

Research Article

Quantitative Scintigraphy Evaluated the Relationship between ^{131}I Therapy and Salivary Glands Function in DTC Patients: A Retrospective Analysis

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Purpose. Quantitative scintigraphy to evaluate salivary gland function changes in patients with differentiated thyroid cancer (DTC) after iodine-131 (^{131}I) treatment. **Methods.** A total of 458 patients with DTC grouped by sex and age were included. Salivary gland scintigraphy was performed to evaluate salivary gland function before and after ^{131}I treatment. The uptake fraction (UF), uptake index (UI), and excretion fraction (EF) of two pairs of parotid glands and submandibular glands were measured and compared. The Chi-square test was conducted according to function impairment count. **Results.** Salivary gland function in different age groups and sexes were quite different, especially for women <55 years old, who had decreased UF, UI, and EF of all four glands without basal injury. The secretion or uptake function of some salivary glands with basic function impairment before ^{131}I treatment was increased after iodine treatment. Only a small percentage of males showed reduced functional parameters after several treatments. The most significant difference in the count of impairment for the four salivary glands were the first and third examinations, which was more evident in women. The submandibular gland had the most significant reduction in uptake. **Conclusion.** Changes in salivary gland function are more common in young females being treated for DTC. Impairment of salivary gland function is correlated with the number of treatments and the cumulative dose of ^{131}I . Some salivary gland functions impaired before ^{131}I treatment were enhanced in the early treatment.

1. Introduction

Differentiated thyroid cancer (DTC) is a common endocrine malignancy. According to statistics, 900,590 people were diagnosed with thyroid cancer in the United States. 52,070 people are expected to be diagnosed with thyroid cancer in 2019 [1]. Surgical treatment of thyroid cancer followed by removal of residual thyroid using iodine-131 (^{131}I) is a common treatment approach, but ^{131}I can induce salivary gland damage [2–4]. ^{131}I is absorbed on the membranes of thyroid follicular cells and cancer cells through the reactive sodium iodide transporter (NIS) [5]. Salivary glands expressing NIS can also absorb ^{131}I , and the accumulation of salivary gland ^{131}I is about 30 to 40 times that of plasma levels [6]. The radiation dose of high-concentration ^{131}I is sufficient to cause salivary gland damage and affect their function [7].

Salivary gland dysfunction is mainly reflected in decreased saliva secretion. Saliva is essential for the preservation of oral health. Saliva's functions include buffering, lubricating, mineralizing, and cleaning oral tissues [8]. Saliva also has antibacterial, antiviral, and antifungal properties [9]. Changes in the quantity or quality of saliva can affect the integrity of the oral tissues leading to the appearance of conditions like dental caries, periodontal diseases, and various other oral and pharyngeal disorders [10, 11]. In addition, salivary gland dysfunction is characterized by difficulty swallowing, dental disease, and loss of taste. DTC patients who had one or more ^{131}I treatments may experience the above discomfort [12–14]. Their quality of life was affected. Therefore, determining and protecting salivary gland function in DTC patients should not be ignored. There are many examinations to assess salivary gland function, including salivary gland scintigraphy with $^{99\text{m}}\text{Tc}$ -pertechnetate [3, 4, 15], neck ultrasonography [16], salivary flow rate measurement of the whole or individual gland [17]. $^{99\text{m}}\text{Tc}$ -pertechnetate is commonly used in hospitals. Because it can quantify the uptake or secretory from individual salivary glands and calculate their function [18–20].

Through salivary gland scintigraphy, we found that some patients had impaired salivary gland function before ^{131}I treatment. We defined it as an impairment of the basic function of the salivary glands. The impairment of basic function could be associated with different factors, including Sjögren syndrome [21, 22], salivary gland obstructive disease [23], salivary gland infection [24], obesity and diabetes [25, 26], aging [27, 28] and so on. The changes in the salivary gland's function in these patients after ^{131}I treatment are worth discussing.

This study aimed to analyze the changes in salivary gland uptake and excretion function following ^{131}I treatment. And to study the relationships between the function change and different genders, age groups. The results provide clinical guidance for the protection of salivary function in DTC patients undergoing ^{131}I treatment.

2. Materials and Methods

2.1. Patients. A retrospective analysis of the hospital files from the Department of Nuclear Medicine of Tianjin

Medical University General Hospital was in this study. The salivary gland scintigraphy parameters and inpatient treatment database of DTC patients from the hospital were used. We reviewed information for DTC patients who received ^{131}I therapy from November 2014 to December 2018. All enrolled patients had two or more pre-hospital scintigraphy of salivary glands. A total of 458 patients with DTC grouped by sex and age were included. Patients with the above information missing were excluded. Total thyroidectomy was performed for all patients by thyroid surgeons, and DTC was diagnosed by postoperative pathology. According to the ATA Guidelines, we selected N1b or M1 DTC patients [2]. All patients received ^{131}I treatment 6 weeks postoperatively. Before treatment with ^{131}I , patients were advised to have a low-iodine diet for 3 weeks. After the first radioiodine treatment, patients in the study received one or more radioiodine treatments. Salivary gland function parameters were recorded by 370 MBq (10 mCi) $^{99\text{m}}\text{Tc}$ -Pertechnetate salivary gland scintillation before ^{131}I treatment. The interval between each treatment was ~6 months. The protocol for evaluating salivary gland scintigraphy is shown in Figure 1. Patients with residual thyroid tissue were given 2.96 to 5.55 GBq (30–150 mCi) dosages for each treatment [2, 29].

2.2. Salivary Gland Imaging Protocol. According to our previous reports, pre-ablation salivary gland imaging was performed under thyroid-stimulating hormone (TSH) stimulation in the morning, 4 h before the first ^{131}I intake [3, 4]. Patients were asked to fast before salivary gland imaging. Single-photon emission computed tomography was performed on a Discovery NM/CT 670 (General Electric Medical Systems, Chicago, IL, USA) while subjects laid on their back. A low-energy, parallel hole, high-resolution collimator was used with a peak value of 140 keV and a window width of 20%. Each patient received 370 MBq $^{99\text{m}}\text{Tc}$ -pertechnetate intravenously through the cubital vein. After injection, the dynamic images were continuously shot on a 256×256 matrix at minute/frame with zoom 1.5 for 15 minutes. The patients were given 0.2 g oral vitamin C at the 8th minute after injection; they were instructed to chew quickly and then keep the tablet under the tongue for about 1 minute. To accurately calculate the delivered radioactivity dose, we measured the radioactivity count in the syringe before and after the injection. Patients underwent a radio-nuclide scan as described above before every ^{131}I treatment. Salivary gland imaging was also performed under TSH stimulation.

