 100 outputs. Further, the densities and spacings of plotted vectors differ between the  
332 visualisations. Thus, to avoid crowding, in the visualisations from VOX-FE lines  
333 representing strain vectors were drawn at every fourth node in models 1, 2 and 3 and at  
334 every eighth node in the larger models, 4 and 5, over the areas of interest.

### 335 **Global model deformation**

336 It is important to note that there are two different definitions of the term 'deformation'. In  
337 material science and in the context of morphometrics, 'deformation' refers to changes in  
338 size and shape (local or global). This is the definition followed here since it reflects the  
339 quantities measured by strains, i.e. how the finite elements deform under load. This differs  
340 from the definition of 'deformation' used occasionally in mechanics (see Truesdell and Noll  
341 2004, p.48) where it may refer to the displacement of nodes of the FE model between  
342 unloaded and loaded states.

343 Global model deformations (changes in size and shape) resulting from applied loads were  
344 compared between FE models through Procrustes size and shape analyses based on 51  
345 craniofacial landmarks (described in Table 1, Supporting information) and visualised in Fig.  
346 3c). During size and shape analysis, coordinates are rotated and translated, thus preserving  
347 the changes in model size as well as shape due to loading. The resulting size and shape  
348 coordinates are then submitted to principal components analysis (PCA; O'Higgins et al.,  
349 2012; Fitton et al., 2015). Visualisations of predicted changes in cranial size and shape due  
350 to loading and the differences in modes of deformation among models used the surface  
351 corresponding to model 1, warped to the mean unloaded landmark configuration before  
352 further warping to represent model deformations. Two Cartesian transformation grids were  
353 drawn over the mean landmark configuration, and warped with the surface to facilitate

354 interpretation of visualised deformations (Fitton et al. 2012; O'Higgins et al. 2012). Since  
355 landmarks are placed only once on the CT-derived surface representing all the models,  
356 there is no measurement error associated to the method.

357

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## 358 Results

359 The experimental setup was replicated in VOX-FE for each of the models 1-5. The  
360 locations of each constrained point and applied load, plus the predicted vs. actual bite force  
361 measured *in vitro* were used to achieve accurate model and load orientation. The  
362 experimentally measured bite force in the most anatomically accurate model, 5, was  
363  $176.84 \pm 9.44$  N and the predicted bite force was 177.11 N. Repeating this setup, model 4  
364 predicted 177.21 N of bite force, whereas low-resolution models 1, 2 and 3 predicted 182,  
365 182.54 and 182.55 N of bite force respectively.

366 The results of the strain and global model deformation analyses are presented below.

### 367 Measured vs. predicted strains

368 In general, the strain contour plots predicted by the FEAs differ among models in  
369 magnitude but show similar distributions of regions of relatively high and low strain (Figs.  
370 2b, and 2c with adjusted strain ranges to improve visualisation). This is also evident from  
371 the plots of strain magnitudes (Figs. 4 and 5) where strains from the FE simulations are  
372 compared with the *in vitro* ranges. The match is better for lines 1 and 2 than for lines 3 and  
373 4. By comparing models 1, 2 and 4 with model 5, it appears that the main effect of  
374 representing regions of cancellous bone as solid cortical bone and reconstructing sinus and  
375 nasal walls was to increase model stiffness. Comparing FE models with each other and  
376 with the results from DSPI, the 'solid' model 1 shows strains three to four times lower than  
377 the *in vitro* results and the strains predicted for the other models (Figs. 4 and 5). Overall,  
378 models 2 to 5 showed similar strain magnitudes. However, models 2 and 4 (with  
379 incompletely reconstructed sinus and nasal walls) show the largest discrepancy with the  
380 values measured *in vitro* (particularly  $\epsilon_3$  values; Fig. 5) and the lowest correlations (Table 2)  
381 with model 5 of strains traced along the lines drawn over the frontal process of the maxilla  
382 (see Figs. 3a and 3b). Model resolution (comparing models 2 vs. 4 and 3 vs. 5) over the  
383 limited range assessed in this study does not have an effect on strain magnitude.

384 There are some differences in strain magnitudes between models and the experimentally  
385 measured strains, and between models 1, 2 and 4 compared to model 5 (the most accurate).  
386 However, the directions of the principal strain vectors are very consistent among models.  
387 These mainly consist of vertical compression and transverse tension of the nasal notch  
388 (Fig. 6) and of the infero-medial margin of the orbital opening in the frontal process of the

389 maxilla (Fig. 7). This is evident despite the differences described earlier in the ways strain  
390 vectors are displayed in the DSPI and VOX-FE outputs.

### 391 **Global model deformation**

392 The PCA of size and shape variables confirms and clarifies the findings from the analyses  
393 of strains with regard to differences and similarities in modes of deformation. In the plots  
394 of principal components (PCs), model deformations are represented by lines connecting  
395 the loaded and unloaded models (Fig. 8). Global deformations generally consist of dorso-  
396 ventral bending of the maxilla mainly at the level of the nasal notch. The deformations of  
397 models 1, 3 and 5 are virtually the same in direction (mode of deformation), varying only in  
398 magnitude with model 1 deforming less. Models 2 and 4 deform to greater degree and in  
399 subtly different ways from the others, with more vertical compression of the nasal aperture  
400 and lateral displacement of the mid to upper parts of the nasal margins. They also deform  
401 more asymmetrically than the other models. The magnitudes of model deformation due to  
402 loading are very small. As such, to aid visualisation the warpings in Fig. 8 were magnified  
403 250 times.

404

### 405 **Discussion**

406 The aim of the present study was to validate the performance of FE models of a human  
407 cranium and to assess their sensitivity to variations in anatomical detail and, secondarily, in  
408 model resolution. This is important because finite element models of crania are increasingly  
409 used to assess and compare function.

410 For this, a wet cadaveric human cranium was loaded experimentally, simulating a bite at the  
411 left upper incisor and the resulting strains and reaction force at the incisor were measured.  
412 These were then compared to the strains predicted by FE models built using two different  
413 simplification approaches: presence or absence of cancellous bone and inner sinus and  
414 nasal walls, and high or low resolution. It was hypothesised that there are no differences in  
415 distribution, magnitude and direction between the principal strains predicted by FE models  
416 built using different segmentation approaches, and between these and the principal strains  
417 measured *in vitro*.

418 Bite forces were measured during the loading experiments and the predicted bite force was  
419 obtained from each model after loading. The vector of the load applied to the

420 neurocranium was adjusted until the bite force predicted in model 5 matched the force  
421 measured *in vitro*. A change in  $0.1^\circ$  in load orientation (or skull orientation) produced a  
422 difference of about 1 N in predicted bite force. The predicted bite forces from the lower  
423 resolution models were up to 3% higher when the same loads and constraints were applied  
424 to them, presumably reflecting subtle differences in how the applied load is transferred to  
425 the constraints when model resolution is reduced.

426 Model sensitivity to varying construction approaches was assessed in terms of strain  
427 magnitudes, contour plots and principal strain vector orientations. To date, this study  
428 presents the largest full field surface strain measurement and comparison carried out on a  
429 cranium. Additionally a Procrustes size and shape analysis compared global deformations  
430 among models.

431 The results of experiments conducted to test the hypotheses and considerations with  
432 regard to the use of simplifications when building FE models of the human cranium are  
433 discussed below.

