

The published manuscript is available at EurekaSelect via
<https://www.eurekaselect.com/openurl/content.php?genre=article&doi=10.2174/1381612826666200513112607>.

Curcumin for the management of periodontal diseases: a review

Abstract:

Periodontal disease is one of the most common cause of tooth loss among adults. Research shows that inflammation is one of the crucial components in the initiation and progression of periodontitis. Various herbal medicines have recently been receiving attention for the management of periodontitis owing to their general safety and multitude of pharmacological actions. Curcumin, a bioactive polyphenol extracted from *Curcuma longa*, possesses antioxidant, antimicrobial, antiinflammatory, analgesic and anticarcinogenic properties. Several studies have assessed the usefulness of curcumin against periodontal diseases and the results have shown equivalent or even higher efficacy compared to the regularly used drugs for the management of periodontitis such as chlorhexidine. Herein, we review the experimental and clinical findings on the anti-periodontitis effects of curcumin and the pharmacological mechanisms underlying these effects.

Keywords: Inflammation; Gingivitis; Herbal medicine; Curcumin; Periodontitis; Herbal mouthwash

Introduction

Poor oral/dental health causes significant pain and suffering and therefore affects general health (1). Loss of teeth due to periodontitis often causes discomfort and endangers the esthetics and function. Studies have revealed a possible link between systemic health problems and periodontitis (2, 3). More than **one-half** in adults are affected by periodontal diseases (4-8). There is a recent worldwide increase in the use of medicinal plants in the management of various conditions including periodontal diseases due to their efficacy and favorable side effect profile (9).

Literature search

In this review, we searched manuscripts with the following keywords: antioxidant capacity, anti-inflammatory, gingivitis, herbal medicine, curcumin, periodontitis, herbal mouthwashes, alternative medicine. The search databases include Google Scholar, PubMed, Web of Science, Scopus from 1990 - 2019 using the EndNote software.

Periodontal disease

Periodontal disease is a set of inflammatory conditions affecting the gingiva and the supporting structures of the periodontium(10, 11). Periodontal diseases are classified into gingival diseases and periodontitis (12). The gingival disease is usually being characterized by inflammatory dental plaque accumulation in the gingival tissues. The clinical presentation of the gingival disease includes areas of swelling, redness and bleeding. **Periodontal ligament and alveolar bone are not affected** (13, 14). Gingivitis means inflammation of the gingiva which can lead to

periodontitis (15, 16). Periodontitis is an condition where there is periodontal tissue inflammation resulting in alveolar bone destruction (13, 17).

The clinical signs of periodontitis include changes in the gingival tissue morphology, gingival overgrowth is one of the modifications that occurs in chronic periodontitis may be detectable clinically. Polymorphonuclear leukocytes and monocytes pass through the subepithelial connective tissue through the junctional epithelium and into the gingival sulcus (18). Bleeding upon probing and periodontal pocket formation that lead to tooth loss if untreated (19). The periodontal pocket facilitates bacterial colonization and subgingival plaque formation (13, 17) (Figure 1).

Various non-surgical interventions such as application of numerous antimicrobials and chemotherapeutic agents including chlorhexidine, triclosan, cetylpyridinium chloride have been tried for the management of periodontal diseases. Some cases will need surgical management. Since the etiology of periodontitis is multifactorial and complex etiologies management of periodontitis can be challenging (20) (21).

Herbal medicine strategy

Herbal medicines were used to manage various conditions since time immemorial (22). Since natural medicines have potentially fewer side effects and lower costs than synthetic drugs, the use of phytopharmaceuticals has become widespread worldwide. Natural remedies have been shown to have antioxidant, antiseptic and anti-inflammatory effects (21, 23) which can be potentially beneficial in preventing and treating periodontal diseases.

Curcumin

Curcumin (diferuloylmethane) is a natural compound obtained from *Curcuma longa* (24, 25). Besides to its extensive culinary use, curcumin has been used for centuries for its medicinal properties to manage various conditions (26-28) (29, 30). Curcumin has been shown to have antioxidant (31, 32), antimicrobial (33), anti-inflammatory (34-37), analgesic (38), antimicrobial (33) and anticarcinogenic and chemosensitizing properties (39-42). Curcumin can be safely consumed up to 8g per day (43). Various formulations of curcumin are available including oral and topical formulations (44, 45). We have performed a comprehensive review of literature on the usefulness of curcumin in periodontitis.

Effect of curcumin on experimental models of periodontitis

One study aimed to assess the effect of resveratrol and curcumin on the progression of experimental periodontitis in rats. Intergroup comparisons showed higher bone-loss in the placebo group compared to the active treatment groups (46). Gu et al (47), evaluated the effect of orally administration of 4-methoxycarbonylcurcumin (CMC 2.5) for 3 weeks in Streptozotocin (STZ) induced periodontal disease. The results showed that CMC 2.5 reduced inflammatory markers including Matrix metalloproteinase 9 (MMP-9), Matrix metalloproteinase 13 (MMP-13) and Interleukin 1 beta (IL-1 β). Daily intragastric administration of curcumin inhibits the expression of Interleukin 6 (IL-6), tumor necrosis factor alpha (TNF- α), Prostaglandin E2 (PGE2), cyclooxygenase-2 (COX-2) in the gingival tissues in ligature-induced periodontal disease in the rat. There was also a significant reduction of inflammatory cell infiltrate and increased collagen content and fibroblastic cell numbers in the curcumin-treated animals (48). Daily curcumin administration to rats inhibited the inflammatory markers in the gingival tissues

(49). Besides, curcumin treatment resulted in a significant increase in collagen and fibroblasts. Curcumin (5–30 $\mu\text{mol/L}$) dose-dependently inhibited COX-2 mRNA and protein synthesis in *P. gingivalis* LPS -stimulated human gingival fibroblasts (HGFs) (50).