2.3. Image Analysis. First, circular regions of interest (ROIs) were manually drawn on the parotid and submandibular glands. Parotid glands showed a similar unified background area in the bilateral temporal-orbital region, while submandibular glands appeared as a similar unified background area in the bilateral supraclavicular region. The sizes and positions of these ROIs remained the same for each scanning session. An imaging system was used to generate time-activity curves for $^{99\text{m}}\text{Tc}$ -pertechnetate uptake and excretion in counts per minute. Based on these ROIs counts and the

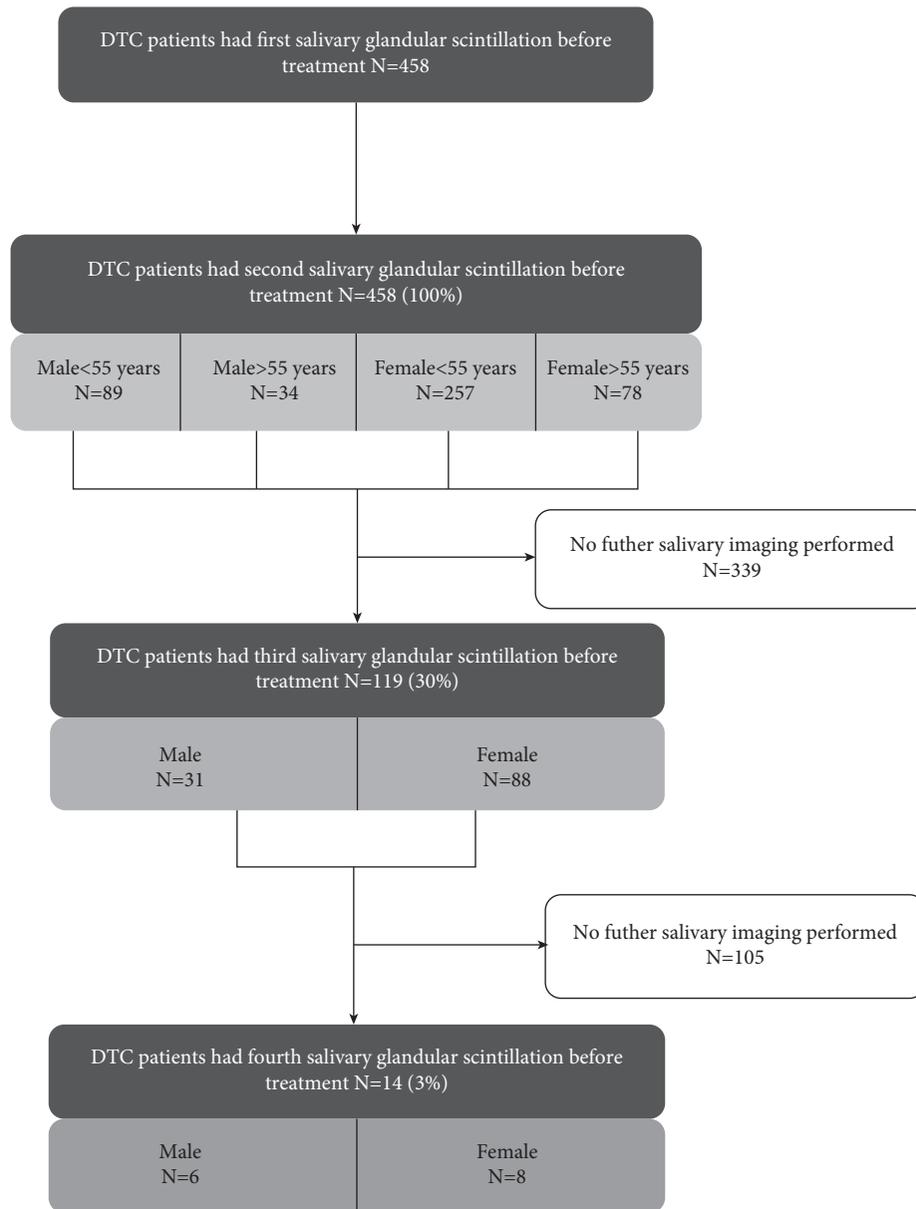


FIGURE 1: Case follow-up process. Salivary glandular scintillation imaging data were collected from DTC patients before the first, second, third, and fourth ^{131}I treatments. The group was compared according to the number of patients in each treatment course.

subsequent time-activity curves, the salivary gland functional indicators were derived using the following modified formulas [3, 4, 30, 31] (Figure 2):

2.3.1. Uptake Fraction (UF). $UF = (\text{salivary gland maximum uptake count minute} - \text{salivary gland background count corresponding to maximum count minute}) / (\text{count in the syringe before injection} - \text{count in the syringe after injection})$

2.3.2. Uptake Index (UI). $UI = (\text{salivary gland maximum uptake count minute} - \text{salivary gland background count corresponding to maximum uptake count minute}) / \text{salivary gland background count corresponding to maximum uptake count minute}$

2.3.3. Excretion Fraction (EF). $EF = (\text{salivary gland maximum uptake count minute} - \text{salivary gland minimum uptake count minute after vitamin C}) / \text{salivary gland background count corresponding to maximum uptake count minute}$

UI and UF reflect the uptake function of salivary glands, while EF reflects the secretion function.

2.4. Diagnostic Criteria for Salivary Gland Function. Salivary gland function impairment was established based on the diagnostic criteria of the Department of Nuclear Medicine, Tianjin Medical University General Hospital, with reference to previous studies and modified in our institute [18, 19, 32, 33]. Parameters obtained by salivary scintigraphy, the peak uptake (maximum salivary gland uptake count/second count at peak uptake) was set to <50 counts/s