#### 434 **Measured vs. predicted strains**

435 This study used a voxel-based approach for FE mesh generation that is fast and automated,  
436 facilitating the process of model construction (Keyak et al. 1990; Lengsfeld et al. 1998).  
437 The results show that, irrespective of model geometry and resolution, the FE models  
438 predict strain distributions (i.e. distribution of regions of relatively high or low strain) that  
439 are similar to those measured in the cranium under experimental loading. The main  
440 differences are in strain magnitudes; with the results from models with cortical and  
441 cancellous bone represented separately being closest to the values measured *in vitro*. Among  
442 these models, those with careful reconstruction of sinus and nasal walls showed the best  
443 overall fit to *in vitro* data. This is expected; anatomically more accurate FE models behave  
444 more similarly to the real cranium under experimental loadings than do simplified models  
445 (Marinescu et al. 2005; Strait et al. 2005; Kupczik et al. 2007). In the frontal process of the  
446 maxilla,  $\epsilon_1$  strains of models 2 and 4 better match the *in vitro* strain magnitudes than the  
447 remaining models, but only for a part of the traced line lengths.  $\epsilon_3$  strains in models 2 and 4  
448 differ from the *in vitro* range (Fig. 5). The strain magnitudes along the traced lines (on Fig.  
449 3a) show the lowest correlation with model 5 for models 2 and 4 (Table 2). These results  
450 reflect an issue in model building where the sinus and nasal walls are thinner than the width  
451 of a voxel. By excluding the walls, the model is more flexible; for  $\epsilon_1$  this results in a closer

452 match in parts but for  $\epsilon_3$  a worse match than if the walls are reconstructed. This problem of  
453 how to represent very thin structures in low resolution models has no clear solution.  
454 However the models with reconstructed sinus and nasal walls generally perform more  
455 reliably than those without, and hence reconstructing them, even though they appear  
456 thicker than they are in reality, would be a reasonable way to address this problem.

457 In model 1 where cancellous bone is represented as a solid material with properties of  
458 cortical bone, strains were on average about 3.5 times lower than in the more detailed  
459 models. Thus, not including cancellous bone as a low modulus distinct material produces a  
460 significant increase in model stiffness. However, surface strain distributions (rather than  
461 magnitudes) in the contour maps remain approximately consistent among all models (1, 3  
462 and 5) with reconstructed sinus and nasal walls. This is more evident when the contour  
463 plots of these three models are scaled individually to use a similar range of the colour map  
464 (Fig. 2c). These results parallel those of (Fitton et al. 2015) and support the use of the  
465 simplification approaches used here if *relative* rather than *absolute* magnitudes of strains are  
466 of interest since they have limited local impacts on strain contours. **The reduction in strains  
467 due to stiffening of the cancellous bone material between models reflects the findings of  
468 Renders et al. (2011) who noted a reduction in stresses with increasing trabecular mineral  
469 density heterogeneity in study of bone from the mandibular condyle.** These findings are of  
470 importance in FEA studies where accurate representation of cancellous bone or sinus and  
471 nasal walls is not possible such as in fossils or damaged archaeological material or where  
472 the construction of high resolution models is impractical. However, attention should be  
473 paid when comparison is made among individuals of significantly different sizes, where  
474 there is a possibility that the distribution of cancellous bone differs allometrically (i.e. larger  
475 individuals having disproportionately more extensive areas of cancellous bone and vice  
476 versa), potentially impacting on modes of deformation (Chamoli and Wroe 2011).

477 Model resolution, over the limited range assessed here, has no appreciable effect on model  
478 performance, and suggests that the model is close to convergence in the areas investigated.  
479 However, since there was no CT scan with a higher resolution available, increasing model  
480 resolution in this study was effected by increasing element number, this may not accurately  
481 replicate the true differences in resolution of scan data.

482 The effect of another parameter of importance in FEA, material properties, was not  
483 considered in this study although it is known that cranial skeletal material properties are  
484 heterogeneous (McElhaney et al. 1970; Dechow et al. 1993; Peterson and Dechow 2003;

485 Schwartz-Dabney and Dechow 2003). The use of linearly elastic, isotropic material  
486 properties of bone homogeneously throughout the skull is common in FEA (Kupczik et al.  
487 2009; Wroe et al. 2010; Bright and Gröning 2011; Gröning et al. 2012). Using  
488 heterogeneous material properties improved model accuracy in a study by Strait et al.  
489 (2005), but this required a large amount of preliminary work in mapping and representing  
490 heterogeneity and it considerably increased model complexity to achieve solution.  
491 **Moreover, determination of material properties is impossible in fossil material and**  
492 **impractical in studies based on medical CTs from living individuals, which are usually of**  
493 **too low a resolution to allow accurate material property determination based on Hounsfield**  
494 **units.** However, several validation and sensitivity analyses support the use of simplified,  
495 homogeneous, material properties throughout the skull, since such models achieved results  
496 reasonably close to experimental data (Strait et al. 2005; Kupczik et al. 2007; Gröning et al.  
497 2009; Szwedowski et al. 2011). The empirical findings of the present study indicate that  
498 using linearly elastic, isotropic and homogeneous material properties for the cranium and  
499 teeth, results in good concordance between predicted and measured strain contours when  
500 the sinus and nasal walls are represented in the model. However this depends on accuracy  
501 in representing model geometry, in replicating the experimental loading conditions, and on  
502 the choices made with regard to material properties. In the present study we directly  
503 measured  $E$  in two locations, the maxillary tuberosity ( $E = 16.3 \pm 3.7$  GPa) and the  
504 zygomatic arch ( $E = 21.9 \pm 2.7$  GPa). It turned out that using an intermediate value,  
505 achieved strain magnitudes that reasonably matched measured ones, but other values for  $E$   
506 could also have been chosen and the choice of homogenous, isotropic material properties  
507 is arguably a source of error that would tend to make the model more or less flexible  
508 (affecting magnitude rather than mode of deformation). In this regard it is worth noting  
509 that, in a study in which material properties of a macaque skull were varied, Berthaume et  
510 al. (2012), found that 'large variations in modest-to-high *strains* and lower variations in  
511 modest-to-high *stresses* occur due to variation in material property values'. Thus, beyond  
512 the impact of simplifications of the FE model described here, errors in allocation of  
513 material properties also produce errors and so uncertainties with regard to estimated  
514 strains. The sum of such errors could potentially have a significant impact on, and limit,  
515 comparative studies of cranial biting performance. Further, Daegling et al. (2015) found  
516 that there is significant individual variation of material properties in the mandible, such that  
517 to incorporate them in a specific model, requires specimen specific measurement.  
518 However, we achieve a good match between strains in our most detailed homogenous,

519 isotropic model and those measured experimentally. Given that errors in material property  
520 allocation can have a marked effect, and that specimen specific data are not readily acquired  
521 (although they can be approximated directly from CT density) it seems reasonable to prefer  
522 simplified homogenous isotropic properties when accurate and detailed specimen specific  
523 data are not available.

524 Considering all of these results, model construction using simplification approaches that  
525 preserve sinus and nasal wall anatomy such as those described here (models 1, 3 and 5)  
526 does not appear to impact greatly on mode of deformation. However, variations in  
527 predicted strains among these models indicate that accurate estimates of strain magnitude  
528 are more difficult to achieve. It is only because we have experimental validation data that  
529 we have confidence in these predicted strain magnitudes. With fossils or in circumstances  
530 where experimental validation is impossible predicted strain magnitudes will suffer from  
531 error of unknown degree. Does this mean that prediction of cranial deformation is not  
532 possible without prior validation? A consideration of global deformations is informative in  
533 this regard.