Mau et al(51), showed that curcumin reduced receptor activator of nuclear factor- κB ligand (RANKL)-induced osteoclast differentiation and the expression of osteoclastic specific genes in a dose-dependent manner. Curcumin reduced tartrate-resistant acid phosphatase (TRAP)-positive and polymorphonuclear cells infiltration and alveolar bone destruction in ligature-induced experimental periodontitis model. Local administration of nanocurcumin reduced inflammatory bone resorption of hemi-maxillae in a lipopolysaccharides (LPS)-induced model of periodontal disease (52).

Administration of curcumin significantly reduced bone resorption in mandibles and RANKL, receptor activator of nuclear factor- κB (RANK) and osteoprotegerin (OPG) in mandibles in rat ligature-induced periodontitis. It also reduced the inflammatory cytokine expression levels in gingival tissues of the experimental periodontitis animals (53). Application of 2% curcumin gel lead to statistically significant reduction in the probing pocket depth (PPD) and gingival index (GI). GI was determined and recorded at 4 gingival sites per tooth according to the following criteria: (0) normal gingiva, (1) mild gingivitis without bleeding on probing, (2) moderate gingivitis with bleeding on probing, and (3) severe gingivitis with ulceration and spontaneous bleeding. The sum of the scores from the four areas of each tooth was divided by 4 to derive the GI for that tooth. The average GI value obtained after calculating individual GI (54).

There was no significant difference in the morphometric analysis at mesiobuccal, midbuccal, distobuccal, mesiopalatal, midpalatal, and distopalatal site following administration of curcumin gel in an experimental periodontitis model of rat (55). Systemic administration of curcumin and piperine increased Transforming growth factor beta (TGF- β) levels, reduced Nuclear factor- κ B (NF- κ B) activation and reduced cellular infiltrate, associated with enhanced collagen content and accelerated soft tissue repair in ligature-induced periodontitis. Although, only curcumin increased early bone repair in ligature-induced periodontitis (56).

Effects of curcumin on periodontal diseases in clinical studies

In a randomized study the efficacy of curcumin was compared to chlorhexidine in the management of chronic periodontitis. There was a greater reduction of clinical attachment level (CAL) and PPD after curcumin than chlorhexidine after 30 days (57). In another randomized clinical study (split-mouth design) administration of curcumin gel (10 mg *curcuma longa* extract/gram) significantly decreased plaque index (PI), GI, probing depth (PD) and clinical attachment loss after 45 days (58). In a single-blind, randomized study, PI, GI and saliva collection for ROM (whole oxidant capacity of saliva) in three groups (control (saline), curcumin mouth rinse and chlorhexidine mouth rinse groups) were performed. The curcumin group showed a significant reduction in ROM contents after 4 weeks (59). In patients with chronic periodontitis application of curcumin gel (1 mg/ml) along with scaling and root planing lead to reduction in the number of periopathogens. There was also an improvement in various clinical parameters including PPD, CAL, PI and bleeding index (BI) after 6 months compared to the control group which received only scaling and root planing (60). In a double-blinded randomized

study, the efficacy of curcumin and chlorhexidine mouth rinses on PI, GI and sulcus bleeding index (SBI) in gingivitis patients was evaluated. There was a clinically significant reduction in all parameters in curcumin and chlorhexidine groups (61). In another pilot randomized study, the efficacy of 1% curcumin solution as an adjunct to thorough scaling and root planing in patients with chronic periodontitis was evaluated. There was a significant improvement in BOP, redness, plaque index and PPD in the curcumin group after one month compared to chlorhexidine gluconate and a positive control (saline) group. The microbiological analysis showed significant reduction in the number of BANA positive sites in the curcumin group after 1 month and by the end of study period (62). Application of curcumin collagen sponge or chlorhexidine (CHX) chips (Periocol-CG) for chronic periodontitis leads to a significant reduction in PI and GI scores at the end of the 6-month study period. There was also a significant improvement in the microbiological parameters, PPD and CAL levels in both groups (63).

Another study investigated the effect of curcumin and doxycycline in the inhibition of **Matrix metalloproteinase 9 (MMP-9)** activity in gingival tissue samples from chronic periodontitis patients. Curcumin demonstrated 61.01% reduction in the MMP-9 activity at 1500 µg/ml concentration and doxycycline demonstrated 59.58% decrease in the MMP-9 activity at 300 µg/ml concentration (64). One study evaluated the efficacy of subgingival application of 0.2% chlorhexidine gel and 2% curcumin gel as an adjunct to scaling and root planing in the management of mild to moderate periodontal pockets. Both agents had an impact on mild to moderate periodontal pockets, but the curcumin gel was more efficient than the chlorhexidine gel (65). The effects of curcumin and chlorhexidine mouth-washes was assessed in three groups

(patients who underwent scaling and root planing followed by the use of curcumin mouthwash, patients underwent scaling and root planing followed by the use of chlorhexidine mouthwash, patients underwent only scaling and root planing). When compared to the scaling and root planing group there was a significant improvement in clinical parameters in the other groups (66). A randomized single-blinded (split-mouth) study was conducted to assess the effect of curcumin as an adjunct to scaling and root planing in patients with chronic periodontitis. The results showed a decrease in the CAL, probing depth, PI, GI, and microbiologic parameters (*Porphyromonas gingivalis*, *Tannerella forsythia*, and *Treponema denticola*) after curcumin gel application (67). *In-situ* gel formulations of curcumin containing 2% curcumin significantly reduced the PD, BI, and to a lesser extent of PI after one-month treatment in patients with chronic periodontitis (68). A randomized, double-blinded, parallel study compared the IL-1 β and CCL28 levels after application of curcumin extract, chlorhexidine and chlorhexidine-metronidazole and metronidazole in an experimental gingivitis human model. The increase of IL-1 β and CCL28 in the curcumin and chlorhexidine-metronidazole groups was significantly less than that of the chlorhexidine alone group (69). In a single-blind, randomized study, the periodontitis patients received scaling and root planing after which test either curcumin gel (*C. longa* extract-10 mg) or ornidazole were injected. Administration of curcumin gel significantly reduced PD, PI and clinical attachment loss than ornidazole group after 1 month in these patients with chronic periodontitis (70).