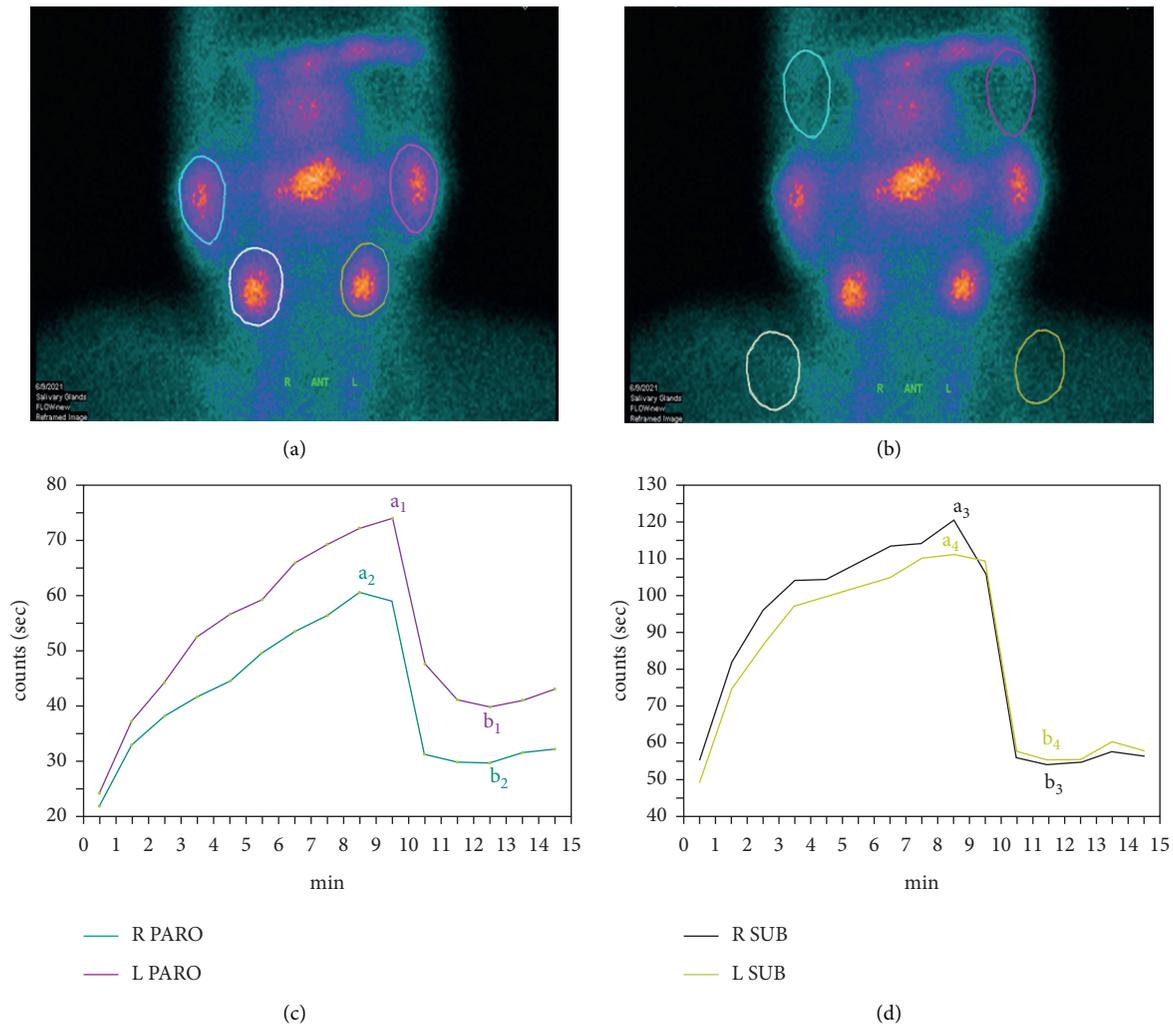


FIGURE 2: Physiological uptake and excretion of ^{99m}Tc -pertechnetate in parotid and submandibular salivary glands detected by dynamic salivary gland scintigraphy. (a) A clear depiction of the ROIs and ROIs of the salivary and mandibular glands was obtained from all frames. (b) The parotid gland was in the bilateral temporal-orbital region, and the mandibular gland was in the bilateral supraclavicular region to draw similar background areas, and the background ROIs count was recorded. (c-d) Time-activity curve of each salivary gland. The ordinate of a_1 - a_4 corresponds to the maximum minute count of each salivary gland. The ordinate of b_1 - b_4 corresponds to the count of each salivary gland at the minimum uptake minute after vitamin C.

with reduced intake function, and EF was set to $<30\%$ with reduced secretory function. A reduced diagnosis of either or both of these above salivary glands is dysfunction. The patients with salivary gland dysfunction before the first admission were those with impaired basic salivary gland function.

2.5. Statistical Analysis. All data for males and females were analyzed separately and are presented as either mean \pm standard deviation or median (upper quartile, lower quartiles). Statistical analysis was performed by using Statistical Package for Social Sciences (SPSS version 25.0, IBM Corp., Armonk, NY, USA) software. Mann-Whitney U tests were used to compare values of the same patient before and after the first treatment. Kruskal-Wallis tests were performed to evaluate and compare salivary gland function in patients who underwent multiple ^{131}I treatments. After

dividing men and women into separate groups, the distribution of salivary gland injuries before the first treatment and before the second to fourth treatments were analyzed by chi-square test.

3. Results

Among the 458 patients, most (72.9%) were female with a mean age of 46 ± 12 years (range 14–76 years). The lowest and highest doses of ^{131}I were 30 and 550 mCi, respectively. Patient demographic data are summarized in Table 1.

From the scintigraphy examinations, the analysis according to age and sex groups evidenced salivary gland function is more sensitive in females than in males. The sensitivity of salivary gland function was ranked from large to small, in order of <55 years female, ≥ 55 years female, <55 years male, ≥ 55 years male ($p = 0.05$). After treatment with

TABLE 1: Clinical characteristics of total patients.

Characteristic	Total	Male	Female	<i>P</i>
Age (years)	46 ± 12/14–76	46 ± 12/22–75	45 ± 12/14–76	0.442
Number/percentage		124/27%	334/73%	
¹³¹ I activity (mCi)	128 ± 78/30–550	131 ± 68/30–450	126 ± 80/30–550	0.145

Age and ¹³¹I activity are stated as median ± standard deviation/range.

¹³¹I, the UI, UF, or EF values of patients without impairment of basic salivary gland function tended to decrease. The secretion or uptake function of some salivary glands with basic function impairment before ¹³¹I treatment was increased after iodine treatment. Table S1 summarizes the number of patients, age distribution, and cumulative dose before one or more ¹³¹I treatments. The UI, UF, and EF of each group were compared before and after ¹³¹I treatment (Tables 2–4).

We found that the difference between the first and third injury counts of salivary gland damage on salivary gland scintigraphy tests before hospitalization was the most significant. There were statistically significant differences between the four salivary glands in both the male and female groups. It was more pronounced in the female group (female, *P* = 0.001; male, *P* < 0.05). The percentage of damaged salivary glands increased with the number of treatments in both sexes, while the percentage of normal salivary glands gradually decreased (Table 5, Figure 1S). The cumulative dose of ¹³¹I received by patients over several treatments is shown in Figure 2S. Chi-square test was performed to determine the relationship between the number of patients by sex and the impairment of salivary gland function before each ¹³¹I treatment (Table S2). There was a statistical difference in the left submandibular gland injury count between sexes before the first treatment (*p* < 0.05).

4. Discussion

Salivary gland damage is a common manifestation of DTC patients after ¹³¹I therapy [34]. ¹³¹I is mainly concentrated in the duct system of the salivary glands. The radiation causes debris buildup that narrows the lumen, and this obstruction can lead to an injurious process that results in glandular degeneration. Salivary gland scintigraphy examination is necessary for the objective evaluation of the reproducibility of salivary gland function [20]. These measurement parameters mainly assess gland uptake and secretion capacity [15, 35, 36]. We performed salivary gland scintigraphy to identify relevant salivary gland parameters and then analyzed the factors that affect salivary gland function changes in DTC patients after each ¹³¹I treatment.

4.1. Sex and Age. By comparing salivary glands without basic functional impairment, we found that functional changes were related to age and sex. The salivary glands of younger patients were more sensitive to ¹³¹I treatment than older patients, and female patients were more likely to show decreased function. Other studies have examined age- and

sex-dependent differences in salivary gland function changes. Liu et al. assessed iodine dynamics and salivary gland dosimetry after ¹³¹I treatment and showed that women's parotid iodine intake was often higher than men's [37]. This suggests that female salivary glands are more susceptible to radiation, leading to decreased function following ¹³¹I treatment. Almeida et al. found that patient sex was associated with the uptake phase on salivary glands scintigraphy. Intake of all major salivary gland was decreased in men compared with women. Patient age was the strongest predictor of parotid gland dysfunction as it affects the stage of parotid gland uptake and elimination on salivary gland scintigraphy [38]. Another study revealed the presence of epidermal and nerve growth factors (EGF and NGF) in salivary glands and described their roles in cell growth and differentiation. They are detected at higher levels in the submandibular glands of males than females [39], indicating that male salivary gland cells have stronger repair and regeneration abilities. Animal studies showed sex-attributed differences in wound healing patterns in submandibular glands between male and female mice. In males, the number of convoluted tubules rich in EGF and NGF (involved in cell proliferation and neurogenesis, respectively) was higher than that in females [40, 41]. These sex-differences observed in mice may help explain why the incidence of salivary gland disease tends to be much higher in women than in men (i.e., approximately 9:1) [42].

4.2. Different Basic Functions. Before the first ¹³¹I treatment, we selected patients with basic impairment of salivary gland function. After the first ¹³¹I treatment, some salivary gland parameters were different in the female group, with most changes in the group younger than 55 years old. After the second ¹³¹I treatment, the bilateral submandibular gland EF values were different in females but not males. Interestingly, these altered functional parameters all showed an upward trend rather than the expected decline. We consider that this may be related to the compensatory increase in the function of glandular cells under certain stress states. Poradovskaia et al. showed that after ablating or removing one submandibular salivary gland in rats, the contralateral gland responded by increasing cell proliferation with concomitant increases in the size of the cells and nuclei by 10% and 17%, respectively. Burlage et al. observed that pilocarpine preconditioning induced proliferation of acinar and intercalated duct cells in rats, which could explain the observed enhanced compensatory response in salivary glands [43]. Compensatory proliferation is a mechanism to replace lost cells in rapidly cycling tissues [44]. After an initial singular dose ¹³¹I dose of 100 mCi, the salivary glands might increase

TABLE 2: Comparison of salivary gland function in patients before secondary treatment.