#### 534 **Global model deformation**

535 In terms of global deformation, it is apparent that model sensitivity to how the internal  
536 sinus and nasal walls are reconstructed differs from and has greater overall impact than  
537 sensitivity to the presence of cancellous bone or variations in model resolution. Thus in the  
538 PC plot of Fig. 8 the three models (models 1, 3 and 5) with reconstructed sinus and nasal  
539 walls deform very similarly (direction of vector connecting unloaded and loaded models),  
540 differing mainly in the magnitude of deformation (length of vector connecting unloaded  
541 and loaded models). These deform differently (direction and magnitude) to models in  
542 which the sinus and nasal walls are omitted (models 2 and 4). These models manifest a  
543 higher degree and somewhat different modes of dorso-ventral maxillary bending. This  
544 contrasts with the effects of not representing cancellous bone as a separate material (model  
545 1 vs models 3 and 5), where the major impact is on the magnitude (vector length) rather  
546 than mode (vector direction) of deformation. Model resolution when varied over the range  
547 assessed in this study has little effect among models 3 and 5, whereas between models 2  
548 and 4, without inner sinus and nasal walls, the difference between models is comparatively  
549 larger.

550 It should be borne in mind that the PCA of size and shape offers quite a different insight  
551 into model performance than analyses of stresses and strains. Thus, Procrustes size and  
552 shape analyses of global deformations describe general features of deformation such as  
553 dorso-ventral bending or twisting (O'Higgins et al. 2012) while stresses and strains are  
554 relevant to prediction of failure/fracture and possibly, remodelling activity.

### 555 **Wider considerations**

556 It should be noted that the physical cranium was loaded non-physiologically because of  
557 practical constraints, but the FE models were loaded identically to allow comparison. Of  
558 course, our findings may differ from those that would have arisen from physiological  
559 loading. For instance, the zygomatic region is relatively unstrained in our study, whereas it  
560 shows high strains in experimental and modelling studies (Strait et al. 2009; Bright and  
561 Gröning 2011; Berthaume et al. 2012; Fitton et al. 2015) and lower strains when the  
562 masseter muscle is deactivated (Fitton et al., 2012). This said, the extent to which these  
563 findings of high zygomatic region strains reflect reality has been questioned by Curtis et al.  
564 (2011), who found that inclusion of temporal fascia in an FE model of a macaque greatly  
565 reduced strains in this region. Beyond this limitation, only one loading scenario, at a single  
566 bite point has been assessed. Both the non-physiological and limited loading scenarios used  
567 in this study may well lead to its findings not reflecting the full complexity and detail of  
568 differences among modelling approaches and between these and the physical cranium. This  
569 should be borne in mind when generalising from the present findings.

570 Using diverse approaches to comparing FE model performance (strain contour maps,  
571 strain vector magnitudes and directions, and global model deformation), we have  
572 demonstrated that simplifications in model geometry and material properties impact on the  
573 validity of FEA results. Some types of simplification such as model 1 (one material) result  
574 in smaller degrees of deformation, a 'stiffening of the cranium' (Figs. 2 and 8), while others  
575 (e.g. inaccurate lateral nasal wall reconstruction in models 3 and 4) impact on both mode  
576 and magnitude of deformation (Figs. 2 and 8). Previous work has shown that other  
577 decisions in model construction, such as varying relative force magnitudes among jaw  
578 closing muscles, impact on both mode of deformation and strain contours, while total  
579 applied muscle force impacts more on magnitude of deformation and strains (Fitton et al,  
580 2012).

581 This is important because it means that unless each model whose performance is to be  
582 compared has been separately refined using specimen specific validation data there will  
583 always be a degree of uncertainty concerning differences in mode and degree of  
584 deformation which will impact strain contour maps, strain magnitudes and assessments of  
585 global deformation. Such validation is difficult in extant and impossible in living humans  
586 and fossil material.

587 However, through this and the many validation and sensitivity analyses cited above, we  
588 know that some types of error (material properties, muscle force vector magnitudes,  
589 simplifications in model geometry of certain types) will affect magnitude rather more than  
590 mode of deformation. Further, other types of error (in e.g. relative muscle activation,  
591 muscle force vector directions, simplifications in model geometry of certain types) will  
592 impact more on mode than magnitude of deformation. Thus carefully designed  
593 experiments that keep constant muscle vectors and relative activations and apply certain  
594 simplifications of model geometry (that do not affect e.g. nasal wall anatomy) and use the  
595 same degree of homogeneity and isotropy of material properties may produce reasonable  
596 results with regard to mode but not magnitude of deformation. In such cases comparisons  
597 should cautiously be based on relative strains within models or the direction components  
598 of vectors of global deformation to minimise the risk of reaching erroneous conclusions.  
599 The validity of such analyses will, however, depend on the validity of the assumption of  
600 constant muscle load vectors and on how geometry has been simplified in each as well as  
601 on the magnitude the biological signal (the true differences in performance) relative to the  
602 magnitude of error. Much is yet to be learned through careful sensitivity and validation  
603 studies before the impact of modelling and loading errors is fully understood and the field  
604 can be confident that differences in model performance reflect biological reality.

605 It may be more secure to adopt an explicitly experimental approach to the application of  
606 FEA to comparative cranial functional analyses, asking specific questions about the impact  
607 of particular aspects of morphology on cranial performance. This approach maintains all  
608 aspects of the model and loading constant except for the feature of interest (e.g. sutures,  
609 periodontal ligament; Moazen et al. 2009; Wood et al. 2011; Wang et al. 2012) which is  
610 modified and the impact on performance assessed.

611 The present study was limited by several factors. Significant but, we believe adequately  
612 corrected for (see methods), is the issue of comparing surface strains projected onto a  
613 plane (DSPI output) with predicted strains over a 3D surface. Beyond this, the use of a

614 single cadaveric specimen, does not allow us to assess variation in the validity of outputs  
615 over a range of different morphologies. This is a limitation that is imposed by the  
616 complexity of obtaining human material for such work and the effort and resources  
617 required to carry out the detailed experimental and subsequent modelling work. Uniquely,  
618 in the present study we are able to present comprehensive sensitivity and validation using a  
619 single specimen and the largest and most directly measured map of surface strains to date.  
620 The findings indicate that a fairly simple model (model 5) is able to replicate the mode and  
621 magnitude of deformation of the physical cranium. However, the several sources of error  
622 in model building have different degrees of impact on mode and magnitude of deformation  
623 and hence, on the strain contours and magnitudes. This calls for great care in the  
624 application of FEA in the wider, comparative context. Finally, all of the considerations we  
625 raise in this paper with regard to error in comparison of cranial performance are likely to  
626 also apply to greater or lesser degree to other skeletal elements.

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## 629 Conclusion

630 By comparing the strains predicted by a series of FE models of the human cranium with  
631 those measured *in vitro* in the actual specimen, the impacts of different modelling  
632 simplifications on predicted deformations were assessed. The hypothesis that there are no  
633 differences in strains predicted by the FE models and those measured in the cranium was  
634 falsified. Thus, the performance of all models differed to some degree from that of the  
635 experimentally loaded cranium. However, even though the model built with only cortical  
636 bone and teeth as distinct materials showed strain magnitudes that were about 3.5 times  
637 lower than the experimentally loaded cranium, the mode of deformation was very similar.  
638 Omitting internal sinus and nasal walls led to alterations in both modes and magnitudes of  
639 deformation.