Potential mechanisms for the anti-periodontitis effects

The hallmark of periodontal disease is inflammation and bone loss (71). Bone remodeling maintains the integrity of the skeleton by formation of bone via osteoblasts and removal of mineralized bone by osteoclasts. The RANKL and its OPG receptor play an important role in the bone remodeling regulation (72). RANKL and OPG have an imperative role in the healing of destructive periodontal disease (73, 74). RANKL is expressed by osteoblasts, fibroblasts, chondrocytes, activated T cells and B cells stromal cells as well as other mesenchymal cells (75, 76). Curcumin suppresses the RANKL/RANK/OPG expression thereby inhibiting the inflammatory response and bone loss during experimental periodontitis (53). The NF- κ B activation increases the expression of various inflammatory cytokines and chemokines involved in the pathogenesis of various inflammatory diseases (77). NF- κ B was activated when oral epithelial cells were exposed to *Porphyromonas gingivalis* and *Fusobacterium nucleatum* which are periodontopathogens that induced apoptosis of monocytes and neutrophils (78-80).

The two main signaling pathways activated downstream of toll like receptor 4 (TLR4) are p38 MAPK and NF- κ B. They are considered as the ‘general indicators of inflammatory activity’ (81). TLRs recognize and respond to various types of microbial challenges. Activation of TLR4 enhances the activation of mitogen-activated protein kinases and the translocation of nuclear NF- κ B (82). Bacterial infection elicits an inflammatory response that will eventually exacerbate bone destruction (83). The RANK ligand-induced osteoclastogenesis is mediated by NF- κ B. NF- κ B pathway inhibition results in inhibition of osteoclast formation as well as bone resorptive activity (84).

There is a close relationship between reactive oxygen species (ROS) and periodontitis (85-89). ROS have been implicated in inducing oxidative damage to pathogens (90, 91). However, overproduction of ROS can result in oxidative damage which is strongly associated with periodontal destruction (17, 92).

The dental plaque harbors several pathogens which stimulate the release a number of inflammatory cytokines including TNF- α and interleukins. These cytokines attract polymorphonuclear cells (PMNs) to the infection site. In response to this bacterial challenge, PMN secretes a variety of proteolytic enzymes and an increase in O₂ production (93). ROS can aggravate inflammatory injury through NF- κ B (94, 95) and induce apoptosis through c-Jun N-terminal kinase (JNK) activation (96). A reduction in ROS levels has shown to reduce bone loss (97).

The biological mechanisms of curcumin-mediated effects involve regulation of several molecular targets, including protein kinases, cytokines, growth factors, transcription factors and other enzymes such as cyclooxygenase-2. Curcumin inhibits the NF- κ B activation pathway, reduces the synthesis of COX-2 (50), and inhibits the signaling of TLR4 (98) Curcumin inhibits the expression of inflammatory cytokines (99-102). This evidence suggests that curcumin potentially ameliorates the initial stages of periodontitis (50). Curcumin promotes healing of wound by migration of fibroblasts (103, 104).

Advances in the formulations of curcumin

Curcumin can be used in various formulations including soaps, cosmetics, capsules, ointments, energy drinks and capsules (44). The therapeutic use of curcumin is limited due to its rapid

metabolism and low solubility resulting in poor bioavailability (105).(106-108). Various formulations such as micelles, nanoparticles, liposomal vesicles, phospholipid complexes, nano-emulsions and polymers are being tried to improve the efficiency of curcumin delivery (109). Synthesis of curcumin derivative with a carbonyl substituent at the C-4 position increases its anti-inflammatory therapeutic properties due to the presence of an additional electron-withdrawing group. 4-methoxycarbonylcurcumin has a methoxycarbonyl group at C4 which improves its solubility, acidity as well as greater albumin and zinc-binding capacities. This modification potentiates the MMP-inhibitory effects of 4-methoxycarbonylcurcumin compared to curcumin (110, 111). (47). Zambrano et al. (52) found that nanoparticles synthesized from poly(lactic acid and co-glycolic acid) increased the half-life of curcumin by 15-fold in rats (112). These nanoparticle formulations allow for chemical modifications that may increase its absorption (113). In patients with chronic periodontitis *in-situ* gel-forming formulations will form strong gels after application at the delivery site thereby increasing the duration of contact of active formulation in the site (68, 114). Encapsulated curcumin in nanoparticles, enhanced the water solubility of curcumin. Conversely, free curcumin exhibited a better photodynamic property than curcumin -nanoparticle(115).

Conclusion

Periodontal disease remission/control is characterized by a significant inflammation reduction, some improvement in other clinical parameters, and a stabilization of disease progression. Ideally, restoration to periodontal stability should be considered a major therapeutic goal and can be achieved by controlling inflammation and infection, decreasing predisposing factors, and

controlling modifying factors. Curcumin is an effective and safe alternative to several common medications and has a multitude of therapeutic benefits in various diseases. Curcumin can counterbalance periodontal inflammation, oxidative stress and dental destruction. There is growing evidence that curcumin show equivalent or even higher efficacy compared with the regularly used medications for the management of periodontitis such as chlorhexidine. Curcumin analogs have also been investigated in the management of periodontitis. Therefore, the development of chemically modified curcumin analogs with improved anti-inflammatory and anti-periodontitis effects is an ongoing attempt that could lead to the introduction of novel anti-periodontitis drug candidates in the future.