Scintigraphic parameters	Male older than or equal to 55 years						Female under 55 years						Female older than or equal to 55 years					
	Male under 55 years		Male older than or equal to 55 years		Female under 55 years		Female older than or equal to 55 years		Female under 55 years		Female older than or equal to 55 years		Female under 55 years		Female older than or equal to 55 years			
	Damage before treatment	Alter firsttreatment	P	Damage before treatment	Alter firsttreatment	P	No damage before treatment	Alter firsttreatment	P	Damage before treatment	Alter firsttreatment	P	No damage before treatment	Alter firsttreatment	P	Damage before treatment	Alter firsttreatment	P
RP	0.039 (0.027-0.071)	0.039 (0.030-0.058)	0.866	0.016 (0.011-0.014)	0.018 (0.013-0.014)	0.180	0.047 (0.026-0.066)	0.043 (0.025-0.056)	0.116	0.043 (0.022-0.051)	0.043 (0.020-0.048)	0.000	0.043 (0.026-0.049)	0.038 (0.020-0.049)	0.000	0.025 (0.010-0.040)	0.027 (0.015-0.036)	0.715
UI	1.792 (1.337-2.293)	1.806 (1.424-2.423)	0.799	0.967 (0.589-0.862)	0.960 (0.707-0.732)	0.655	1.787 (1.176-2.175)	1.946 (1.264-2.643)	0.133	1.946 (1.264-2.643)	1.946 (1.264-2.643)	0.000	2.105 (1.621-2.618)	1.920 (1.467-2.369)	0.000	1.314 (0.774-2.224)	1.169 (0.836-1.466)	0.465
EF	0.109 (0.070-0.218)	0.461 (0.354-0.539)	0.064	0.306 (0.190-0.270)	0.268 (0.195-0.207)	0.655	0.444 (0.301-0.488)	0.396 (0.302-0.513)	0.151	0.465 (0.211-0.521)	0.465 (0.211-0.521)	0.001	0.496 (0.428-0.580)	0.467 (0.404-0.533)	0.001	0.176 (0.143-0.173)	0.359 (0.143-0.558)	0.144
LP	0.044 (0.019-0.048)	0.046 (0.027-0.050)	0.113	0.023 (0.011-0.023)	0.021 (0.012-0.034)	0.180	0.044 (0.024-0.072)	0.045 (0.025-0.067)	0.197	0.032 (0.013-0.036)	0.032 (0.020-0.048)	0.000	0.038 (0.020-0.053)	0.036 (0.028-0.048)	0.000	0.020 (0.010-0.110)	0.020 (0.013-0.047)	0.069
UI	0.815 (0.724-1.093)	1.689 (1.288-2.142)	0.079	1.072 (0.594-1.014)	1.312 (0.732-1.237)	0.180	1.858 (1.388-2.379)	1.567 (1.261-2.538)	0.443	1.770 (1.308-2.291)	1.770 (1.308-2.291)	0.000	2.013 (1.569-2.548)	1.841 (1.464-2.347)	0.000	1.494 (0.940-2.044)	1.560 (1.012-2.019)	0.401
EF	0.124 (0.104-0.154)	0.486 (0.377-0.549)	0.043	0.198 (0.053-0.244)	0.390 (0.227-0.388)	0.655	0.440 (0.339-0.511)	0.397 (0.304-0.498)	0.061	0.383 (0.120-0.357)	0.383 (0.139-0.467)	0.001	0.494 (0.420-0.564)	0.468 (0.394-0.535)	0.000	0.188 (0.167-0.240)	0.390 (0.281-0.487)	0.208
RS	0.033 (0.023-0.049)	0.048 (0.042-0.062)	0.000	0.023 (0.021-0.030)	0.026 (0.021-0.036)	0.285	0.057 (0.048-0.067)	0.045 (0.030-0.063)	0.011	0.034 (0.020-0.042)	0.034 (0.020-0.049)	0.000	0.047 (0.035-0.061)	0.042 (0.033-0.052)	0.000	0.010 (0.000-0.000)	0.019 (0.010-0.029)	0.345
UI	2.156 (1.297-2.530)	2.796 (2.209-3.705)	0.016	1.796 (0.818-1.989)	1.883 (1.262-2.093)	0.285	3.074 (2.395-5.531)	2.604 (2.127-3.239)	0.024	2.018 (1.582-2.324)	2.018 (1.582-2.324)	0.000	2.490 (2.002-2.945)	2.401 (1.997-2.945)	0.000	1.602 (0.986-1.907)	1.402 (0.868-1.907)	0.893
EF	0.171 (0.151-0.251)	0.200 (0.171-0.254)	0.020	0.030 (0.034-0.281)	0.030 (0.029-0.338)	0.593	0.060 (0.044-0.431)	0.060 (0.038-0.400)	0.264	0.030 (0.016-0.141)	0.030 (0.016-0.141)	0.000	0.030 (0.025-0.037)	0.030 (0.025-0.037)	0.000	0.132 (0.120-0.200)	0.132 (0.120-0.200)	0.043
SI	0.027 (0.020-0.020)	0.045 (0.010-0.040)	0.000	0.020 (0.020-0.020)	0.020 (0.020-0.020)	0.000	0.060 (0.060-0.060)	0.060 (0.060-0.060)	0.000	0.029 (0.024-0.035)	0.029 (0.024-0.035)	0.000	0.040 (0.040-0.040)	0.040 (0.040-0.040)	0.000	0.020 (0.020-0.020)	0.020 (0.020-0.020)	0.000
EF	0.161 (0.161-0.161)	0.377 (0.377-0.377)	0.000	0.377 (0.377-0.377)	0.377 (0.377-0.377)	0.000	0.377 (0.377-0.377)	0.377 (0.377-0.377)	0.000	0.173 (0.139-0.198)	0.173 (0.139-0.198)	0.000	0.380 (0.380-0.380)	0.330 (0.277-0.428)	0.000	0.110 (0.110-0.110)	0.210 (0.110-0.390)	0.128
EF	0.161 (0.161-0.161)	0.377 (0.377-0.377)	0.000	0.377 (0.377-0.377)	0.377 (0.377-0.377)	0.000	0.377 (0.377-0.377)	0.377 (0.377-0.377)	0.000	0.173 (0.139-0.198)	0.173 (0.139-0.198)	0.000	0.380 (0.380-0.380)	0.330 (0.277-0.428)	0.000	0.110 (0.110-0.110)	0.210 (0.110-0.390)	0.128

UF: UI, EF stated as median (upper quartile, lower quartiles), RP: Right submandibular; LS: Left submandibular; RS: Right submandibular; LP: Left parotid; RS: Right submandibular; UF: Uptake index; EF: Excretion fraction.

TABLE 3: Comparison of salivary gland function in patients before third treatment.