640 The second hypothesis, that there are no differences in magnitudes and modes of  
641 deformation among finite element models of the same skull built using different  
642 approaches, was falsified. Modes of deformation (as assessed by strain vectors, contour  
643 plots and a size and shape analysis) are less sensitive to how cancellous bone is represented  
644 and to variations in model resolution, over the limited range examined here, than to  
645 variations in sinus and nasal wall representation. Thus, simplifications of cancellous bone  
646 anatomy have an impact on magnitude rather than mode of deformation while under-  
647 representation of very thin bony structures such as are found in the sinus and nasal walls  
648 impacts on both mode and magnitude of deformation. These differences suggest that  
649 comparative FEA studies of biting performance among crania will likely suffer from error,  
650 due to uncertainty in the modelling process. The extent to which this error limits our ability  
651 to make ecological inferences from crania is likely significant but requires thorough  
652 investigation.

653

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661 **Authors' contributions**

662 VT-I, LCF and PO'H: study conception and design. VT-I: FE model construction. VT-I  
663 and PO'H: DSPI and FE data analysis. VT-I, LCF, MJF and PO'H: DSPI experiments,  
664 interpretation of results and manuscript writing.

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810 **Tables**

811 **Table 1.** Characteristics of the finite element models. Young's modulus: Bone=17  
 812 GPa; cortical bone=17 GPa; cancellous bone=56 MPa; teeth=50 GPa.

Model	Voxel size (mm)	No. of elements	Materials	Material volume		Features
				mm <sup>3</sup>	%	
Model 1	0.48 x 0.48 x 0.48	4,028,280	Bone (cortical+cancellous)	448,472.94	97.96	Full manual reconstruction of sinus bony walls.
			Teeth	9,316.41	2.04	
Model 2	0.48 x 0.48 x 0.48	3,326,922	Cortical bone	327,851.44	86.71	Partial (threshold based) reconstruction of inner sinus bony walls.
			Cancellous bone	40,916.34	10.82	
			Teeth	9,316.53	2.46	
Model 3	0.48 x 0.48 x 0.48	3,504,595	Cortical bone	347,999.16	87.38	Full manual reconstruction of sinus bony walls.
			Cancellous bone	40,960.09	10.28	
			Teeth	9,316.53	2.34	
Model 4	0.35 x 0.35 x 0.35	8,817,889	Cortical bone	327,113.15	86.74	Like model 2.
			Cancellous bone	40,734.59	10.80	
			Teeth	9,284.42	2.46	
Model 5	0.35 x 0.35 x 0.35	9,241,525	Cortical bone	345,217.06	87.34	Like model 3.
			Cancellous bone	40,749.30	10.31	
			Teeth	9,284.29	2.35	

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815 **Table 2.** Correlation of strain magnitudes between the most detailed model (5) and the  
 816 other models.

Model 5	Principal strains	Linear correlations ( $r$ )			
		Model 1	Model 2	Model 3	Model 4
Line 1	$\epsilon_1$	0.91	0.83	0.97	0.90
	$\epsilon_3$	0.91	0.93	0.98	0.97
Line 2	$\epsilon_1$	0.98	0.96	0.97	0.99
	$\epsilon_3$	0.96	0.97	0.98	0.99
Line 3	$\epsilon_1$	0.80	0.71	0.88	0.75
	$\epsilon_3$	0.85	0.81	0.90	0.73
Line 4	$\epsilon_1$	0.85	0.25	0.85	0.36
	$\epsilon_3$	0.87	0.36	0.87	0.34

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819 **Figure Legends**

820 **Fig. 1.** Experimental setup for *in vitro* strain measurement. (a) Vertical compressive load  
821 applied to the calvarium (upper arrow) simulating a left central incisor bite (lower arrow).  
822 The asterisk shows the DSPI sensor attached to the infraorbital region. (b) DSPI-based  
823 surface strain measurement, where the unstrained surface (upper image) provides a speckle  
824 interferogram that changes under load. The change is quantified in a phase map (middle  
825 image). Surface strains are calculated from 3D displacements, and expressed as colour-  
826 coded strain contour plots and strain vector orientations (lower image). The position of the  
827 nose is shown for reference.

828 **Fig. 2.** Cranium and finite element models. (a) Coronal section of the CT (Cranium) and  
829 the five FE models showing the results produced by different segmentations; green  
830 represents cortical bone, red represents cancellous bone and white represents teeth. (b)  
831 Cranium with overlaid DSPI results, and FE models showing maximum principal strain  $\epsilon_1$   
832 (upper row) and minimum principal strain  $\epsilon_3$  (lower row) strain contour plots. (c) Adjusted  
833 ranges of  $\epsilon_1$  (upper row) and  $\epsilon_3$  (lower row) contour plots for models 1, 3 and 5 to match  
834 the strain distributions of DSPI on the cranium, and models 2 and 4.

835 **Fig. 3.** Lines for extracting strain magnitudes and landmarks for size and shape analysis. (a)  
836 Landmark lines on the FE model surface. (b) Corresponding lines in the DSPI outputs. (c)  
837 Landmarks for Procrustes size and shape analysis.

838 **Fig. 4.** *In vitro* vs. predicted strain magnitudes across the infraorbital region. The grey area  
839 represents the mean measured (DSPI) strains  $\pm 2$  standard deviations (SD). The strain  
840 magnitudes predicted for model 1 multiplied by 3.5 were also plotted; this approximately  
841 corrects for increased model stiffness due to infilled cancellous bone.

842 **Fig. 5.** *In vitro* vs. predicted strain magnitudes across the frontal process of the maxilla. The  
843 grey area represents the mean measured (DSPI) strains  $\pm 2$  standard deviations (SD). The  
844 strain magnitudes predicted for model 1 multiplied by 3.5 were also plotted; this  
845 approximately corrects for increased model stiffness due to infilled cancellous bone.

846 **Fig. 6.** *In vitro* vs. predicted directions of strains in the infraorbital region. Black lines  
847 represent the vectors of strains in 2D (DSPI) and 3D (FE models). (a) maximum principal  
848 strain  $\epsilon_1$  and (b) minimum principal strain  $\epsilon_3$ . To best match contours and to facilitate the  
849 identification of corresponding regions, vector magnitudes in the FEA outputs and ranges  
850 of each strain contour plot have been independently adjusted.

851 **Fig. 7.** *In vitro* vs. predicted directions of strains in the frontal process of the maxilla. Black  
852 lines represent the vectors of strains in 2D (DSPI) and 3D (FE models). (a) Maximum  
853 principal strain  $\epsilon_1$  and (b) minimum principal strain  $\epsilon_3$ . To best match contours and to  
854 facilitate the identification of corresponding regions, vector magnitudes in the FEA outputs  
855 and ranges of each strain contour plot have been independently adjusted.

856 **Fig. 8.** Principal components analysis of size and shape variables based on 51 landmarks  
857 representing deformation of models 1 to 5 under a simulated incisor bite respect to the  
858 unloaded cranium. Deformations are magnified 250 times to facilitate visualisation.

