

# No conclusive evidence of group selection in spiders

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Any field study showing convincing evidence of group selection [GS] would be a significant contribution to the field of evolutionary biology. Pruitt and Goodnight (2014)<sup>1</sup> [PG14] claim to provide such evidence in a 14-18 months field experiment on spiders. However, we contend that critical flaws in their predictions, assumptions, methods and interpretations undermine this claim. The data presented are unreliable and are equally consistent with GS and individual-level selection [ILS]; thus, the authors cannot credibly conclude that GS has produced the observed patterns.

## 1. Predictions: No critical test of GS

Evaluating GS involves, at a bare minimum, estimating and comparing both individual and group fitness, as stated by previous reviews<sup>2,3</sup> and performed by other studies<sup>4,5</sup>. Yet PG14 did not estimate individual fitness, and so cannot evaluate the relative importance of GS compared to ILS. The chosen species, *Anelosimus studiosus*, is solitary, rarely forms groups<sup>6</sup>, and shows no evidence of reproductive restraint or skew within groups<sup>7</sup>. Thus, individual and group fitness are not expected to conflict and are generally confounded, emphasising how crucial it is, firstly, to formulate predictions capable of distinguishing ILS and GS explanations and, secondly, to estimate individual fitness.

Both of PG14's predictions could follow equally well from ILS as from GS: Prediction 1) "Compositions [i.e. within-group phenotypic frequencies] that approximate the normal mixtures that characterize each site will enjoy greater success". Merely demonstrating differential survival of groups does not allow the authors to distinguish successful groups from groups of successful individuals. PG14's "group trait" is a group size dependent behavioural polymorphism. Experimental changes in this "group trait" (i.e. manipulating group size and phenotype frequency) may directly affect within-group individual fitness just as well as whole-group fitness<sup>8-11</sup>. Specifically, creating experimental groups that deviate from locally stable polymorphisms may reduce mean *individual* fitness, rendering group extinction more likely. The prediction of differential group extinction can therefore result from ILS just as plausibly as from GS. Similarly ambiguous is Prediction 2) "Colonies should only be able to adaptively hone compositions when composed of native individuals". If 'Native colonies' can "adaptively" change phenotype frequencies over time, this may occur via several mechanisms, as PG14 mention (plasticity, phenotype-biased dispersal, etc.). Yet, any of these mechanisms may evolve by ILS, a possibility ignored by PG14.

## 2. Assumptions: Unreliable selection pressure

PG14's conclusions rest upon the assumption that 'naturally-occurring mixtures' (i.e. field phenotypic frequencies, PG14's Fig. 1a) represent consistent selection pressures across years. Yet, the years of measurement were patchy (2007-2014), differed among sites and often did not overlap (Table 1). Indeed, PG14 sampled significantly different phenotypic mixtures and group sizes among years at each site (mixtures:  $p = 1.1 \times 10^{-5}$ ; group size:  $p = 7.8 \times 10^{-14}$ ; Fisher's combined  $p$ -value across separate Kruskal-Wallis tests for each site) but ignored this variation and pooled dissimilar data. In four out of nine samples taken at high resource sites, the selection pressure was no different from zero (non-significant correlations between mixtures and group size; separate lm for each year at each site), and in two out of the three low resource sites the relationship between mixture and group size differed significantly from year to year (Norris Dam:  $p = 0.0074$ ; Don Carter:  $p = 0.017$ ; lm: interaction between log(group size) and year on phenotypic mixtures). Moreover, half of the sites had not been assessed for four to six years. These measurements cannot be assumed to represent consistent, current selection pressures.

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### 59 **3. Methods: Unreliable group trait**

60 PG14 infer GS by comparing parental [P] and grand-offspring [F2] generations based on parental  
61 traits alone: they compare P-phenotypic compositions of P-colonies with P- (not F2)  
62 compositions of surviving F2-colonies (PG14's Fig. 1c is identical to Fig. 1b, minus extinct  
63 colonies: F2-colonies are depicted with their grandparents' compositions). It stretches credibility  
64 to assume that past compositions are visible to selection but present compositions are not.  
65 Indeed, 'Foreign colonies' changed to display F2-compositions in a pattern *opposite* to the  
66 assumed selection pressure (Fig. 1; PG14's Fig. 2). These changes mean that F1-compositions  
67 presumably also differed from P-compositions (and were visible to selection during that  
68 generation) but F1 was not assessed (Fig. 1).