Scintigraphic parameters	Male group before third treatment				Female group before third treatment				
	Damage before treatment	After first treatment	After second treatment	P	No damage before treatment	After first treatment	After second treatment	P	
RP		Number: 2	Number: 29			Number: 2	Number: 86		
UF	0.034 (0.018-0.033)	0.057 (0.019-0.067)	0.044 (0.012-0.055)	0.223	0.050 (0.031-0.068)	0.049 (0.032-0.060)	0.043 (0.026-0.061)	0.485	0.037 (0.028-0.048)
UI	1.844 (1.054-1.712)	2.967 (0.865-3.586)	2.387 (0.864-2.717)	0.607	1.867 (1.527-2.391)	1.898 (1.440-2.819)	2.016 (1.277-3.038)	0.485	1.952 (1.549-2.479)
EF	0.254 (0.064-0.317)	0.506 (0.304-0.455)	0.299 (0.148-0.300)	0.607	0.486 (0.433-0.544)	0.478 (0.352-0.581)	0.493 (0.340-0.579)	0.422	0.485 (0.415-0.552)
LP		No case				Number: 31			
UF					0.041 (0.035-0.065)	0.045 (0.033-0.056)	0.036 (0.027-0.053)	0.206	0.019 (0.009-0.025)
UI					1.971 (1.523-2.460)	1.897 (1.348-2.457)	1.615 (1.359-2.576)	0.597	1.686 (1.393-2.281)
EF					0.510 (0.442-0.558)	0.464 (0.402-0.550)	0.455 (0.347-0.545)	0.086	0.479 (0.420-0.544)
RS		Number: 2				Number: 29			
UF	0.034 (0.017-0.033)	0.033 (0.016-0.034)	0.032 (0.021-0.027)	1.000	0.058 (0.046-0.080)	0.054 (0.042-0.066)	0.051 (0.038-0.065)	0.018	0.052 (0.039-0.068)
UI	1.553 (0.664-1.665)	1.821 (0.961-1.770)	1.636 (1.189-1.265)	0.607	3.171 (2.438-3.936)	3.105 (2.284-3.663)	3.052 (2.415-3.624)	0.639	2.685 (2.161-3.528)
EF	0.096 (0.029-0.115)	0.122 (0.038-0.146)	0.073 (0.032-0.078)	0.223	0.381 (0.335-0.467)	0.375 (0.271-0.416)	0.396 (0.318-0.468)	0.343	0.404 (0.308-0.498)
LS		Number: 2				Number: 29			
UF	0.031 (0.011-0.036)	0.033 (0.009-0.040)	0.023 (0.009-0.025)	0.607	0.056 (0.044-0.079)	0.048 (0.040-0.065)	0.050 (0.040-0.059)	0.024	0.033 (0.023-0.051)
UI	1.482 (0.656-1.568)	1.570 (0.645-1.711)	1.030 (0.421-1.125)	0.223	3.584 (2.667-4.089)	3.236 (2.655-3.797)	3.053 (2.546-3.685)	0.166	1.891 (1.437-2.864)
EF	0.169 (0.123-0.131)	0.072 (0.000-0.108)	0.034 (0.016-0.034)	0.223	0.418 (0.316-0.492)	0.372 (0.295-0.436)	0.404 (0.321-0.528)	0.166	0.169 (0.119-0.199)

UF, UI, EF stated as median (upper quartile, lower quartiles). Examination before first, second and third treatments expressed as I, II, and III. The values were p values after pairwise comparison of I, II, III. RP: Right parotid; LP: Left parotid; RS: Right submandibular; LS: Left submandibular; UF: Uptake fraction; UI: Uptake index; EF: Excretion fraction.

TABLE 4: Comparison of salivary gland function in patients before fourth treatment.

Scintigraphy parameters	Male group before fourth treatment number: 6			Female group before fourth treatment number: 8			P		
	Before treatment	After first treatment	After second treatment	After third treatment	Before treatment	After first treatment		After second treatment	After third treatment
RP									
UF	0.054 (0.024–0.072)	0.072 (0.035–0.089)	0.051 (0.025–0.077)	0.052 (0.015–0.076)	0.047 (0.043–0.056)	0.032 (0.031–0.046)	0.033 (0.018–0.037)	0.010 (0.009–0.042)	0.013
UI	2.237 (1.492–2.725)	2.317 (1.691–3.522)	2.252 (1.425–3.257)	2.127 (1.323–2.668)	2.371 (1.860–2.885)	1.690 (1.537–2.151)	1.388 (0.808–1.844)	0.707 (0.529–1.887)	0.003
EF	0.485 (0.330–0.559)	0.542 (0.460–0.705)	0.458 (0.364–0.547)	0.403 (0.004–0.548)	0.543 (0.500–0.567)	0.478 (0.439–0.487)	0.158 (0.023–0.446)	0.050 (0.022–0.407)	0.001
LP									
UF	0.040 (0.035–0.065)	0.052 (0.039–0.067)	0.041 (0.029–0.070)	0.031 (0.010–0.045)	0.041 (0.035–0.050)	0.035 (0.027–0.048)	0.033 (0.026–0.044)	0.025 (0.018–0.035)	0.034
UI	1.958 (1.661–2.357)	2.3628 (1.468–2.616)	1.623 (1.307–3.080)	1.675 (1.213–1.995)	2.051 (1.765–2.396)	1.684 (1.534–2.175)	1.635 (0.963–2.104)	1.576 (0.472–2.029)	0.010
EF	0.515 (0.480–0.589)	0.487 (0.385–0.575)	0.419 (0.216–0.548)	0.287 (0.047–0.48)	0.475 (0.454–0.526)	0.445 (0.297–0.485)	0.411 (0.031–0.501)	0.429 (0.016–0.518)	0.522
RS									
UF	0.053 (0.040–0.064)	0.057 (0.036–0.076)	0.050 (0.030–0.067)	0.040 (0.014–0.076)	0.052 (0.041–0.059)	0.039 (0.035–0.048)	0.043 (0.035–0.054)	0.037 (0.030–0.044)	0.021
UI	2.147 (1.884–2.778)	3.041 (2.015–3.985)	2.703 (1.927–3.185)	2.726 (1.660–4.742)	2.988 (2.479–3.495)	2.465 (2.125–3.067)	2.649 (2.306–2.857)	2.447 (2.239–2.601)	0.119
EF	0.386 (0.338–0.444)	0.352 (0.246–0.471)	0.394 (0.315–0.466)	0.422 (0.181–0.475)	0.437 (0.373–0.455)	0.282 (0.254–0.352)	0.284 (0.218–0.405)	0.348 (0.143–0.437)	0.010
LS									
UF	0.052 (0.044–0.056)	0.045 (0.043–0.078)	0.054 (0.041–0.063)	0.045 (0.015–0.070)	0.049 (0.042–0.056)	0.042 (0.030–0.047)	0.042 (0.034–0.049)	0.037 (0.030–0.040)	0.392
UI	2.605 (1.676–3.068)	3.420 (2.807–3.783)	3.023 (2.450–3.487)	2.904 (2.145–3.613)	2.550 (2.338–3.107)	2.620 (1.897–3.204)	2.666 (2.135–3.045)	2.472 (2.344–2.949)	0.789
EF	0.397 (0.350–0.456)	0.371 (0.255–0.427)	0.420 (0.297–0.504)	0.376 (0.166–0.453)	0.332 (0.269–0.407)	0.289 (0.259–0.355)	0.274 (0.194–0.405)	0.327 (0.129–0.415)	0.327

UF, UI, EF stated as median (upper quartile, lower quartiles). Examination before first, second, and third treatments expressed as I, II, III, and IV. The values were P values after pairwise comparison of I, II, III, IV. RP: Right parotid; LP: Left parotid; RS: Right submandibular; LS: Left submandibular; UF: Uptake fraction; UI: Uptake index; EF: Excretion fraction.

TABLE 5: The number of salivary gland injuries before the last three treatments compared with the first treatment.