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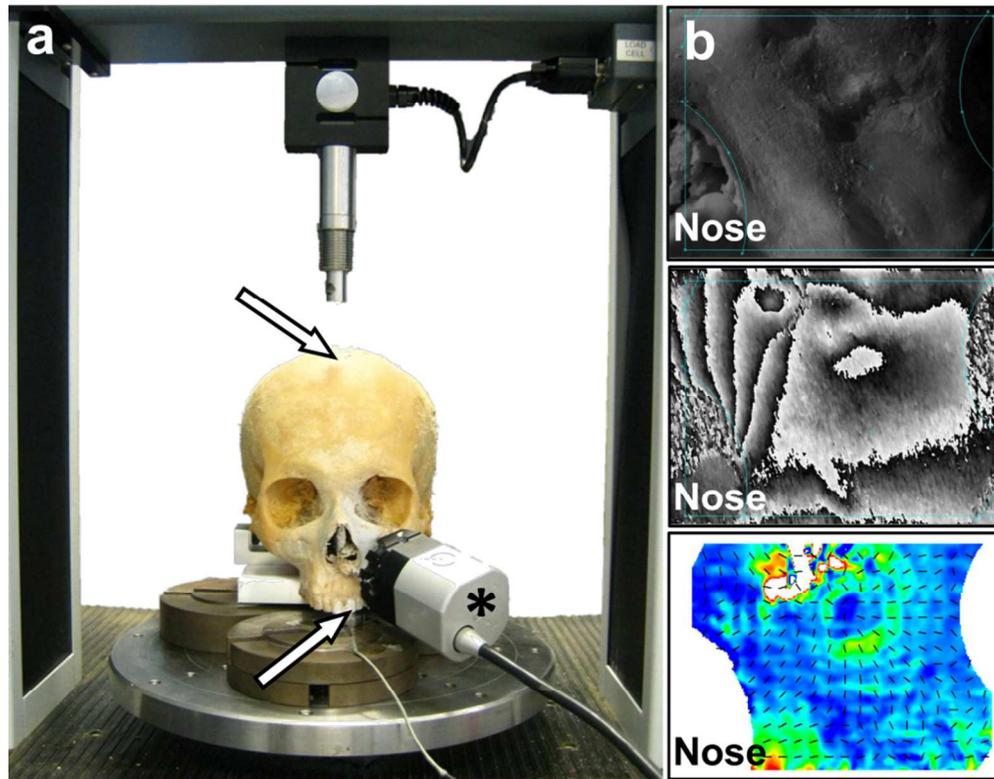


Fig. 1. Experimental setup for in vitro strain measurement. (a) Vertical compressive load applied to the calvarium (upper arrow) simulating a left central incisor bite (lower arrow). The asterisk shows the DSPI sensor attached to the infraorbital region. (b) DSPI-based surface strain measurement, where the unstrained surface (upper image) provides a speckle interferogram that changes under load. The change is quantified in a phase map (middle image). Surface strains are calculated from 3D displacements, and expressed as colour-coded strain contour plots and strain vector orientations (lower image). The position of the nose is shown for reference.  
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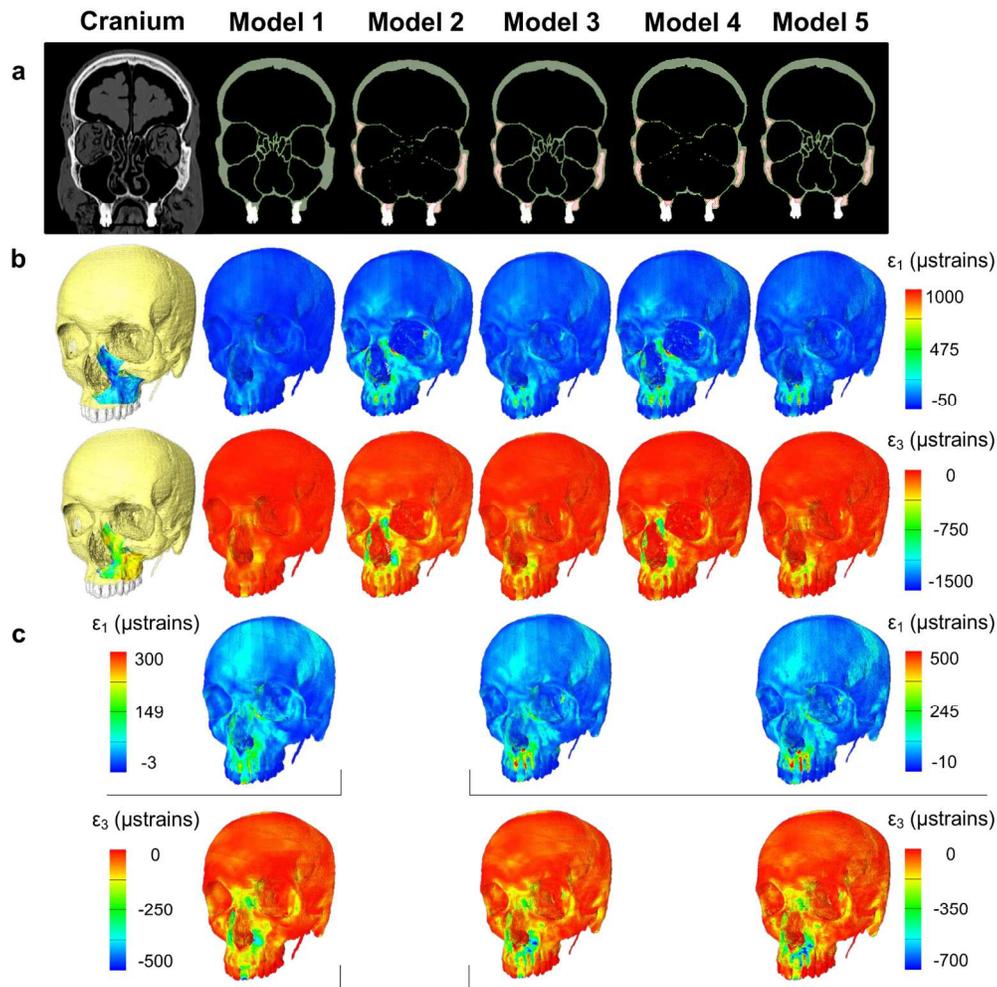


Fig. 2. Cranium and finite element models. (a) Coronal section of the CT (Cranium) and the five FE models showing the results produced by different segmentations; green represents cortical bone, red represents cancellous bone and white represents teeth. (b) Cranium with overlaid DSPI results, and FE models showing maximum principal strain  $\epsilon_1$  (upper row) and minimum principal strain  $\epsilon_3$  (lower row) strain contour plots. (c) Adjusted ranges of  $\epsilon_1$  (upper row) and  $\epsilon_3$  (lower row) contour plots for models 1, 3 and 5 to match the strain distributions of DSPI on the cranium, and models 2 and 4.  
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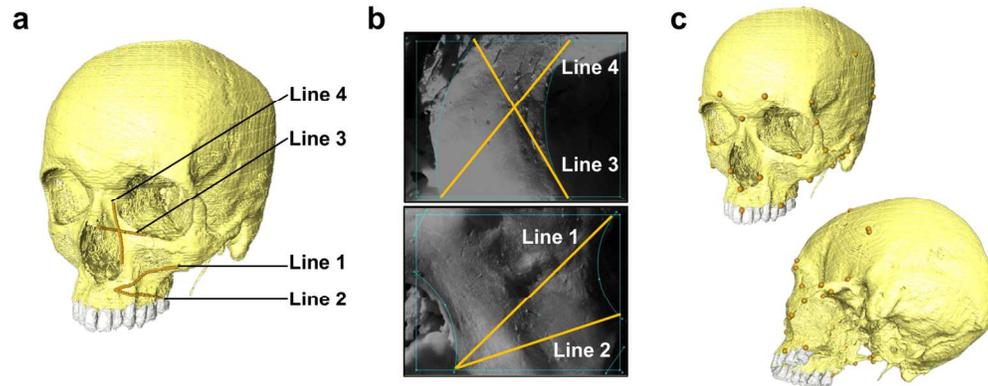


Fig. 3. Lines for extracting strain magnitudes and landmarks for size and shape analysis. (a) Landmark lines on the FE model surface. (b) Corresponding lines in the DSPI outputs. (c) Landmarks for Procrustes size and shape analysis.