69 Changes *within* generations were also not considered (Fig. 1). Around egg hatching<sup>12</sup>, colonies  
70 peak in size, after which mortality and dispersal decrease colony size. Phenotypic composition  
71 cannot be considered a stable "group trait" when its proposed selective advantage is a function of  
72 group size, and group size changes nonlinearly over time. Compounding this, compositions of P  
73 and F2 were apparently measured at different developmental stages (Fig. 1): a serious flaw, as  
74 individual phenotypes are affected by reproductive status<sup>13</sup>. PG14 compared sexually mature  
75 females (P) with grandchildren (F2) that presumably were juvenile, mixed-gender, and receiving  
76 maternal care (Fig. 1; mothers die off in October<sup>12</sup>; juvenile spiders are unsexable).

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### 78 **4. Interpretations: No evidence of evolution**

79 None of their findings supports PG14's puzzling claim to have demonstrated a "marked  
80 evolutionary response to GS". Rather, after two generations, surviving 'Foreign colonies' *failed*  
81 to change phenotypic compositions in site-appropriate ways (instead changing to express  
82 compositions appropriate for their original site), suggesting a lack of genetic change over the  
83 experiment (Fig. 1). PG14 argue that this constitutes evidence that mechanisms for adjusting  
84 compositions are locally adapted due to historical GS, but provide no justification for this claim:  
85 while they provide data suggesting phenotypes themselves may be partially heritable, there is no  
86 evidence that this "adjustment mechanism" has undergone genetic change and, again, no attempt

to reject ILS as an explanation. ‘Foreign colonies’ may revert to their native phenotype compositions without genetic change, for example due to persistent maternal or epigenetic effects, either of which may respond to ILS or GS. The evolutionary mechanisms shaping population-level differences, whether in phenotype frequencies or the means by which these change over time, simply have not been addressed in this paper.

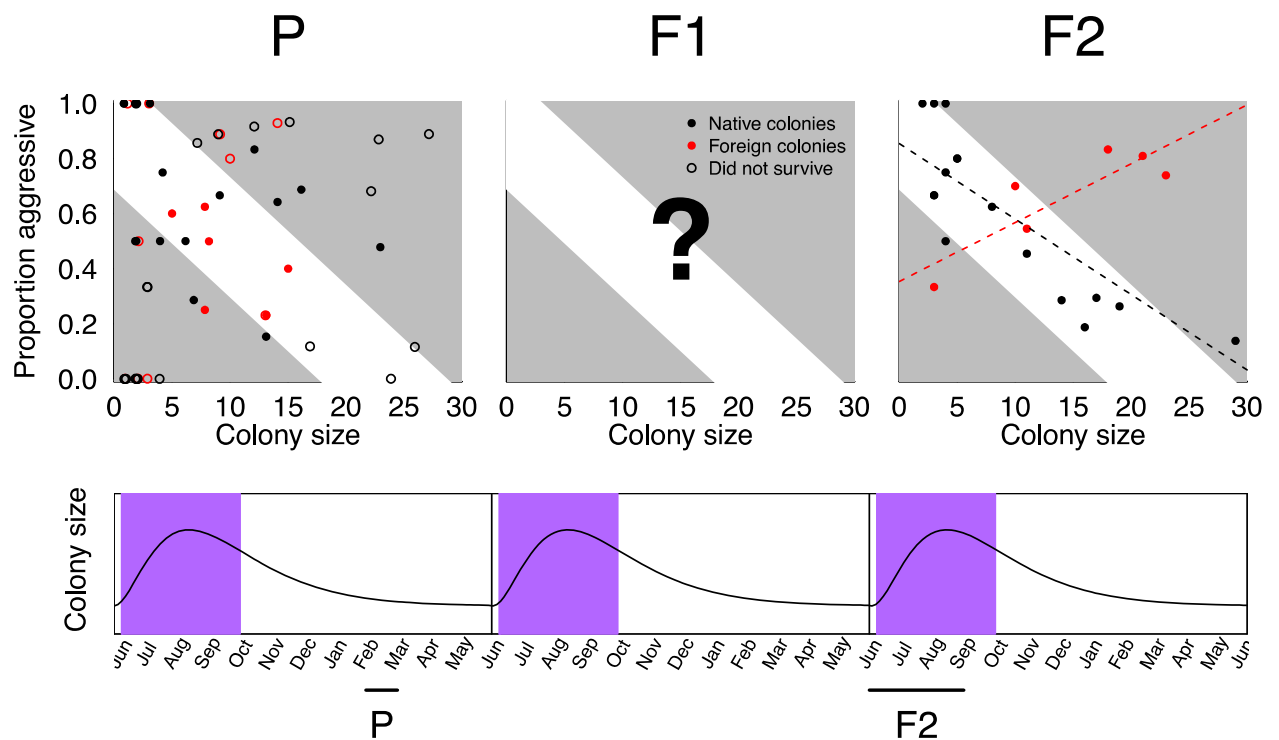
We stress that we would welcome any field study demonstrating that GS causes genetic change over generations in ways inconsistent with ILS. Given recent high-profile exchanges over the relative importance of GS<sup>14,15</sup>, such a paper would be a significant contribution to the field. Unfortunately, PG14 is not such a study.