Conflict of interest: None.

References

1. Sheiham A. Oral health, general health and quality of life. *SciELO Public Health*; 2005.
2. Manjunath B, Praveen K, Chandrashekar B, Vatchala Rani R, Bhalla A. Periodontal infections: a risk factor for various systemic diseases. *National Medical Journal of India*. 2011;24(4):214.
3. Shekar BRC, Nagarajappa R, Suma S, Thakur R. Herbal extracts in oral health care-A review of the current scenario and its future needs. *Pharmacognosy reviews*. 2015;9(18):87.
4. Petersen PE, Ogawa H. The global burden of periodontal disease: towards integration with chronic disease prevention and control. *Periodontology* 2000. 2012;60(1):15-39.

5. Najafi MH, Taheri M, Mokhtari MR, Forouzanfar A, Farazi F, Mirzaee M, et al. Comparative study of 0.2% and 0.12% digluconate chlorhexidine mouth rinses on the level of dental staining and gingival indices. *Dental research journal*. 2012;9(3):305.
6. Ghanbari H, Forouzanfar A, Fatemi K, Mokhtari MR, Abrishami M, Ebrahiminik Z, et al. Modified Widman flap procedure: With or without periodontal dressing? *Open Journal of Stomatology*. 2012;2(03):170.
7. Forouzanfar A. CLINICAL APPLICATION OF ANTIBIOTICS FOR THE MANAGEMENT OF PERIODONTAL DISEASE, A SYSTEMATIC.
8. Eke PI, Thornton-Evans GO, Wei L, Borgnakke WS, Dye BA, Genco RJ. Periodontitis in US adults: National health and nutrition examination survey 2009-2014. *The Journal of the American Dental Association*. 2018;149(7):576-88. e6.
9. Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. *Frontiers in pharmacology*. 2014;4:177.
10. Cowan LT, Lakshminarayan K, Lutsey PL, Folsom AR, Beck J, Offenbacher S, et al. Periodontal disease and incident venous thromboembolism: The Atherosclerosis Risk in Communities study. *Journal of clinical periodontology*. 2019;46(1):12-9.
11. Linden GJ, McClean KM, Woodside JV, Patterson CC, Evans A, Young IS, et al. Antioxidants and periodontitis in 60–70-year-old men. *Journal of clinical periodontology*. 2009;36(10):843-9.
12. Wiebe CB, Putnins EE. The periodontal disease classification system of the American Academy of Periodontology-an update. *JOURNAL-CANADIAN DENTAL ASSOCIATION*. 2000;66(11):594-9.
13. Williams RO, Feldmann M, Maini RN. Anti-tumor necrosis factor ameliorates joint disease in murine collagen-induced arthritis. *Proceedings of the National Academy of Sciences*. 1992;89(20):9784-8.
14. Moghaddam MA, Salehinejad J, Mokhtari MR, Mohammadipour HS, Forouzanfar A. PALATAL PERIPHERAL OSSIFYING FIBROMA ALONG WITH GENERALIZED MARGINAL GINGIVITIS: A CASE REPORT.
15. Offenbacher S. Periodontal diseases: pathogenesis. *Annals of periodontology*. 1996;1(1):821-78.

16. Valkenburg C, Van der Weijden FA, Slot DE. Plaque control and reduction of gingivitis: The evidence for dentifrices. *Periodontology* 2000. 2019;79(1):221-32.
17. Waddington R, Moseley R, Embery G. Periodontal Disease Mechanisms: Reactive oxygen species: a potential role in the pathogenesis of periodontal diseases. *Oral diseases*. 2000;6(3):138-51.
18. Muñoz-Carrillo JL, Hernández-Reyes VE, García-Huerta OE, Chávez-Ruvalcaba F, Chávez-Ruvalcaba MI, Chávez-Ruvalcaba KM, et al. Pathogenesis of Periodontal Disease. *Periodontal Disease-Diagnose Considerations*: IntechOpen; 2019.
19. Pihlstrom BL, Michalowicz BS, Johnson NW. Periodontal diseases. *The lancet*. 2005;366(9499):1809-20.
20. Ramesh A, Varghese SS, Doraiswamy JN, Malaiappan S. Herbs as an antioxidant arsenal for periodontal diseases. *Journal of intercultural ethnopharmacology*. 2016;5(1):92.
21. Nugala B, Namasi A, Emmadi P, Krishna PM. Role of green tea as an antioxidant in periodontal disease: The Asian paradox. *Journal of Indian Society of Periodontology*. 2012;16(3):313.
22. Manju K, Jat R, Anju G. A review on medicinal plants used as a source of anticancer agents. *International Journal of Drug Research and Technology*. 2017;2(2):6.
23. Chapple IL. Oxidative stress, nutrition and neutrogenomics in periodontal health and disease. *International journal of dental hygiene*. 2006;4:15-21.
24. Pulido-Moran M, Moreno-Fernandez J, Ramirez-Tortosa C, Ramirez-Tortosa M. Curcumin and health. *Molecules*. 2016;21(3):264.
25. Forouzanfar F, Barreto G, Majeed M, Sahebkar A. Modulatory effects of curcumin on heat shock proteins in cancer: A promising therapeutic approach. *BioFactors* (Oxford, England). 2019.
26. Mantzorou M, Pavlidou E, Vasios G, Tsagalioti E, Giaginis C. Effects of curcumin consumption on human chronic diseases: a narrative review of the most recent clinical data. *Phytotherapy research*. 2018;32(6):957-75.
27. Marchiani A, Rozzo C, Fadda A, Delogu G, Ruzza P. Curcumin and curcumin-like molecules: from spice to drugs. *Current medicinal chemistry*. 2014;21(2):204-22.