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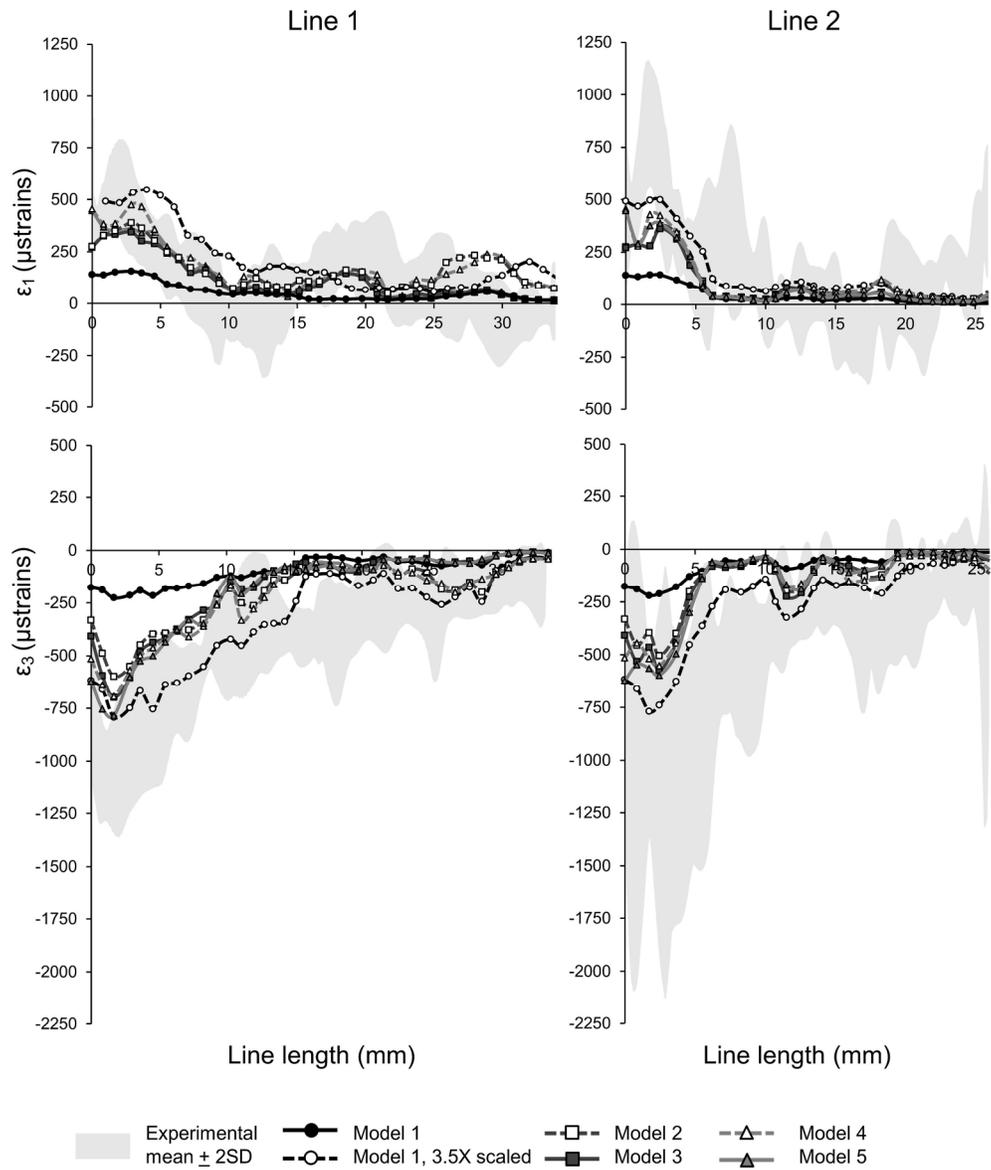


Fig. 4. In vitro vs. predicted strain magnitudes across the infraorbital region. The grey area represents the mean measured (DSPI) strains + 2 standard deviations (SD). The strain magnitudes predicted for model 1 multiplied by 3.5 were also plotted; this approximately corrects for increased model stiffness due to infilled cancellous bone.  
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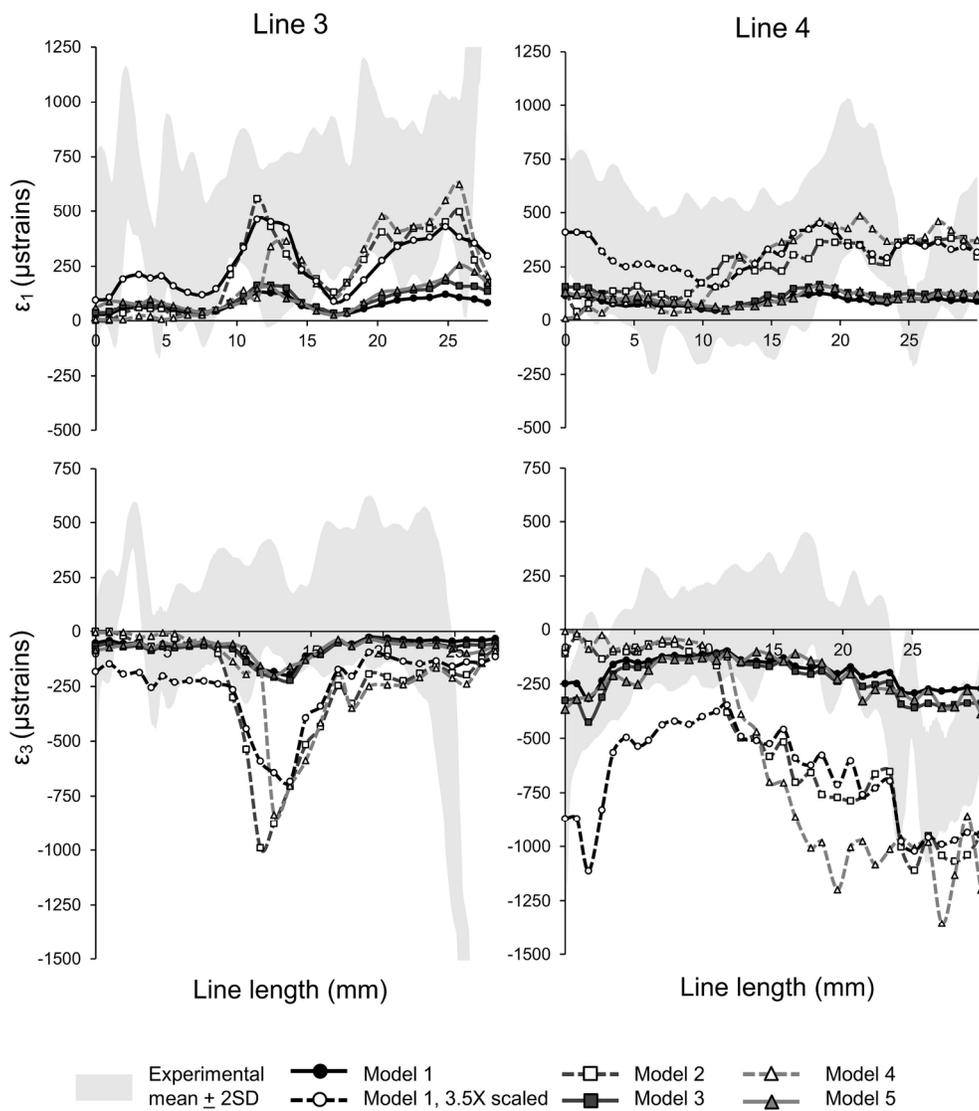