Scintigraphic parameters	Damage	Male										Female				
		First	Second	P	First	Third	P	First	Fourth	P	First	Second	P	Third	Fourth	P
RP	Hurt	8 (6.5%)	8 (6.5%)	1.000	8 (6.5%)	8 (25.8%)	0.005	17 (5.1%)	2 (33.3%)	0.069	17 (5.1%)	16 (5.1%)	0.859	29 (33%)	5 (62.5%)	0.001
	No hurt	115 (93.5%)	115 (93.5%)		23 (74.2%)			318 (94.9%)	4 (66.7%)		318 (94.9%)	319 (94.9%)		59 (67%)	3 (37.5%)	
LP	Hurt	5 (4.1%)	9 (7.3%)	0.271	5 (4.1%)	7 (22.6%)	0.003	25 (7.5%)	3 (50%)	0.003	25 (7.5%)	23 (6.9%)	0.881	24 (27.3%)	2 (25%)	0.125
	No hurt	118 (95.9%)	114 (92.7%)		24 (87.4%)			310 (92.5%)	3 (50%)		310 (92.5%)	312 (93.1%)		64 (72.7%)	6 (75%)	
RS	Hurt	12 (9.8%)	19 (15.4%)	0.179	12 (9.8%)	8 (25.8%)	0.032	44 (13.1%)	2 (33.3%)	0.128	44 (13.1%)	48 (14.3%)	0.653	36 (40.9%)	2 (25%)	0.292
	No hurt	111 (90.2%)	104 (84.6%)		23 (74.2%)			291 (86.9%)	4 (66.7%)		291 (86.9%)	287 (85.7%)		52 (59.1%)	6 (75%)	
LS	Hurt	11 (8.9%)	22 (17.9%)	0.040	11 (8.9%)	8 (25.8%)	0.027	57 (17%)	2 (33.3%)	0.112	57 (17%)	62 (18.5%)	0.613	39 (44.3%)	4 (50%)	0.036
	No hurt	112 (91.1%)	101 (82.1%)		23 (74.2%)			278 (83%)	4 (66.7%)		278 (83%)	273 (81.5%)		49 (55.7%)	4 (50%)	

The sequence number of examinations before treatments. Expressed as number of cases (values of number of cases/total cases of this examination before treatments × 100%). RP: Right parotid; LP: Left parotid; RS: Right submandibular; LS: Left submandibular.

uptake to maintain secretory function [4]. Our results are consistent with those of the above-mentioned studies. It is believed that salivary glands with slight damage in the basal state have a certain compensatory function. After the initial radiation injury, the compensatory function of gland cells is activated by stress, manifesting as increased uptake or excretion. With the increase of radiation dose and the passage of time after ^{131}I treatment, this compensatory function gradually decreases or disappears. In this study, we also selected uninjured salivary glands and compared the functional parameters before and after ^{131}I therapy. Notably, these salivary glands were more likely to be affected than those that were already impaired before treatment. Most showed functional reduction without the phenomenon of functional compensatory increase. This provides new ideas for clinical treatment. For example, salivary gland protection should be strengthened during ^{131}I treatment, especially in patients with normal basal function.

4.3. Different Glands. The salivary gland imaging results after the first and second ^{131}I treatments showed that the submandibular glands are more sensitive than the parotid ones, and the most common change was a decreased UF value representing impaired uptake. We analyzed whether this difference was related to salivary gland cell characteristics and salivary gland structure. Damage to the microvascular endothelial cells in salivary glands caused by radiotherapy is one of the causes of impaired gland function [45, 46]. It leads to microvascular dysfunction and the production of ceramide and reactive oxygen species (ROS) that can induce gland dysfunction. ROS scavengers are used to protect salivary gland function in radiotherapy patients [47]. By inhibiting or eliminating aberrant oxidation reactions, it is possible to reduce damage to salivary gland function caused by radiation. One study found that levels of salivary non-enzymatic antioxidants and antioxidant enzymes in the saliva secreted by the parotid gland were higher than those secreted by the submandibular salivary glands [48]. Therefore, salivary gland function changes due to parotid microvascular injury are not as serious as those caused by submandibular microvascular injury. Another group showed that the saliva-to-serum ^{131}I concentration rates in the parotid gland of mice and humans were 0.59 and 4.6, respectively, while those in the submandibular gland were 5.1 and 6.9 [49]. The ^{131}I concentration was higher in the submandibular glands of both species. An investigation showed that murine duct cells in the different salivary glands varied greatly in their ability to concentrate iodide, so it could be shown that ^{131}I was mainly concentrated in the ducts of the submandibular glands in mice, with lower levels in the parotid gland and very little in sublingual gland ducts [50]. Because of their ability to concentrate ^{131}I , the submandibular glands are more susceptible to radiation damage.

After the first ^{131}I treatment, we performed a second scintigraphy scan. We found that the males younger than 55 and females older than 55 showed a tendency of decreased function of the left parotid gland compared to the right

parotid gland. The differential changes in the left and right glands after radiation have been reported in several studies and may be due to the asymmetric concentration of radioactive iodine in the salivary glands [3, 15, 31].

4.4. Treatment Frequency and ^{131}I Dose. The Chi-square test showed that the difference between the number of damaged and undamaged salivary glands in males and females increased significantly with the increase of treatment times, and the percentage of damaged glands also increased gradually. The number of treatments was correlated with the ^{131}I cumulative dose. A correlation between radiation dose and salivary gland dysfunction was previously reported [51, 52]. A salivary glandular scintillation imaging study showed that ~30% of salivary parenchymal function was lost following a single ^{131}I dose of 6 GBq (162 mCi), with a cumulative dose of 35 GBq (945 mCi) resulting in complete loss of glandular function [53]. Parthasarathy and Crawford argued that significant side effects were rarely seen at doses <3.7 GBq (100 mCi) [54].

5. Limitations

Our results should be considered in the context of some limitations. First, this was a retrospective study with no survey to assess patient symptoms and signs, so it was not possible to add more conditions (e.g., dry mouth, difficulty swallowing, loss of taste) for case screening. Second, there was a small number of patients, especially among the group treated more than four times, which limited our analysis of salivary gland function in patients with DTC treated for more than 2–3 years. Third, clinical parameters and test data of some patients were missing. Finally, the lack of significant findings in males maybe because they only accounted for 27% of the study cohort.

6. Conclusions

This study quantitatively compared salivary scintigraphy parameters in DTC patients after multiple ^{131}I treatments. Salivary gland function sensitivities are quite variable in different ages and sexes, with the highest sensitivity in women younger than 55. After treatment, the uptake or secretion function of some salivary glands with impaired basic function increased. Decreased salivary gland function is significantly related to the number of ^{131}I treatments and the cumulative dose. The parotid glands have the most significant reduction in uptake.

Data Availability

The data generated in the study are included in this article. The database is available upon request.

Disclosure

XL and LY are co-first authors. ZM and YW are co-corresponding authors.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

XL and LY contributed equally to the study. ZM and YW contributed equally to the study.