28. Mhillaj E, Tarozzi A, Pruccoli L, Cuomo V, Trabace L, Mancuso C. Curcumin and Heme Oxygenase: Neuroprotection and Beyond. *International journal of molecular sciences*. 2019;20(10):2419.
29. Yang M, Akbar U, Mohan C. Curcumin in Autoimmune and Rheumatic Diseases. *Nutrients*. 2019;11(5):1004.
30. Lelli D, Sahebkar A, Johnston TP, Pedone C. Curcumin use in pulmonary diseases: State of the art and future perspectives. *Pharmacological research*. 2017;115:133-48.
31. Menon VP, Sudheer AR. Antioxidant and anti-inflammatory properties of curcumin. The molecular targets and therapeutic uses of curcumin in health and disease: Springer; 2007. p. 105-25.
32. Panahi Y, Khalili N, Sahebi E, Namazi S, Karimian MS, Majeed M, et al. Antioxidant effects of curcuminoids in patients with type 2 diabetes mellitus: a randomized controlled trial. *Inflammopharmacology*. 2017;25(1):25-31.
33. De R, Kundu P, Swarnakar S, Ramamurthy T, Chowdhury A, Nair GB, et al. Antimicrobial activity of curcumin against *Helicobacter pylori* isolates from India and during infections in mice. *Antimicrobial agents and chemotherapy*. 2009;53(4):1592-7.
34. Chainani-Wu N. Safety and anti-inflammatory activity of curcumin: a component of tumeric (*Curcuma longa*). *The Journal of Alternative & Complementary Medicine*. 2003;9(1):161-8.
35. Sahebkar A, Cicero AFG, Simental-Mendia LE, Aggarwal BB, Gupta SC. Curcumin downregulates human tumor necrosis factor-alpha levels: A systematic review and meta-analysis of randomized controlled trials. *Pharmacol Res*. 2016;107:234-42.
36. Abdollahi E, Momtazi AA, Johnston TP, Sahebkar A. Therapeutic effects of curcumin in inflammatory and immune-mediated diseases: A nature-made jack-of-all-trades? *Journal of Cellular Physiology*. 2018;233(2):830-48.
37. Karimian MS, Pirro M, Majeed M, Sahebkar A. Curcumin as a natural regulator of monocyte chemoattractant protein-1. *Cytokine and Growth Factor Reviews*. 2017;33:55-63.
38. Matsushita Y, Ueda H. Curcumin blocks chronic morphine analgesic tolerance and brain-derived neurotrophic factor upregulation. *Neuroreport*. 2009;20(1):63-8.

39. Park J, Contreas CN. Anti-carcinogenic properties of curcumin on colorectal cancer. *World journal of gastrointestinal oncology*. 2010;2(4):169.
40. Mirzaei H, Masoudifar A, Sahebkar A, Zare N, Nahand JS, Rashidi B, et al. MicroRNA: A novel target of curcumin in cancer therapy. *Journal of Cellular Physiology*. 2017;233(4):3004-15.
41. Rezaee R, Momtazi AA, Monemi A, Sahebkar A. Curcumin: A potentially powerful tool to reverse cisplatin-induced toxicity. *Pharmacological Research*. 2017;117:218-27.
42. Teymouri M, Pirro M, Johnston TP, Sahebkar A. Curcumin as a multifaceted compound against human papilloma virus infection and cervical cancers: A review of chemistry, cellular, molecular, and preclinical features. *BioFactors (Oxford, England)*. 2017;43(3):331-46.
43. Mirzaei H, Shakeri A, Rashidi B, Jalili A, Banikazemi Z, Sahebkar A. Phytosomal curcumin: A review of pharmacokinetic, experimental and clinical studies. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*. 2017;85:102-12.
44. Gupta SC, Patchva S, Aggarwal BB. Therapeutic roles of curcumin: lessons learned from clinical trials. *The AAPS journal*. 2013;15(1):195-218.
45. Hewlings SJ, Kalman DS. Curcumin: A Review of Its' Effects on Human Health. *Foods*. 2017;6(10):92.
46. Corrêa M, Pires P, Ribeiro F, Pimentel S, Casarin R, Cirano F, et al. Systemic treatment with resveratrol and/or curcumin reduces the progression of experimental periodontitis in rats. *Journal of periodontal research*. 2017;52(2):201-9.
47. Gu Y, Lee H-M, Napolitano N, Clemens M, Zhang Y, Sorsa T, et al. 4-methoxycarbonyl curcumin: a unique inhibitor of both inflammatory mediators and periodontal inflammation. *Mediators of inflammation*. 2013;2013.
48. Guimarães MR, Coimbra LS, de Aquino SG, Spolidorio LC, Kirkwood KL, Rossa Jr C. Potent anti-inflammatory effects of systemically administered curcumin modulate periodontal disease in vivo. *Journal of periodontal research*. 2011;46(2):269-79.
49. Guimarães MR, de Aquino SG, Coimbra LS, Spolidorio LC, Kirkwood KL, Rossa Jr C. Curcumin modulates the immune response associated with LPS-induced periodontal disease in rats. *Innate immunity*. 2012;18(1):155-63.