Fig. 5. In vitro vs. predicted strain magnitudes across the frontal process of the maxilla. The grey area represents the mean measured (DSPI) strains + 2 standard deviations (SD). The strain magnitudes predicted for model 1 multiplied by 3.5 were also plotted; this approximately corrects for increased model stiffness due to infilled cancellous bone.  
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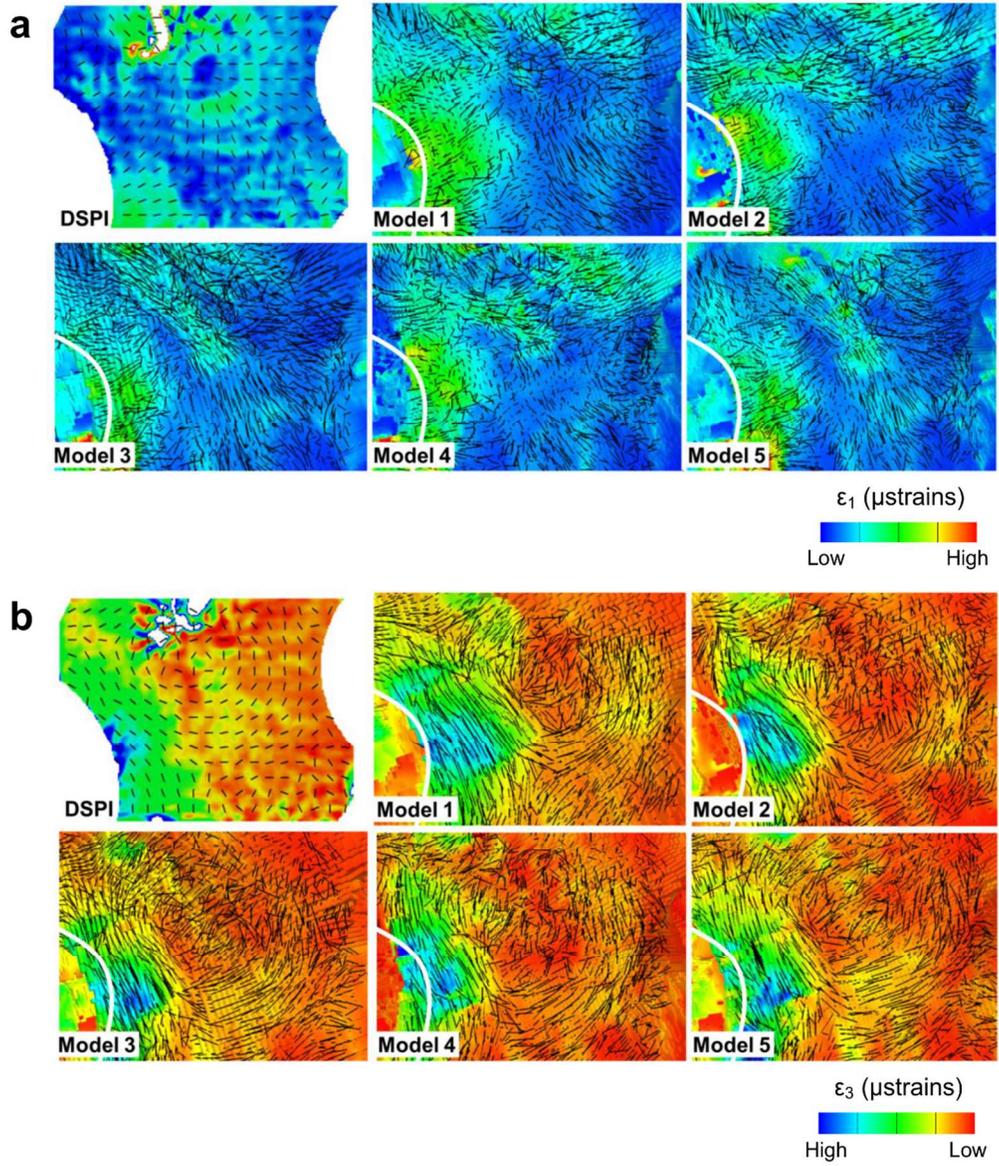


Fig. 6. In vitro vs. predicted directions of strains in the infraorbital region. Black lines represent the vectors of strains in 2D (DSPI) and 3D (FE models). (a) maximum principal strain  $\epsilon_1$  and (b) minimum principal strain  $\epsilon_3$ . To best match contours and to facilitate the identification of corresponding regions, vector magnitudes in the FEA outputs and ranges of each strain contour plot have been independently adjusted. 110x130mm (300 x 300 DPI)