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Supplementary Materials

Table S1 summarizes the number of patients, age distribution, and cumulative dose before one or more ^{131}I treatments. Table S2: Chi-square test was performed to determine the relationship between the number of patients by sex and the impairment of salivary gland function before each ^{131}I treatment. There was a statistical difference in the left submandibular gland injury count between sexes before the first treatment ($p < 0.05$). Figure S1: The percentage of damaged salivary glands increased with the number of treatments in both sexes. Figure S2: The cumulative dose of ^{131}I received by patients over several treatments. (*Supplementary Materials*)

References

- [1] K. D. Miller, L. Nogueira, A. B. Mariotto et al., "Cancer treatment and survivorship statistics," *CA: A Cancer Journal for Clinicians*, vol. 69, no. 5, pp. 363–385, 2019.
- [2] B. R. Haugen, E. K. Alexander, K. C. Bible et al., "2015 American thyroid association management guidelines for adult patients with thyroid nodules and differentiated thyroid cancer: the American thyroid association guidelines task force on thyroid nodules and differentiated thyroid cancer," *Thyroid*, vol. 26, no. 1, pp. 1–133, 2016.
- [3] A. Upadhyaya, P. Zhou, Z. Meng et al., "Radioprotective effect of vitamin E on salivary glands after radioiodine therapy for differentiated thyroid cancer," *Nuclear Medicine Communications*, vol. 38, no. 11, pp. 891–903, 2017.
- [4] A. Upadhyaya, Z. Meng, P. Wang et al., "Effects of first radioiodine ablation on functions of salivary glands in patients with differentiated thyroid cancer," *Medicine*, vol. 96, no. 25, p. e7164, 2017.
- [5] J.-K. Chung, H. W. Youn, J. H. Kang, H. Y. Lee, and K. W. Kang, "Sodium iodide symporter and the radioiodine treatment of thyroid carcinoma," *Nuclear Medicine and Molecular Imaging*, vol. 44, no. 1, pp. 4–14, 2010.
- [6] R. K. Badam, J. Suram, D. B. Babu et al., "Assessment of salivary gland function using salivary scintigraphy in pre and post radioactive iodine therapy in diagnosed thyroid carcinoma patients," *Journal of Clinical and Diagnostic Research: Journal of Clinical and Diagnostic Research*, vol. 10, 2016.
- [7] K. M. D. La Perle, D. C. Kim, N. C. Hall et al., "Modulation of sodium/iodide symporter expression in the salivary gland," *Thyroid*, vol. 23, no. 8, pp. 1029–1036, 2013.
- [8] C. Llana-Puy, "The rôle of saliva in maintaining oral health and as an aid to diagnosis, *Medicina Oral, Patología Oral y Cirugía Bucal*, vol. 11, no. 5, 2006.
- [9] E. Kaufman, I. B. Lamster, and I. B. Lamster, "The diagnostic applications of saliva- A review," *Critical Reviews in Oral Biology & Medicine*, vol. 13, no. 2, pp. 197–212, 2002.
- [10] M. Lenander-Lumikari and V. Loimaranta, "Saliva and dental caries," *Advances in Dental Research*, vol. 14, no. 1, pp. 40–47, 2000.
- [11] J. C. Atkinson, M. Grisius, W. Massey, and W. Massey, "Salivary hypofunction and xerostomia: diagnosis and treatment," *Dental Clinics of North America*, vol. 49, no. 2, pp. 309–326, 2005.
- [12] E. N. Klein Hesselink, A. H. Brouwers, J. R. de Jong et al., "Effects of radioiodine treatment on salivary gland function in patients with differentiated thyroid carcinoma: a prospective study," *Journal of Nuclear Medicine*, vol. 57, no. 11, pp. 1685–1691, 2016.
- [13] R. Solans, J. A. Bosch, P. Galofré et al., "Salivary and lacrimal gland dysfunction (sicca syndrome) after radioiodine therapy," *Journal of Nuclear Medicine: Official Publication, Society of Nuclear Medicine*, vol. 42, no. 5, pp. 738–743, 2001.
- [14] S. Hyer, A. Kong, B. Pratt, C. Harmer, and C. Harmer, "Salivary gland toxicity after radioiodine therapy for thyroid cancer," *Clinical Oncology*, (Royal College of Radiologists), vol. 19, no. 1, pp. 83–86, 2007.
- [15] J.-q. Wu, H.-j. Feng, W. Ouyang et al., "Systematic evaluation of salivary gland damage following I-131 therapy in differentiated thyroid cancer patients by quantitative scintigraphy and clinical follow-up," *Nuclear Medicine Communications*, vol. 36, no. 8, pp. 819–826, 2015.
- [16] E. Horvath, V. Skoknic, S. Majlis et al., "Radioiodine-induced salivary gland damage detected by ultrasonography in patients treated for papillary thyroid cancer: radioactive iodine activity and risk," *Thyroid*, vol. 30, no. 11, pp. 1646–1655, 2020.
- [17] D. Muralidharan, N. Fareed, P. V. Pradeep et al., "Qualitative and quantitative changes in saliva among patients with thyroid dysfunction prior to and following the treatment of the dysfunction," *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*, vol. 115, no. 5, pp. 617–623, 2013.
- [18] T. Gupta, C. Hotwani, S. Kannan et al., "Prospective longitudinal assessment of parotid gland function using dynamic quantitative perchneate scintigraphy and estimation of dose-response relationship of parotid-sparing radiotherapy in head-neck cancers," *Radiation Oncology*, vol. 10, no. 1, p. 67, 2015.
- [19] T. Aksoy, P. O. Kiratli, B. Erbas, and B. Erbas, "Correlations between histopathologic and scintigraphic parameters of salivary glands in patients with Sjögren's syndrome," *Clinical Rheumatology*, vol. 31, no. 9, pp. 1365–1370, 2012.
- [20] D. A. Anjos, E. C. S. C. Etchebehere, A. O. Santos et al., "Normal values of [99mTc]perchneate uptake and