50. Hu P, Huang P, Chen MW. Curcumin attenuates cyclooxygenase-2 expression via inhibition of the NF- κ B pathway in lipopolysaccharide-stimulated human gingival fibroblasts. *Cell biology international*. 2013;37(5):443-8.
51. Mau L-P, Cheng W-C, Chen J-K, Shieh Y-S, Cochran DL, Huang R-Y. Curcumin ameliorates alveolar bone destruction of experimental periodontitis by modulating osteoclast differentiation, activation and function. *Journal of functional foods*. 2016;22:243-56.
52. Zambrano LM, Brandao DA, Rocha FR, Marsiglio RP, Longo IB, Primo FL, et al. Local administration of curcumin-loaded nanoparticles effectively inhibits inflammation and bone resorption associated with experimental periodontal disease. *Scientific reports*. 2018;8.
53. Zhou T, Chen D, Li Q, Sun X, Song Y, Wang C. Curcumin inhibits inflammatory response and bone loss during experimental periodontitis in rats. *Acta Odontologica Scandinavica*. 2013;71(2):349-56.
54. L e H. The gingival index, the plaque index and the retention index systems. *The Journal of Periodontology*. 1967;38(6):610-6.
55. Hosadurga RR, Rao S, Jose J, Rompicharla NC, Shakil M, Shashidhara R. Evaluation of the efficacy of 2% curcumin gel in the treatment of experimental periodontitis. *Pharmacognosy research*. 2014;6(4):326.
56. Guimaraes-Stabili MR, de Aquino SG, de Almeida Curylofo F, Tasso CO, Rocha FRG, de Medeiros MC, et al. Systemic administration of curcumin or piperine enhances the periodontal repair: a preliminary study in rats. *Clinical oral investigations*. 2019;23(8):3297-306.
57. Anitha V, Rajesh P, Shanmugam M, Priya BM, Prabhu S, Shivakumar V. Comparative evaluation of natural curcumin and synthetic chlorhexidine in the management of chronic periodontitis as a local drug delivery: A clinical and microbiological study. *Indian Journal of Dental Research*. 2015;26(1):53.
58. Anuradha B, Bai YD, Sailaja S, Sudhakar J, Priyanka M, Deepika V. Evaluation of anti-inflammatory effects of curcumin gel as an adjunct to scaling and root planing: a clinical study. *Journal of international oral health: JIOH*. 2015;7(7):90.
59. Arunachalam LT, Sudhakar U, Vasanth J, Khumukchum S, Selvam VV. Comparison of anti-plaque and anti-gingivitis effect of curcumin and chlorhexidine mouth rinse in the treatment of gingivitis: A clinical and biochemical study. *Journal of Indian Society of Periodontology*. 2017;21(6):478.

60. Bhatia M, Urolagin SS, Pentyala KB, Urolagin SB, KB M, Bhoi S. Novel therapeutic approach for the treatment of periodontitis by curcumin. *Journal of clinical and diagnostic research: JCDR*. 2014;8(12):ZC65.
61. Chatterjee A, Debnath K, Rao NKH. A comparative evaluation of the efficacy of curcumin and chlorhexidine mouthrinses on clinical inflammatory parameters of gingivitis: A double-blinded randomized controlled clinical study. *Journal of Indian Society of Periodontology*. 2017;21(2):132.
62. Gottumukkala SN, Koneru S, Mannem S, Mandalapu N. Effectiveness of sub gingival irrigation of an indigenous 1% curcumin solution on clinical and microbiological parameters in chronic periodontitis patients: A pilot randomized clinical trial. *Contemporary clinical dentistry*. 2013;4(2):186.
63. Gottumukkala SN, Sudarshan S, Mantena SR. Comparative evaluation of the efficacy of two controlled release devices: Chlorhexidine chips and indigenous curcumin based collagen as local drug delivery systems. *Contemporary clinical dentistry*. 2014;5(2):175.
64. Guru SR, Kothiwale SV, Saroch N, Guru RC. Comparative evaluation of inhibitory effect of curcumin and doxycycline on matrix metalloproteinase-9 activity in chronic periodontitis. *Indian Journal of Dental Research*. 2017;28(5):560.
65. Hugar SS, Patil S, Metgud R, Nanjwade B, Hugar SM. Influence of application of chlorhexidine gel and curcumin gel as an adjunct to scaling and root planing: A interventional study. *Journal of natural science, biology, and medicine*. 2016;7(2):149.
66. Muglikar S, Patil KC, Shivswami S, Hegde R. Efficacy of curcumin in the treatment of chronic gingivitis: a pilot study. *Oral health & preventive dentistry*. 2013;11(1).
67. Nagasri M, Madhulatha M, Musalaiah S, Kumar PA, Krishna CM, Kumar PM. Efficacy of curcumin as an adjunct to scaling and root planning in chronic periodontitis patients: A clinical and microbiological study. *Journal of pharmacy & bioallied sciences*. 2015;7(Suppl 2):S554.
68. Nasra MM, Khiri HM, Hazzah HA, Abdallah OY. Formulation, in-vitro characterization and clinical evaluation of curcumin in-situ gel for treatment of periodontitis. *Drug delivery*. 2017;24(1):133-42.
69. Pulikkotil S, Nath S. Effects of curcumin on crevicular levels of IL - 1 β and CCL 28 in experimental gingivitis. *Australian dental journal*. 2015;60(3):317-27.