## Figures and tables

**Table 1.** Collection years for each field site. Data from years marked with ‘X’ were pooled within sites to create the naturally-occurring mixtures (Fig. 1a in PG14) used to infer site-specific selection pressures. Grey shaded years indicate the years during which the experiment took place.

		Collection Years							
	Site	2007	2008	2009	2010	2011	2012	2013	2014
High Resource Sites	Melton Hill	X	X	X		X	X		
	Little River	X	X						
	Moccasin Creek							X	X
Low Resource Sites	Norris Dam	X	X	X		X	X		
	Clinch River		X		X				
	Don Carter			X	X				

107 **Figure 1**



108  
109 **Figure 1: Overview of methods and results from PG14**

110 **Upper panel:** Distribution of experimental colonies placed in all of six field sites in the parental  
111 [P] generation; the missing information of the next generation [F1]; and the distribution of final  
112 compositions of the grand-offspring generation [F2]. Phenotypic compositions, i.e. proportions  
113 of aggressive individuals in each colony, are plotted against colony sizes. We present data only  
114 from one low-resource field site, ‘Don Carter’, to illustrate the setup. Black dots represent  
115 ‘Native colonies’ (created with spiders collected at Don Carter); red dots represent ‘Foreign  
116 colonies’ (spiders collected at high-resource field site ‘Moccasin Creek’). Full circles of both  
117 colours in P are colonies that were still alive in F2 (equal to PG14’s Fig. 1c); empty circles are  
118 colonies that had gone extinct by F2. The white band represents the proposed selection pressure  
119 at that field site: a regression line fitted on phenotypic compositions and colony sizes of  
120 ‘naturally-occurring’ colonies at Don Carter (based here on colonies of sizes up to 30; its  
121 thickness chosen arbitrarily). Dotted lines in F2 represent regressions of the final F2-  
122 compositions of the surviving colonies: ‘Native’ (black) versus ‘Foreign’ (red). Surviving

colonies had P-compositions close to the white ‘selection band’, but F2-compositions differed according to site of origin: ‘Native’ F2-compositions were close to the selection band while ‘Foreign’ F2-compositions followed a positive regression, dissimilar to the selection band. Setup and results were similar in the two additional low-resource sites while the three high-resource sites showed opposite trends (i.e. selection bands were positive regression lines while the ‘Foreign’ F2-regressions showed negative correlations). *Note that although ‘Foreign colonies’ end up opposite to the proposed selection pressure, PG14 still conclude that their proposed selection pressure was supported.* **Lower panel** shows how the size of an *A. studiosus* colony is expected to vary within years with a peak around egg hatching during summer. The period of maternal care is marked in purple. Black lines marked with ‘P’ and ‘F2’ indicate at which point in the life cycle PG14 performed the behavioral assays to determine the phenotypic compositions of colonies. *Note that group sizes and phenotypic compositions of P and F2 were measured at different points, comparing sexually mature females (P) with juvenile grandchildren (F2) during maternal care at a stage where offspring sex cannot be determined.*

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P

F1

F2