- excretion fraction by major salivary glands," *Nuclear Medicine Communications*, vol. 27, no. 4, pp. 395–403, 2006.
- [21] G. B. Proctor, A. M. Shaalan, and A. M. Shaalan, "Disease-induced changes in salivary gland function and the composition of saliva," *Journal of Dental Research*, vol. 100, no. 11, pp. 1201–1209, 2021.
- [22] J. Y. Barr, X. Wang, P. A. Kreiger, S. M. Lieberman, and S. M. Lieberman, "Salivary-gland-protective regulatory T-cell dysfunction underlies female-specific sialadenitis in the non-obese diabetic mouse model of Sjögren syndrome," *Immunology*, vol. 155, no. 2, pp. 225–237, 2018.
- [23] Y. Q. Zhang, X. Ye, Y. Meng, Y. N. Zhao, D. G. Liu, and G. Y. Yu, "Evaluation of parotid gland function before and after endoscopy-assisted stone removal," *Journal of Oral and Maxillofacial Surgery: Official Journal of the American Association of Oral and Maxillofacial Surgeons*, vol. 77, no. 2, 2019.
- [24] A. Chern, A. O. Famuyide, G. Moonis, A. K. Lalwani, and A. K. Lalwani, "Sialadenitis: a possible early manifestation of covid-19," *The Laryngoscope*, vol. 130, no. 11, pp. 2595–2597, 2020.
- [25] M. Knaś, M. Maciejczyk, K. Sawicka et al., "Impact of morbid obesity and bariatric surgery on antioxidant/oxidant balance of the unstimulated and stimulated human saliva," *Journal of Oral Pathology & Medicine: Official Publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*, vol. 45, no. 6, pp. 455–464, 2016.
- [26] M. Maciejczyk, J. Matczuk, M. Żendzian-Piotrowska et al., "Eight-week consumption of high-sucrose diet has a pro-oxidant effect and alters the function of the salivary glands of rats," *Nutrients*, vol. 10, no. 10, 2018.
- [27] J.-S. Choi, I. S. Park, S.-k. Kim, J.-Y. Lim, Y. M. Kim, and Y.-M. Kim, "Analysis of age-related changes in the functional morphologies of salivary glands in mice," *Archives of Oral Biology*, vol. 58, no. 11, pp. 1635–1642, 2013.
- [28] W. Pedersen, M. Schubert, K. Izutsu, T. Mersai, E. Truelove, and E. Truelove, "Clinical science age-dependent decreases in human submandibular gland flow rates as measured under resting and post-stimulation conditions," *Journal of Dental Research*, vol. 64, no. 5, pp. 822–825, 1985.
- [29] L. Lin and L. Sijin, "Clinical guidelines for 131I treatment of Graves' hyperthyroidism (2021 edition)," *Chinese Society of Nuclear Medicine*, vol. 41, no. 04, pp. 242–253, 2021, (in Chinese).
- [30] B. Fallahi, D. Beiki, S. M. Abedi et al., "Does vitamin E protect salivary glands from I-131 radiation damage in patients with thyroid cancer?" *Nuclear Medicine Communications*, vol. 34, no. 8, pp. 777–786, 2013.
- [31] Y.-S. An, J.-K. Yoon, S. J. Lee et al., "Symptomatic late-onset sialadenitis after radioiodine therapy in thyroid cancer," *Annals of Nuclear Medicine*, vol. 27, no. 4, pp. 386–391, 2013.
- [32] K. Shizukuishi, S. Nagaoka, Y. Kinno et al., "Scoring analysis of salivary gland scintigraphy in patients with Sjögren's syndrome," *Annals of Nuclear Medicine*, vol. 17, no. 8, pp. 627–631, 2003.
- [33] J. M. Roesink, M. A. Moerland, A. Hoekstra, P. P. V. Rijk, C. H. Terhaard, and C. H. J. Terhaard, "Scintigraphic assessment of early and late parotid gland function after radiotherapy for head-and-neck cancer: a prospective study of dose-volume response relationships," *International Journal of Radiation Oncology, Biology, Physics*, vol. 58, no. 5, pp. 1451–1460, 2004.
- [34] G. Sunavala-Dossabhoy, "Radioactive iodine: an unappreciated threat to salivary gland function," *Oral Diseases*, vol. 24, no. 1-2, pp. 198–201, 2018.
- [35] M. Caglar, M. Tuncel, R. Alpar, and R. Alpar, "Scintigraphic evaluation of salivary gland dysfunction in patients with thyroid cancer after radioiodine treatment," *Clinical Nuclear Medicine*, vol. 27, no. 11, pp. 767–771, 2002.
- [36] K. S. Jo, Y.-S. An, S. J. Lee et al., "Significance of salivary gland radioiodine retention on post-ablation 131I scintigraphy as a predictor of salivary gland dysfunction in patients with differentiated thyroid carcinoma," *Nuclear Medicine and Molecular Imaging*, vol. 48, no. 3, pp. 203–211, 2014.
- [37] B. Liu, R. Huang, A. Kuang et al., "Iodine kinetics and dosimetry in the salivary glands during repeated courses of radioiodine therapy for thyroid cancer," *Medical Physics*, vol. 38, no. 10, pp. 5412–5419, 2011.
- [38] J. P. Almeida, Á. E. Sanabria, E. N. P. Lima, L. P. Kowalski, and L. P. Kowalski, "Late side effects of radioactive iodine on salivary gland function in patients with thyroid cancer," *Head & Neck*, vol. 33, no. 5, pp. 686–690, 2011.
- [39] R. Levi-Montalcini and S. Cohen, "Effects of the extract of the mouse submaxillary salivary glands on the sympathetic system of mammals," *Annals of the New York Academy of Sciences*, vol. 85, pp. 324–341, 1960.
- [40] R. L. Byyny, D. N. Orth, S. Cohen, and E. S. Doayne, "Epidermal growth factor: effects of androgens and adrenergic agents," *Endocrinology*, vol. 95, no. 3, pp. 776–82, 1974.
- [41] D. N. Ishii and E. M. Shooter, "Regulation of nerve growth factor synthesis in mouse submaxillary glands by testosterone," *Journal of Neurochemistry*, vol. 25, no. 68, pp. 43–51, 1975.
- [42] J. E. Brandt, R. Priori, G. Valesini, and D. Fairweather, "Sex differences in Sjögren's syndrome: a comprehensive review of immune mechanisms," *Biology of Sex Differences*, vol. 6, p. 19, 2015.
- [43] F. R. Burlage, H. Faber, H. H. Kampinga, J. A. Langendijk, A. Vissink, and R. P. Coppes, "Enhanced proliferation of acinar and progenitor cells by prophylactic pilocarpine treatment underlies the observed amelioration of radiation injury to parotid glands," *Radiotherapy & Oncology*, vol. 90, no. 2, pp. 253–256, 2009.
- [44] J. Denekamp, "Cell kinetics and radiation biology," *International Journal of Radiation Biology & Related Studies in Physics, Chemistry & Medicine*, vol. 49, no. 2, pp. 357–380, 1986.
- [45] A. P. Cotrim, A. Sowers, J. B. Mitchell, B. J. Baum, and B. J. Baum, "Prevention of irradiation-induced salivary hypofunction by microvessel protection in mouse salivary glands," *Molecular Therapy*, vol. 15, no. 12, pp. 2101–2106, 2007.
- [46] Z. Zhang, F. Cui, C. Cao, Q. Wang, and Q. Zou, "Single-cell RNA analysis reveals the potential risk of organ-specific cell types vulnerable to SARS-CoV-2 infections," *Computers in Biology and Medicine*, vol. 140, Article ID 105092, 2022.
- [47] A. Mizrachi, A. P. Cotrim, N. Katabi et al., "Radiation-induced microvascular injury as a mechanism of salivary gland hypofunction and potential target for radioprotectors," *Radiation Research*, vol. 186, no. 2, pp. 189–195, 2016.
- [48] K. Blochowiak and H. Witmanowski, "Ocena stężenia produktów peroksydacji lipidów w ślinie i w surowicy krwi u pacjentów ze złamaniami żuchwy," *Czasopismo Stomatologiczne*, vol. 63, no. 4, pp. 250–258, 2010.
- [49] B. Cohen, N. B. Myant, and N. B. Myant, "Concentration of salivary iodide: a comparative study," *The Journal of Physiology*, vol. 145, no. 3, pp. 595–610, 1959.
- [50] J. H. Logothetopoulos, N. B. Myant, and N. B. Myant, "Concentration of radio-iodide and 35-S-thiocyanate by the

- salivary glands,” *The Journal of Physiology*, vol. 134, no. 1, pp. 189–194, 1956.
- [51] R. Solans, J. A. Bosch, P. Galofré et al., “Salivary and lacrimal gland dysfunction (Sicca syndrome) after radioiodine therapy,” *Journal of Nuclear Medicine: Official Publication, Society of Nuclear Medicine*, vol. 42, pp. 738–743, 2001.
- [52] C. Alexander, J. B. Bader, A. Schaefer, C. Finke, and C. M Kirsch, “Intermediate and long-term side effects of high-dose radioiodine therapy for thyroid carcinoma,” *Journal of Nuclear Medicine: Official Publication, Society of Nuclear Medicine*, vol. 39, no. 9, pp. 1551–1554, 1998.
- [53] W. Jentzen, E. Schneider, L. Freudenberg et al., “Relationship between cumulative radiation dose and salivary gland uptake associated with radioiodine therapy of thyroid cancer,” *Nuclear Medicine Communications*, vol. 27, no. 8, pp. 669–676, 2006.
- [54] K. L. Parthasarathy and E. S. Crawford, “Treatment of thyroid carcinoma: emphasis on high-dose ¹³¹I outpatient therapy,” *Journal of Nuclear Medicine Technology*, vol. 30, no. 4, pp. 165–173, 2002, quiz 172-3.