70. Ravishankar P, Kumar YP, Anila E, Chakraborty P, Malakar M, Mahalakshmi R. Effect of local application of curcumin and ornidazole gel in chronic periodontitis patients. *International journal of pharmaceutical investigation*. 2017;7(4):188.
71. Cochran DL. Inflammation and bone loss in periodontal disease. *Journal of periodontology*. 2008;79(8 Suppl):1569-76.
72. Suda T, Takahashi N, Udagawa N, Jimi E, Gillespie MT, Martin TJ. Modulation of osteoclast differentiation and function by the new members of the tumor necrosis factor receptor and ligand families. *Endocrine reviews*. 1999;20(3):345-57.
73. Kirkwood KL, Cirelli JA, Rogers JE, Giannobile WV. Novel host response therapeutic approaches to treat periodontal diseases. *Periodontology 2000*. 2007;43(1):294-315.
74. Crotti T, Smith MD, Hirsch R, Soukoulis S, Weedon H, Capone M, et al. Receptor activator NF κ B ligand (RANKL) and osteoprotegerin (OPG) protein expression in periodontitis. *Journal of periodontal research*. 2003;38(4):380-7.
75. Liu YCG, Lerner UH, Teng YTA. Cytokine responses against periodontal infection: protective and destructive roles. *Periodontology 2000*. 2010;52(1):163-206.
76. Kawai T, Matsuyama T, Hosokawa Y, Makihiro S, Seki M, Karimbux NY, et al. B and T lymphocytes are the primary sources of RANKL in the bone resorptive lesion of periodontal disease. *The American journal of pathology*. 2006;169(3):987-98.
77. DiDonato JA, Hayakawa M, Rothwarf DM, Zandi E, Karin M. A cytokine-responsive I κ B kinase that activates the transcription factor NF- κ B. *Nature*. 1997;388(6642):548.
78. Milward M, Chapple I, Wright H, Millard J, Matthews J, Cooper P. Differential activation of NF- κ B and gene expression in oral epithelial cells by periodontal pathogens. *Clinical & Experimental Immunology*. 2007;148(2):307-24.
79. Carayol N, Chen J, Yang F, Jin T, Jin L, Wang C-Y. A dominant function of IKK/NF- κ B signaling in global lipopolysaccharide-induced gene expression. *Journal of Biological Chemistry*. 2006;281(41):31142-51.
80. Ambili R, Janam P. A critique on nuclear factor-kappa B and signal transducer and activator of transcription 3: The key transcription factors in

periodontal pathogenesis. *Journal of Indian Society of Periodontology*. 2017;21(5):350.

81. Lai J-l, Liu Y-h, Liu C, Qi M-p, Liu R-n, Zhu X-f, et al. Indirubin inhibits LPS-induced inflammation via TLR4 abrogation mediated by the NF-kB and MAPK signaling pathways. *Inflammation*. 2017;40(1):1-12.

82. Nussbaum G, Ben-Adi S, Genzler T, Sela M, Rosen G. Involvement of Toll-like receptors 2 and 4 in the innate immune response to *Treponema denticola* and its outer sheath components. *Infection and immunity*. 2009;77(9):3939-47.

83. Verdrengh M, Bokarewa M, Ohlsson C, Stolina M, Tarkowski A. RANKL-targeted therapy inhibits bone resorption in experimental *Staphylococcus aureus*-induced arthritis. *Bone*. 2010;46(3):752-8.

84. Abu-Amer Y. NF-kappaB signaling and bone resorption. *Osteoporosis international : a journal established as result of cooperation between the European Foundation for Osteoporosis and the National Osteoporosis Foundation of the USA*. 2013;24(9):2377-86.

85. Shapira L, Borinski R, Sela MN, Soskolne A. Superoxide formation and chemiluminescence of peripheral polymorphonuclear leukocytes in rapidly progressive periodontitis patients. *Journal of Clinical Periodontology*. 1991;18(1):44-8.

86. Marquis R. Oxygen metabolism, oxidative stress and acid-base physiology of dental plaque biofilms. *Journal of industrial microbiology*. 1995;15(3):198-207.

87. Borba TT, Molz P, Schlickmann DS, Santos C, Oliveira CF, Prá D, et al. Periodontitis: Genomic instability implications and associated risk factors. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*. 2019.

88. Chopra A, Sivaraman K. An update on possible pathogenic mechanisms of periodontal pathogens on renal dysfunction. *Critical reviews in microbiology*. 2019:1-25.

89. Torres MA, Jones JD, Dangl JL. Reactive oxygen species signaling in response to pathogens. *Plant physiology*. 2006;141(2):373-8.

90. Roos D, van Bruggen R, Meischl C. Oxidative killing of microbes by neutrophils. *Microbes and infection*. 2003;5(14):1307-15.

91. Tamaki N, Hayashida H, Fukui M, Kitamura M, Kawasaki K, Nakazato M, et al. Oxidative stress and antibody levels to periodontal bacteria in adults: the Nagasaki Islands study. *Oral diseases*. 2014;20(3).