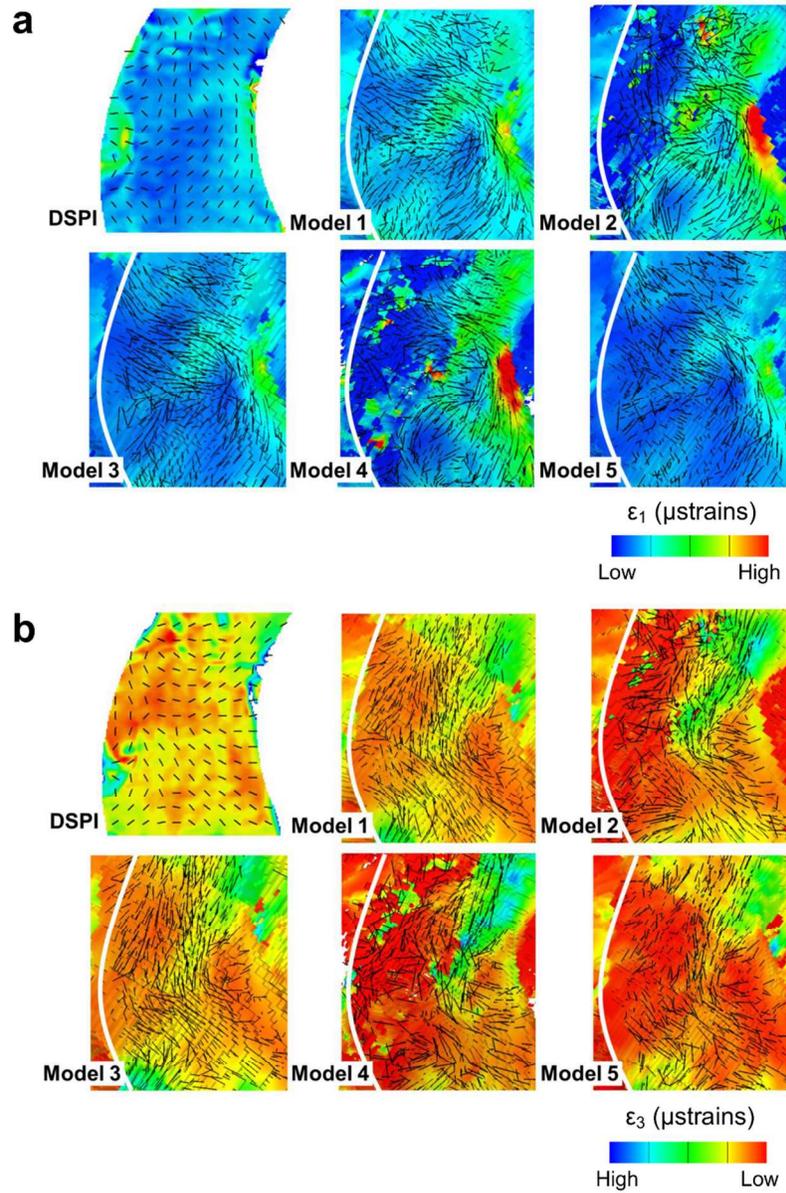


Fig. 7. In vitro vs. predicted directions of strains in the frontal process of the maxilla. Black lines represent the vectors of strains in 2D (DSPI) and 3D (FE models). (a) Maximum principal strain  $\epsilon_1$  and (b) minimum principal strain  $\epsilon_3$ . To best match contours and to facilitate the identification of corresponding regions, vector magnitudes in the FEA outputs and ranges of each strain contour plot have been independently adjusted.

87x131mm (300 x 300 DPI)

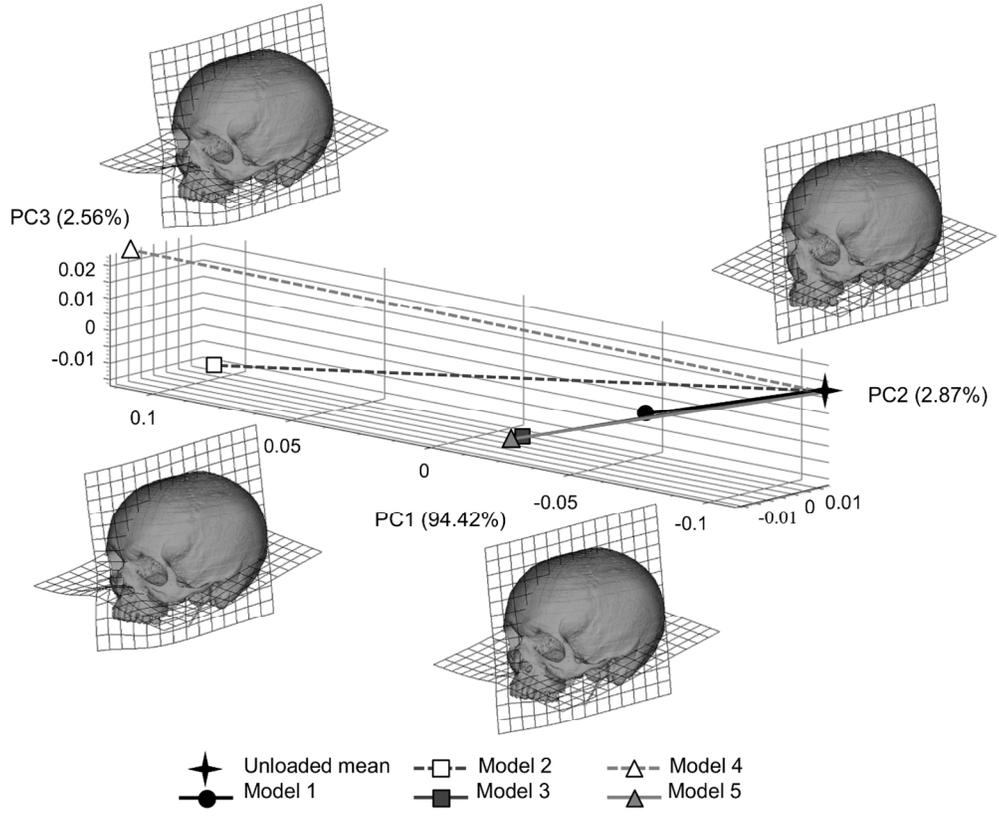


Fig. 8. Principal components analysis of size and shape variables based on 51 landmarks representing deformation of models 1 to 5 under a simulated incisor bite respect to the unloaded cranium. Deformations are magnified 250 times to facilitate visualisation.  
119x98mm (300 x 300 DPI)

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## Supporting Information

**Table 1.** Landmarks for Procrustes size and shape analysis.

No.	Definition
1	Vertex - highest point of the cranial vault.
2	Nasion - intersection between frontonasal and internasal suture.
3	Anterior nasal spine - tip of the anterior nasal spine.
4	Prosthion - most buccal and occlusal point of the interalveolar septum between central incisors.
5	Occiput - most posterior point of the neurocranium.
6&20	Supraorbital torus - most anterior point of supraorbital ridge.
7&21	Infraorbitale - most inferior point of the infraorbital ridge.
8&22	Nasal notch - most lateral part of the nasal aperture.
9&23	First molar - most buccal and mesial point of the junction of the M1 and alveolar process. If M1 is absent, the landmark is in the lowest most buccal point of the interalveolar septum between the second premolar and the next present molar.
10&24	Last molar – most buccal and distal point of the junction between the last molar and the alveolar process.
11&25	Zygo-maxillar - most inferior point of the zygo-maxillary junction.
12&26	Fronto-zygomatic - most lateral point of the fronto-zygomatic junction.
13&27	Fronto-temporal angle - point of intersection between the frontal and temporal processes of the zygomatic bone.
14&28	Zygomatic arch lateral - most lateral point of the zygomatic arch.
15&29	Zygomatic root posterior - most posterior-superior point of the intersection zygomatic root and the squama of the temporal bone.
16&30	Zygomatic root anterior - most anterior point of the intersection between the zygomatic root and the squama of the temporal bone.
17&31	Zygomatic arch medial - most lateral point on the inner face of the zygomatic arch.
18&32	Infratemporal crest - most medial point of the infratemporal crest.
19&33	Eurion - most lateral point of the neurocranium.
34&37	Anterior temporal - most anterior point of the origin of the temporal muscle in the temporal line.
35&38	Superior temporal - most superior point of the origin of the temporal muscle in the temporal line.
36&39	Posterior temporal - most posterior point of the origin of the temporal muscle in the temporal line.
40&43	Anterior masseter - most anterior point of the origin of the masseter muscle in the zygomatic arch.
41&44	Posterior masseter - most posterior point of the origin of the masseter muscle in the zygomatic arch.
42&45	Mid-masseter - midpoint along the origin area of the masseter muscle in the zygomatic arch.
46&49	Superior pterygoid - most superior point of the origin of medial pterygoid muscle in the pterygoid fossa.
47&50	Inferior pterygoid - most inferior point of the origin of medial pterygoid muscle in the pterygoid fossa.
48&51	Mid-ptyergoid - midpoint along the origin area of the medial pterygoid muscle in the pterygoid fossa.