92. Baltacıoğlu E, Yuva P, Aydın G, Alver A, Kahraman C, Karabulut E, et al. Lipid peroxidation levels and total oxidant/antioxidant status in serum and saliva from patients with chronic and aggressive periodontitis. Oxidative stress index: a new biomarker for periodontal disease? *Journal of periodontology*. 2014;85(10):1432-41.
93. Dahiya P, Kamal R, Gupta R, Bhardwaj R, Chaudhary K, Kaur S. Reactive oxygen species in periodontitis. *Journal of Indian Society of Periodontology*. 2013;17(4):411.
94. Gan P, Gao Z, Zhao X, Qi G. Surfactin inducing mitochondria-dependent ROS to activate MAPKs, NF- κ B and inflammasomes in macrophages for adjuvant activity. *Scientific reports*. 2016;6:39303.
95. Sho T, Xu J. Role and mechanism of ROS scavengers in alleviating NLRP3-mediated inflammation. *Biotechnology and applied biochemistry*. 2019;66(1):4-13.
96. Liu W, Gu J, Qi J, Zeng XN, Ji J, Chen ZZ, et al. Lentinan exerts synergistic apoptotic effects with paclitaxel in A549 cells via activating ROS-TXNIP-NLRP3 inflammasome. *Journal of cellular and molecular medicine*. 2015;19(8):1949-55.
97. Kanzaki H, Shinohara F, Kajiya M, Kodama T. The Keap1/Nrf2 protein axis plays a role in osteoclast differentiation by regulating intracellular reactive oxygen species signaling. *Journal of Biological Chemistry*. 2013;288(32):23009-20.
98. Lubbad A, Oriowo M, Khan I. Curcumin attenuates inflammation through inhibition of TLR-4 receptor in experimental colitis. *Molecular and cellular biochemistry*. 2009;322(1-2):127-35.
99. Kim G-Y, Kim K-H, Lee S-H, Yoon M-S, Lee H-J, Moon D-O, et al. Curcumin inhibits immunostimulatory function of dendritic cells: MAPKs and translocation of NF- κ B as potential targets. *The Journal of Immunology*. 2005;174(12):8116-24.
100. Mun SH, Kim HS, Kim JW, Ko NY, Kim DK, Lee BY, et al. Oral administration of curcumin suppresses production of matrix metalloproteinase (MMP)-1 and MMP-3 to ameliorate collagen-induced arthritis: inhibition of the PKC δ /JNK/c-Jun pathway. *Journal of pharmacological sciences*. 2009;111(1):13-21.
101. Boyanapalli SS, Huang Y, Su Z, Cheng D, Zhang C, Guo Y, et al. Pharmacokinetics and Pharmacodynamics of Curcumin in regulating

- anti-inflammatory and epigenetic gene expression. *Biopharmaceutics & drug disposition*. 2018;39(6):289-97.
102. Chandrasekaran CV, Kannan Sundarajan JRE, Gururaja GM, Mundkinajeddu D, Agarwal A. Immune-stimulatory and anti-inflammatory activities of *Curcuma longa* extract and its polysaccharide fraction. *Pharmacognosy Research*. 2013;5(2):71.
103. Sajithlal G, Chithra P, Chandrakasan G. Effect of curcumin on the advanced glycation and cross-linking of collagen in diabetic rats. *Biochemical pharmacology*. 1998;56(12):1607-14.
104. Sidhu GS, Singh AK, Thaloor D, Banaudha KK, Patnaik GK, Srimal RC, et al. Enhancement of wound healing by curcumin in animals. *Wound Repair and Regeneration*. 1998;6(2):167-77.
105. Anand P, Kunnumakkara AB, Newman RA, Aggarwal BB. Bioavailability of curcumin: problems and promises. *Molecular pharmaceutics*. 2007;4(6):807-18.
106. Tønnesen HH. Solubility, chemical and photochemical stability of curcumin in surfactant solutions. *Studies of curcumin and curcuminoids, XXVIII. Die Pharmazie*. 2002;57(12):820-4.
107. Tønnesen HH, Másson M, Loftsson T. Studies of curcumin and curcuminoids. XXVII. Cyclodextrin complexation: solubility, chemical and photochemical stability. *International Journal of Pharmaceutics*. 2002;244(1-2):127-35.
108. Tomren M, Masson M, Loftsson T, Tønnesen HH. Studies on curcumin and curcuminoids: XXXI. Symmetric and asymmetric curcuminoids: stability, activity and complexation with cyclodextrin. *International journal of pharmaceutics*. 2007;338(1-2):27-34.
109. Prasad S, Tyagi AK, Aggarwal BB. Recent developments in delivery, bioavailability, absorption and metabolism of curcumin: the golden pigment from golden spice. *Cancer research and treatment: official journal of Korean Cancer Association*. 2014;46(1):2.
110. Zhang Y, M Golub L, Johnson F, Wishnia A. pKa, zinc-and serum albumin-binding of curcumin and two novel biologically-active chemically-modified curcumins. *Current medicinal chemistry*. 2012;19(25):4367-75.
111. Zhang Y, Gu Y, Lee H-M, Hambardjjeva E, Vranková K, M Golub L, et al. Design, synthesis and biological activity of new polyenolic inhibitors of matrix

metalloproteinases: a focus on chemically-modified curcumins. *Current medicinal chemistry*. 2012;19(25):4348-58.

112. Khalil NM, do Nascimento TCF, Casa DM, Dalmolin LF, de Mattos AC, Hoss I, et al. Pharmacokinetics of curcumin-loaded PLGA and PLGA–PEG blend nanoparticles after oral administration in rats. *Colloids and Surfaces B: Biointerfaces*. 2013;101:353-60.

113. Paka GD, Ramassamy C. Optimization of curcumin-loaded PEG-PLGA nanoparticles by GSH functionalization: investigation of the internalization pathway in neuronal cells. *Molecular pharmaceutics*. 2016;14(1):93-106.

114. Garala K, Joshi P, Shah M, Ramkishan A, Patel J. Formulation and evaluation of periodontal in situ gel. *International journal of pharmaceutical investigation*. 2013;3(1):29.

115. Sakima V, Barbugli P, Cerri P, Chorilli M, Carmello J, Pavarina A, et al. Antimicrobial photodynamic therapy mediated by curcumin-loaded polymeric nanoparticles in a murine model of oral candidiasis. *Molecules*. 2018;23(8):2075.

Figure legend

Figure 1. Pathogenesis of periodontitis.

Tables

Table 1. The effectiveness of curcumin for the management of gingivitis – clinical studies.

Table 2. Experimental model's studies showing the impact of curcumin on periodontitis.